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1 **Gene Expression Profiling During Grape Leaf Development and**
2 **Senescence by High Density Filters**

3
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11 **Keywords:** *Vitis vinifera*, transcription profiling, macroarray, leaf senescence,
12 photosynthesis, proteinases

13
14 **Abstract**

15 **Leaf photosynthesis in grapevines peaks approximately 30 to 40 days after**
16 **unfolding and declines thereafter. In order to achieve high quality fruit is important**
17 **that during the ripening stage the vine's photosynthetic capacity is not diminished**
18 **due to the leaf senescence. To study grapevine leaf senescence at the molecular level,**
19 **a transcription profiling analysis during leaf development and senescence was**
20 **undertaken. A cDNA macroarray containing about 2300 putative unigenes was**
21 **constructed and hybridized with RNA isolated from *Vitis vinifera* 'Pinot noir' leaves**
22 **7, 88 and 131 days-old. Data analysis focused on genes involved in the photosynthetic**
23 **process or belonging to the proteinase family. At this stage of leaf senescence two**
24 **gene categories were mainly affected: the chlorophyll a/b binding (CAB) genes and**
25 **the cysteine-proteinases genes. While CAB transcripts decrease in their abundance,**
26 **cysteine-proteinases are activated.**

27
28 **INTRODUCTION**

29 Leaf development and senescence are biological processes under tight genetic
30 control and characterized by an increase in the photosynthetic rates during leaf expansion
31 and a decline during leaf senescence (Bertamini and Nedunchezian, 2002). To obtain
32 high quality fruit it is important that during the ripening stage the vine's canopy does not
33 lose photosynthetic capacity. Vineyard management practices such as pruning or
34 fertilization can slow down leaf aging but often this will not entirely solve the problem.

35 High-throughput sequencing of expressed sequence tags (ESTs) and gene
36 expression analysis, on a genome-wide scale, have opened new possibilities to shed light
37 on complex biological processes such as leaf senescence (Andersson et al., 2004;
38 Bhalerao et al., 2003). Understanding the molecular biology governing the physiology of
39 such a process may help in maintaining the efficiency of the plant for a greater duration of
40 the season.

41 During the last three years we set up and characterized 6 cDNA libraries from
42 different grape organs and collected a large number of ESTs which are stored in a
43 publicly available database (<http://www.ismaa.it/>) and deposited also in GeneBank
44 (<http://www.ncbi.nlm.nih.gov/dbEST/index.html>). A subset of the cDNA clones from the
45 leaf, shoot, inflorescence bud and berry cDNA libraries (ca. 2300 unique sequences) have
46 been spotted on high density nylon filters and probed with RNA isolated from grapevine
47 leaves at different developmental stages. In this paper we present the results of the

48 analysis of the expression data for those genes that are involved specifically in
49 photosynthesis and in protein degradation, two processes which are strongly affected
50 during leaf senescence.

51

52 MATERIALS AND METHODS

53 The 8th leaf of *Vitis vinifera* L. 'Pinot noir' from 10-year-old vines grafted to
54 '3309C' grown under field conditions with upright growing shoots in San Michele
55 all'Adige (2002) was collected at 3 developmental stages: expanding (7-days-old), mature
56 (88-days-old) and senescent (131-days-old).

57 The 4010 ESTs clones derived from 'Pinot noir' leaf, bud, and berry and 'Regent'
58 shoot, inflorescence cDNA libraries were PCR amplified and double spotted on high
59 density nylon filters (7.3 cm x 11.5 cm) in a 5 x 5 pattern. The clustering of these ESTs
60 had previously revealed that they correspond to 2320 unique sequences. Total RNA was
61 isolated from the leaf samples according to Moser et al., 2004. For the labelling of each
62 cDNA probe 25 µg of total RNA and [³²P]-α-dCTP were used in a reverse transcription
63 reaction. Filters hybridization was carried out at 65°C with a standard procedure
64 (Sambrook et al., 1989). Twelve replicates (including the double spots) of each data point
65 were obtained for each leaf developmental stage.

66 Hybridization signals were quantified by the programme Arrayvision (Imaging
67 Research Inc.) and corrected for local background. Clones for which 90% of the
68 hybridization signals were below a threshold value (95th percentile of the intensities
69 observed in the empty spots) were discarded from the rest of the analysis. Signal
70 intensities were then normalized by using as spiking control the human nebulin gene, a
71 transcript not present in grapevine and added at a concentration of 0.01% of the
72 poly(A)⁺RNA amount used for each probe synthesis. Target spots of human nebulin were
73 spotted on the filters at growing concentrations (0.01 ng µL⁻¹, 0.10 ng µL⁻¹, 10 ng µL⁻¹ and
74 100 ng µL⁻¹) and a linear regression was performed on nebulin data (log of median
75 expression of the array versus log of median expression of the overall nebulin data) for
76 each array. Intercept and regression coefficient of each filter were then applied to the
77 whole data set of the same filter. The result of the normalization process was validated by
78 comparison with the transcript concentration of four genes determined by quantitative real
79 time PCR. As gene expression value for each clone was taken the median value of the 12
80 data replicates.

81

82 RESULTS AND DISCUSSION

83 A high density cDNA array was constructed by double spotting 4010 PCR
84 amplified clones derived from 5 different grape organs, namely, leaf, berry, bud, shoot
85 and inflorescence. About 80% of these clones were previously subjected to single-pass 5'
86 sequencing as part of our EST sequencing project and have been clustered into 2320
87 contigs (ca. 40% redundancy). We were able to annotate about 75% of these tentative
88 unigenes by sequence comparison against the non-redundant protein NCBI database using
89 the programme PSI-BLAST. Although the amplified cDNA on the macroarray were
90 corresponding only in part to leaf sequences, our hybridization results revealed that the
91 high majority of the clones gave a hybridization signal.

92 Since we were mainly interested in understanding the molecular biology
93 underlying the severe morphological and physiological changes that occur going from a
94 young leaf to an old one with respect to the mature stage, we decided to primarily focus
95 our analysis on selected categories of genes expressed at the mature stage as a reference.

96 The genes chosen to be investigated in this study were those related to the photosynthesis
97 and those belonging to the proteinases family since these two functional categories are
98 known to be dramatically affected by the senescence process (Andersson et al., 2004;
99 Gepstein et al., 2003). Although our macroarray does not include all the genes involved in
100 these processes, our data are still highly valuable for two reasons: the paucity of studies
101 on leaf ontogeny and senescence of perennial species and the often strictly correlated
102 expression profiles shown by genes belonging to the same metabolism.

103 The process of leaf senescence involves a shift in a leaf's function: its contribution
104 to carbohydrate production in the plant diminishes because of a decline in its
105 photosynthetic activity and the remobilisation of nitrogen, phosphorus and metals to
106 younger leaves or to other compartments (Quirino et al., 2000). In our study (Fig. 1) we
107 found two different behaviours in the transcripts level of CAB genes on one hand and the
108 Rubisco small subunit and Photosystem II (PSII) and I (PSI) genes on the other hand.
109 Going from the young to the senescent stages the average expression of the CAB genes
110 shows a large decrease becoming half its initial value whereas the constituents of the
111 PSII, PSI and the Rubisco small subunit show little changes. This result would indicate
112 that the CAB transcripts have a peak concentration in the young leaf followed by a
113 constant decrease thereafter and that the other components of the photosynthetic
114 apparatus considered in this study were not yet affected by the onset of senescence. These
115 data are in good agreement with the physiological data collected on the same samples: the
116 activity of electron transport through the whole chain ($H_2O > MV$) measured on isolated
117 thylakoid membranes declined only 15% from the mature to the senescent leaves (data
118 not shown). As depicted in Fig. 1 a decrease in gene expression of the CAB genes and the
119 Rubisco small subunit, much more noticeable for the CAB genes, was observed between
120 the young stage and the mature one. A partial (CAB genes) or full recovery (Rubisco) in
121 the transcripts concentrations occurred then going from the mature to the senescent stage.
122 This large decrease in expression could be at least partially explained by a strong climate
123 change (thunderstorm with hail) that occurred a few days prior to sampling the leaves for
124 the mature stage analysis. Apparently this stress highly affected CAB and Rubisco
125 transcripts accumulation but only minimally affected the expression of the other
126 photosynthetic components.

127 Proteolytic degradation takes place in all senescing plant organs in order to
128 efficiently degrade cell components and re-allocate the nutrients to other organs.
129 Accordingly an increase in the mRNA levels of specific proteases has been observed
130 during senescence both in the model plant *Arabidopsis* (Gepstein et al., 2003) and in
131 autumn aspen leaves (Bhalerao et al., 2003). In all studied senescing systems cellular
132 proteolysis is carried out by similar mechanisms including aspartic- cysteine-proteinases
133 and the ubiquitin degradation pathway. In our filters were included cDNA clones
134 belonging to the cysteine-, subtilisin -, metallo- and ubiquitin specific proteinases. Three
135 classes of proteinase transcripts out of four appeared to accumulate in the leaves from 88
136 to 131 days of age, even though not at the same rate (Fig. 2). The cysteine class of
137 proteinases increased 150% with respect to the subtilisin and ubiquitin specific proteinase
138 classes. The gene for a metalloproteinase showed instead a large decrease in its
139 abundance from the mature to the senescent stage. Our results are in agreement with
140 others that have identified specific cysteine-proteinases among the senescence-associated
141 genes (SAGs), genes that are upregulated during senescence (Chandlee, 2001). Moreover,
142 as previously reported for aspen leaves during autumn (Bhalerao et al., 2003) no major
143 activation of the ubiquitin specific proteinase was observed in grape leaves.

144 In conclusion our ESTs-based macro-array has revealed to be a valuable tool for
145 studying gene expression during grapevine leaf development and particularly in the
146 senescent leaf. Our transcriptional analysis, focused on genes involved in the
147 photosynthetic process or belonging to the family of the proteinases, has demonstrated
148 that at this stage of leaf senescence two gene categories were mainly affected: the CAB
149 genes and the cysteine-proteinases genes. While CAB transcripts decrease in abundance,
150 cysteine-proteinases appear to be activated. It is our intention to extend this type of
151 investigation to older leaves in order to see when a decline in the transcripts level of the
152 other photosynthetic genes occurs.

153

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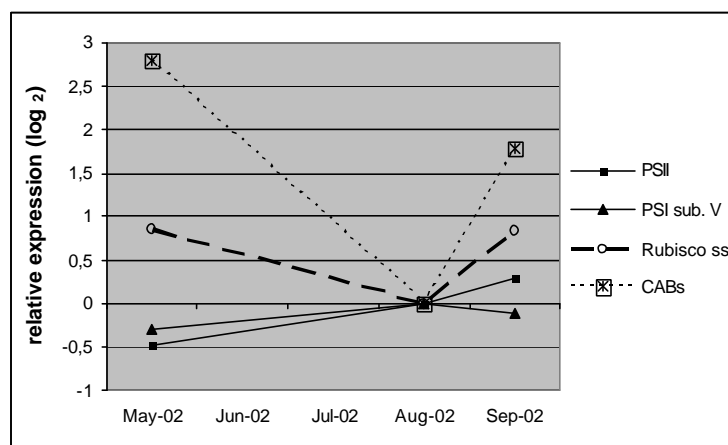
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159 **Literature cited**

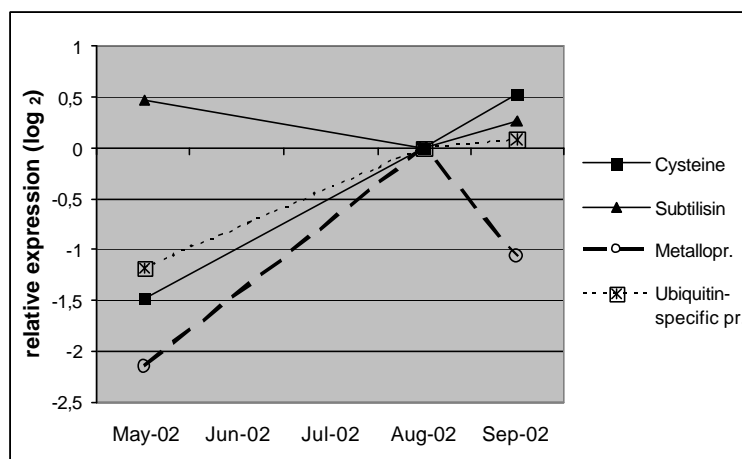
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 188 Fig. 1. Gene expression pattern of photosynthesis-related genes in grape leaves of different ages
 189 (7, 88 and 131 days from unfolding). PSII: psbT (Acc.: CF607382), 10kDa polypeptide
 190 (Acc.: CF609227), psbW (Acc.: CF607350), oxygen evolving enhancer protein 1 (Acc.:
 191 CF609404). PSI: subunit V precursor (Acc.: CF607564). Rubisco small subunit (Acc.:
 192 CF608461). CABs (chlorophyll a/b binding genes): light-harvesting chlorophyll a/b
 193 binding protein (Acc.: 609601 and 606496, 606408, 607287 and 606883, 606963), CAB
 194 binding protein 40 (Acc.: 606286 and 607092 and 606632). The average levels of
 195 expression of the different categories are reported as the logarithm to the base 2 of the
 196 ratio between the expression value at that age and the 88-day-old leaves expression
 197 value to the reference.
 198



199
 200 Fig. 2. Gene expression pattern of several classes of proteinases genes in grape leaves of
 201 different age (7, 88 and 131 days from unfolding). Cysteine: (Acc.: CF606407, 609367,
 202 609241, 609618 and 609506, 610086). Subtilisin: (Acc.: CF609020 and 609479, 609190
 203 and 608895). Metalloproteinase (Acc.: CF607015). Ubiquitin-specific proteinase (Acc.:
 204 CF606767). The average levels of expression of the different categories are reported as
 205 the logarithm to the base 2 of the ratio between the expression value at that age and the
 206 88-day-old leaves expression value as the reference.