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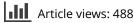
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# The AMPK-FoxO3A axis as a target for cancer treatment

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Key words: AMPK, FoxO3A, cancer, signal-dependent transcription, antitumor therapy, cell metabolism, calorie restriction

FoxO proteins are an evolutionarily conserved subfamily of transcription factors involved in tumor suppression, regulation of energy metabolism and development in several tissues, and are mainly regulated by phosphorylation-dependent nuclear/ cytoplasmic shuttling. The transcriptional activity of FoxO3A, one of the four members of the family, is further modulated by AMPK, one of the key regulators of cellular metabolism, which basically shifts cell machinery from energy-consuming to energy-producing pathways.

We recently demonstrated that the AMPK/FoxO3A energy sensor pathway is still inducible in human cancer cells in response to metabolic stress, as it becomes activated in colorectal and ovarian cancer cells in response to the inhibition of p38 $\alpha$ . Activation of the FoxO3A transcriptional program initially induces autophagy as an attempt to retain energy to survive, whereas under persistent stress conditions it triggers autophagic cell death.

In this review, we focus on the connections between AMPK and FoxO3A, describing their central role as modulators of fundamental processes such as stress resistance, cell metabolism, autophagy and cell death, and highlighting the therapeutic potential of pharmacological modulation of the AMPK-FoxO3A axis.

#### FoxO3A: The "Silent Guardian" against Age-Related Diseases

The *forkhead-box* (Fox) gene family of transcription factors includes more than 100 members, with evolutionarily conserved roles in stress resistance and metabolism, development, differentiation, proliferation and survival. In humans, at least 43 members have been identified and classified in subfamilies recognized by letters (e.g., FoxA).<sup>1</sup>

The FoxO subfamily is composed of FoxO1, FoxO3A, FoxO4, that are ubiquitously expressed, and FoxO6, whose expression is restricted to neural cells. These transcription factors recognize and bind the conserved consensus core recognition motif FHRE (5'TTGTTTAC3') on target gene promoters.<sup>2</sup> As the consensus sequence is common to the various members of the FoxO family, additional mechanisms of regulation are needed to ensure the

\*Correspondence to: Cristiano Simone; Email: simone@negrisud.it Submitted: 12/15/09; Accepted: 12/22/09 Previously published online: www.landesbioscience.com/journals/cc/article/11035 specificity of their transcriptional activity (i.e., multiple interactions with co-regulators, differential expression levels and cell-type and tissue specificity). FoxO proteins are evolutionarily conserved from *C. elegans* to mammals. In invertebrates, the homologous DAF-16 promotes longevity; in mammals, the members of the FoxO family play a role in proliferation/arrest, survival/death, metabolism and autophagy, and have been involved in tumor suppression, regulation of energy metabolism and development in a number of tissues. All these functions are mediated by the specific activation of a coordinated transcriptional program.<sup>3</sup>

The functions of the FoxO proteins have been deeply characterized by employing various animal models.<sup>4</sup> As mentioned above, the first studies were conducted in C. elegans, where DAF-16 has been proven to increase lifespan and regulate nutrient sensing. These functions are conserved in Drosophila, where dFOXO is also involved in insulin signaling. Since in mammals at least three FoxOs are ubiquitously expressed and share several target genes, experiments of individual gene disruption have been necessary to evaluate their physiological roles in development and adult life. FoxO1-null mice display an embryonic lethal phenotype as a consequence of incomplete vascular development.<sup>5</sup> Conversely, FoxO3A- and FoxO4-null mice are viable and grossly indistinguishable from their littermate controls, indicating their dispensability for normal vascular development. However, FoxO3A-null mice show age-dependent infertility due to abnormal ovarian follicular development leading to degeneration,<sup>5,6</sup> and presented severe muscle regeneration impairment.7 Taken together, these experiments indicate that some degree of functional diversification exists between FoxO proteins both in developing and adult animals, without excluding, however, the possibility of functional redundancy. To clarify this issue, all three genes were simultaneously disrupted.<sup>8,9</sup> The conditional triple knock-out developed thymic lymphomas and haemangiomas, suggesting that FoxO proteins may play redundant functions at least in suppressing tumorigenesis, but they are characterized by cell-type and tissue specificity.

In multicellular organisms, all the above mentioned functions carried out by FoxO proteins result in the regulation of the response to oxidative stress, starvation and calorie restriction with the ultimate effect of increasing lifespan and counteracting age-related diseases, such as diabetes and cancer.<sup>10</sup> It is important to highlight that under pathological conditions FoxO activation often results in a phenotypically different outcome compared to what happens in normal conditions. The role of FoxO1 in

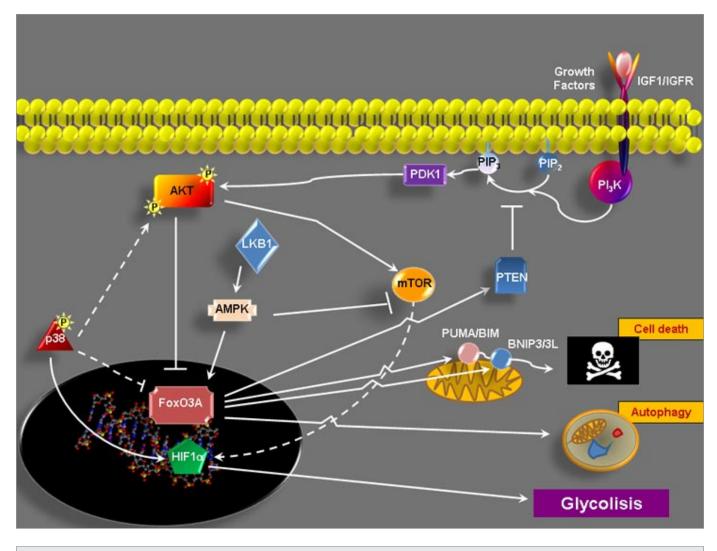


Figure 1. Signaling cascades regulating the balance between energy-consuming and energy-producing pathways.

diabetes,<sup>11</sup> and that of FoxO3A in cancer<sup>12-15</sup> are just two examples. Indeed, albeit gene disruption experiments in mice did not highlight a role for FoxO3A in tumor development, and despite the fact that triple FoxO knockout mice developed only thymic lymphomas and haemangiomas, but no other tumors, FoxO3A is required for colorectal cancer cell death induced by cisplatinum<sup>15</sup> and p38 $\alpha$  inhibitors.<sup>12</sup>

FoxO3A is able to bind to promoters and induce transcription of target genes involved in cell cycle arrest (p21, p27, *CYCLIN G2*, *RBL2*, *BCL6*), cell death (*GADD45*, *BIM*, *FasL*, *PUMA*, *PTEN*, *BNIP3*, *BNIP3L*), cell metabolism (*PGCIα*, *PEPCK*, *UCP2*) and stress resistance (*SOD2*, *Catalase*).<sup>12,13,15-17</sup> Besides, some other genes, like those coding for cyclin D, E and A, are reported to be negatively regulated by FoxOs in a direct or indirect manner.<sup>16,18,19</sup> Moreover, FoxO3A regulates several autophagy-related genes such as *MAP1LC3*, *Gabarapl1*, *ATG12*, *BNIP3* and *BNIP3L*—in muscle cells in response to atrophic signals and in cancer cells in response to the downregulation of the glycolytic cascade promoted by inhibition of the p38α-HIF1α pathway.<sup>12,13,15,20,21</sup> The involvement of FoxO in the regulation of autophagy seems to be evolutionarily conserved, as indicated by the role of dFOXO in Drosophila.  $^{\rm 22}$ 

The above mentioned functions of FoxO3A indicate that it could have an important role in the control of tumor growth. Indeed, FoxO3A overexpression has been shown to inhibit tumor growth in vitro and tumor size in vivo in breast cancer cells.<sup>23,24</sup> Furthermore, cytoplasmic localization of FoxO3A seems to correlate with poor survival in patients with breast cancer.<sup>23</sup> The significance of FoxO3A antitumoral activity is also underscored by studies conducted in leukemia,<sup>25</sup> prostate cancer<sup>26-28</sup> and glioblastoma.<sup>29</sup>

The emerging picture suggests that FoxO3A does not act as a crucial gatekeeper for the well being of the cell, rather it seems to operate as a 'silent guardian', which is called into action when things get worse. In fact, it is not devoted to the monitoring and preservation of the health state of the cells in non-stressed conditions. Conversely, it seems to be required following exogenous stress stimuli (i.e., oxidative or metabolic stress conditions) to help restoring the metabolic homeostasis of the cell. To our knowledge, functional or structural modifications favoring or contributing to the onset of diseases have not been reported to date in FoxO3A locus, except for a chromosomal translocation, t(6; 11), found in a few cases of acute myeloid leukemia, which involves a fusion between MLL and FoxO3A.<sup>25</sup> On the other hand, FoxO3A appears to be preferentially inactivated by deregulated upstream signaling pathways, suggesting that its inactivation might be beneficial to the development of pathological conditions. As a consequence, FoxO3A is still functional and might be a promising target to be activated in order to counteract age-related diseases such as cancer.

#### Kinase-Dependent Regulation of FoxO3A: The Role of AMPK

FoxO proteins are regulated by phosphorylation-dependent nuclear/cytoplasmic shuttling as a result of the activity of the Akt and JNK kinases, which is evolutionarily conserved from invertebrates to humans.<sup>3</sup> Akt directly phosphorylates FoxOs (at the T32, S253 and S315 residues in the case of FoxO3A) and targets them to bind to the 14-3-3 nuclear export protein. This binding excludes FoxO factors from the nucleus, leading to their cytoplasmic accumulation and subsequent degradation. In addition, other kinases involved in oncogenic signals, such as SGK, CK1, DYRK1A and IKK $\beta$ , can inactivate FoxOs with a similar mechanism. On the other hand, JNK1 phosphorylates the 14-3-3 protein inhibiting its binding to FoxOs and promoting their nuclear localization. The observation that stress stimuli trigger the nuclear localization of the FoxO proteins even in the presence of growth factors illustrates the hierarchy of the regulation of these transcription factors, with phosphorylation by JNK (at Thr447 and Thr451 in the case of FoxO4) overriding the effect of phosphorylation by Akt.

As a further mechanism of regulation, it has been recently shown that the activation of the energy sensor pathway, which is triggered by an increased AMP/ATP ratio, leads to the AMPK-mediated phosphorylation of FoxO3A.30 AMPK, the AMP-responsive kinase, is a central metabolic switch found in all eukaryotes that governs glucose and lipid metabolism in response to alterations in nutrients and intracellular energy levels. Importantly, AMPK has been identified as a direct downstream effector of the LKB1 tumor suppressor kinase in a mechanism representing a direct connection between energy metabolism and cell growth control.<sup>31</sup> Indeed, the LKB1/AMPK pathway is clearly involved in the regulation of cell growth of gastrointestinal hamartomas and many other malignancies including those arising in colon, breast, ovarian, pancreatic and lung tissues (the so called Peutz-Jeghers Syndrome, PJS), as well as in melanoma, breast and prostate cancer.31-35

AMPK is able to associate with and phosphorylate FoxO both in nematodes and mammalian cells. In nematodes, AMPK phosphorylates FoxO on six residues: T166, S202, S314, S321, T463 and S466. Also mammalian FoxO3A can be phosphorylated in vitro by AMPK on six sites (T179, S399, S413, S555, S588, S626), five of which are located to the transactivation domain.<sup>30</sup> Mutation of the six residues strongly reduces (84%), but not completely abrogates FoxO3A phosphorylation, suggesting the existence of other putative target sites. FoxO3A is phosphorylated by 2-deoxyglucose-activated AMPK in mammalian cells. The FoxO3A mutant in which all six residues found to be phosphorylated by AMPK were replaced by alanine was severely impaired in transcription, but maintained the ability to bind its cognate sequences, at least in vitro, and to perform nucleo-cytoplasmic shuttling depending on external cues. Even if these data strongly suggest that AMPK plays a role in FoxO3A dependent transcription, they do not exclude the existence of other signaling pathways converging on the same residues to modulate FoxO3A transcriptional activity, nor do they prove a direct role of AMPKmediated FoxO3A phosphorylation in transcriptional modulation. It was recently nicely demonstrated that AMPK is able to modulate NAD<sup>+</sup> metabolism and SIRT1 deacetilase activity, a well known FoxO coactivator.<sup>36-38</sup> In particular, AMPK activation results in increased NAD+ levels that lead to SIRT1 activation and deacetylation-induced activation of its downstream targets, including PGC1α, FoxO1 and FoxO3A. Taken together, these studies depict an intricate scenario, in which AMPK is able to indirectly modulate FoxO3A transcriptional activity by activating SIRT1 under metabolic stress. Besides, AMPK is also required for FoxO3A-dependent transcription of thioredoxin during oxidative stress response.<sup>39</sup> Moreover, recent evidences indicate the existence of an alternative pathway of AMPK activation, triggered by mitochondrial ROS and involving FoxO3A and its target genes (SOD, catalase, PGC1 $\alpha$ ).<sup>40</sup>

Yet, several questions remain unanswered. Although it is largely accepted that activated AMPK is able to directly phosphorylate FoxO3A, the physiological or patho-physiological stimuli triggering AMPK-dependent FoxO3A phosphorylation still have to be identified and, most importantly, it is not clear yet which FoxO3A functions are actually regulated by these multiple phosphorylations.

#### The AMPK-FoxO Axis: Balancing Nutrient Sensing and Healthspan/Lifespan

The effect of food restriction on lifespan was first described in rats in 1935 by McCay and colleagues.<sup>41</sup> Several studies have extended their results to other species from nematodes to mice.<sup>42</sup> Dietary restriction, without malnutrition, extends lifespan and reduces age-dependent diseases including, but not restricted to, cancer and atherosclerosis.43-45 It has to be noted that different dietary regimens seem to activate different molecular pathways, probably depending on specific limiting nutrients, and that various tissues may be involved as signaling emitting centers or signaling responding centres.<sup>46-50</sup> Several pathways, including insulin/ IGF1 and AMPK signaling, were shown to have a role in dietary restriction-dependent lifespan extension. Reduced activity of the insulin/IGF1 pathway was sufficient but not strictly required to extend lifespan in a variety of organisms including worms, flies and mice.<sup>51</sup> Animal experiments indicated that the inverse relationship existing between these two pathways (inhibition of PI3K/ Akt and activation of AMPK) represents an essential intracellular switch for transducing the inhibitory effect of dietary energy restriction on mammary carcinogenesis.52 Among the several

molecular pathways analyzed, activation of the AMPK-FoxO axis was shown to be sufficient and required for the dietary restriction effects to occur on both healthspan and lifespan.<sup>53</sup> In particular, Greer and colleagues have shown that dietary restriction-dependent lifespan extension was suppressed in AMPK defective nematodes and significantly reduced in FoxO mutant strains. On the contrary, the long lifespan of insulin receptor mutant nematodes was further enhanced by dietary restriction. The AMPK-FoxO axis is also required for resistance to oxidative stress, a response that is tightly linked to lifespan extension. Of note, in nematodes AMPK-mediated FoxO-dependent lifespan extension under dietary restriction is not reliant on a transcriptional role of FoxO, as this protein fails to accumulate into the nuclei and maintains a cytoplasmic localization. In contrast, the resistance to oxidative stress mediated by a constitutively active AMPK subunit requires the FoxO-dependent transcription of sod-3. These observations suggest a role for FoxO in lifespan extension in at least two different pathways, which are both dependent on AMPK activity.

### The AMPK-FoxO3A Axis in Cancer

We recently reported that the conserved AMPK/FoxO3A energy sensor pathway is still inducible in human cancer cells in response to metabolic stress. Indeed, colorectal and ovarian cancer cells can activate a FoxO3A-dependent transcriptional program in response to decreased glycolysis caused by inhibition of the  $p38\alpha/HIF1\alpha$  pathway, first leading to autophagy and cell cycle arrest as an attempt to retain energy and increase ATP levels, but then triggering autophagic cell death in conditions of persistent stress.<sup>12,13,15</sup> In these cells, FoxO3A induced the transcription of genes whose protein products promote the overall autophagic flux (ATG6, ATG7, ATG12, GABARAP, GABARAPL2), with MAP1LC3 and Gabarapl1 localizing to large vacuolar structures characterized as autophagolysosomes.<sup>12,13,15</sup> The upregulation of FoxO3A target genes involved in metabolism, such as  $PGC1\alpha$ , PEPCK and UCP2, suggests that, in response to energy depletion triggered by p38 $\alpha$  blockade, cancer cells undergo a gene expression reprogramming leading to the expression of enzymes responsible for the conversion of metabolites produced by the autophagic machinery-mainly fatty acids and aminoacidsinto energy fuel to survive. Indeed, colorectal cancer cells also expressed CPT-1 and MCAD, two enzymes involved in fatty acid catabolism.<sup>12</sup> Following p38a inhibition and in response to the acute energy demand that triggered autophagy, cells exited the cell cycle and accumulated in the G<sub>1</sub> phase, probably as an attempt to retain energy for survival. This was achieved by the FoxO3A-mediated upregulation of the cyclin D transcriptional repressor Bcl-6, of the cdk inhibitors p21 and p27, and of the retinoblastoma family protein p130, finally leading to inhibition of the G<sub>1</sub>/S transition. Prolonged inactivation of p38 $\alpha$  by its specific inhibitor SB202190 promoted an increase in the expression levels of FoxO target genes encoding for the BH3-only proteins PUMA, Bim, BNIP3 and BNIP3L, which correlated to the induction of autophagic cell death.<sup>12,13,15</sup> Interestingly, also PTEN, a phosphatase that inhibits the progression of the PI<sub>3</sub>K/Akt signaling, was found to be upregulated, with FoxO3A

being localized to its promoter together with activated PolII, thus suggesting the existence of a negative feedback loop that possibly accounts for the time-dependent reduction of Akt phospho-activation and kinase activity observed in SB202190treated colorectal and ovarian cancer cells.<sup>12,15</sup> The analysis of other pathways involved in the regulation of FoxO function revealed that JNK-a MAPK that is often turned on as an alternative cascade to p38-was activated together with AMPK in cancer cells.<sup>12,15</sup> The AMPK pathway, which becomes activated when the levels of AMP overcome those of ATP, was required for FoxO3A nuclear accumulation and for the subsequent transcriptional activation of target genes involved in autophagy, metabolism, cell cycle arrest and cell death. At the cellular level, AMPK was necessary for the induction of the autophagic phenotypes observed in SB202190-treated colorectal cancer cells.12 These data were obtained by using pharmacological and genetic approaches to knock-down AMPK in cancer cells and suggest that AMPK plays an important role-whether direct or indirect-in the regulation of FoxO3A intracellular localization. Moreover, our preliminary results showed a significant decrease in the overall amount of FoxO3A in cells in which AMPK was inhibited or depleted. Our results also evidenced that the nuclear pool of endogenous FoxO3A was both enriched and hyperphosphorylated at serine and threonine residues in response to p38a inhibition (Chiacchiera and Simone, unpublished results). At present, however, due to the simultaneous perturbation of other pathways affecting the phospho-status of FoxO3A, a direct mechanism involving the phosphorylation of FoxO3A by AMPK can only be hypothesized. As mentioned above, Greer and coworkers<sup>30</sup> described the direct phosphorylation of FoxO3A by AMPK. Moreover, data gathered in experiments performed on 293 cells expressing exogenous wild type FoxO3A or FoxO3A phosphorylation mutants suggested that AMPK-dependent post-translational modifications are responsible for the induction of FoxO3A transcriptional activity, without affecting its cellular localization. Although our observations led to slightly different conclusions, they might not be in contrast, as different experimental procedures, cell type and stimuli were used in the two studies.

In a variety of cells and tissues analyzed, activated AMPK inhibited ACC, which regulates the formation of fatty acids, and induced CPT-1 activity and beta-oxidation with a consequent increase in the ATP/AMP ratio. These findings are in accordance with the activation of the transcriptional program observed following p38 $\alpha$  blockade in colorectal cancer cells. Collectively, our results showed that p38 $\alpha$  blockade causes tumor growth inhibition both in vitro and in vivo by inducing a FoxO3A-dependent transcriptional program. Indeed, SB202190-treated colorectal tumors showed a striking reduction in cell growth, accompanied by nuclear accumulation of FoxO3A and its target genes p21 and PTEN.<sup>12</sup>

These data confirm the potential of antitumoral strategies designed to interfere with the Warburg effect, which accounts for the constitutive activation of aerobic glycolysis commonly occurring in cancer cells. In this broader frame, modulation of the AMPK pathway might prove an effective approach. In fact, several evidences suggest that deregulation of the AMPK pathway could significantly contribute to the Warburg Effect. For example, mTOR has been identified as an indirect link between AMPK and HIF1 $\alpha$ . Indeed, AMPK suppresses mTOR-dependent expression and transcriptional activation of HIF1 $\alpha$  both in vitro and in vivo.<sup>54</sup> According to studies performed in fibroblasts with overactivated mTOR signaling, fibroblasts deficient for LKB1 or AMPK, tumors from PJS patients and LKB1<sup>+/-</sup> mice showed increased levels of HIF1 $\alpha$  and its target genes.<sup>55</sup>

Taken together, these data suggest that AMPK mediators and effectors, when still functional, might be successfully exploited in cancer therapy to counteract tumor-specific metabolism.

#### Perspectives

Understanding the connections between AMPK and FoxO3A is of utmost importance as these proteins play a central role as modulators of fundamental processes such as stress resistance, cell metabolism, autophagy and cell death. As deregulation of these processes is involved in age-related diseases and tumor formation and progression, the ability to modulate the activity of these proteins may represent a promising therapeutic strategy.

The compelling evidences presented in this review highlight that pharmacological manipulation of the AMPK-FoxO3A axis might represent a beneficial approach for cancer treatment. Indeed, FoxO3A is emerging as a key downstream effector of various anticancer therapeutics, such as p38 inhibitors,<sup>12,13,15</sup> cisplatin (a DNA cross-linker),<sup>14</sup> paclitaxel (a microtubule stabilizer),

#### References

- Burgering BM. A brief introduction to FOXOlogy. Oncogene 2008; 27:2258-62.
- Furuyama T, Nakazawa T, Nakano I, Mori N. Identification of the differential distribution patterns of mRNAs and consensus binding sequences for mouse DAF-16 homologues. Biochem J 2000; 349:629-34.
- Calnan DR, Brunet A. The FoxO code. Oncogene 2008; 27:2276-88.
- Arden KC. FOXO animal models reveal a variety of diverse roles for FOXO transcription factors. Oncogene 2008; 27:2345-50.
- Hosaka T, Biggs WH, 3rd, Tieu D, Boyer AD, Varki NM, Cavenee WK, et al. Disruption of forkhead transcription factor (FOXO) family members in mice reveals their functional diversification. Proc Natl Acad Sci USA 2004; 101:2975-80.
- Castrillon DH, Miao L, Kollipara R, Horner JW, DePinho RA. Suppression of ovarian follicle activation in mice by the transcription factor Foxo3a. Science 2003; 301:215-8.
- Hu P, Geles KG, Paik JH, DePinho RA, Tjian R. Codependent activators direct myoblast-specific MyoD transcription. Dev Cell 2008; 15:534-46.
- Paik JH, Kollipara R, Chu G, Ji H, Xiao Y, Ding Z, et al. FoxOs are lineage-restricted redundant tumor suppressors and regulate endothelial cell homeostasis. Cell 2007; 128:309-23.
- Tothova Z, Kollipara R, Huntly BJ, Lee BH, Castrillon DH, Cullen DE, et al. FoxOs are critical mediators of hematopoietic stem cell resistance to physiologic oxidative stress. Cell 2007; 128:325-39.
- van der Horst A, Burgering BM. Stressing the role of FoxO proteins in lifespan and disease. Nat Rev Mol Cell Biol 2007; 8:440-50.

- Nakae J, Biggs WH, 3rd, Kitamura T, Cavenee WK, Wright CV, Arden KC, et al. Regulation of insulin action and pancreatic beta-cell function by mutated alleles of the gene encoding forkhead transcription factor Foxo1. Nat Genet 2002; 32:245-53.
- Chiacchiera F, Matrone A, Ferrari E, Ingravallo G, Lo Sasso G, Murzilli S, et al. p38alpha blockade inhibits colorectal cancer growth in vivo by inducing a switch from HIF1alpha- to FoxO-dependent transcription. Cell Death Differ 2009; 16:1203-14.
- Chiacchiera F, Simone C. Inhibition of p38alpha unveils an AMPK-FoxO3A axis linking autophagy to cancer-specific metabolism. Autophagy 2009; 5:1030-3.
- Fernández de Mattos S, Villalonga P, Clardy J, Lam EW. FOXO3a mediates the cytotoxic effects of cisplatin in colon cancer cells. Mol Cancer Ther 2008; 7:3237-46.
- 15. Matrone A, Grossi V, Chiacchiera F, Fina E, Cappellari M, Loverro G, et al.  $p38\alpha$  is required for ovarian cancer cell metabolism and survival. Int J Gynecol Cancer 2010; in press.
- Ho KK, Myatt SS, Lam EW. Many forks in the path: cycling with FoxO. Oncogene 2008; 27:2300-11.
- You H, Pellegrini M, Tsuchihara K, Yamamoto K, Hacker G, Erlacher M, et al. FOXO3a-dependent regulation of Puma in response to cytokine/growth factor withdrawal. J Exp Med 2006; 203:1657-63.
- Schmidt M, Fernandez de Mattos S, van der Horst A, Klompmaker R, Kops GJ, Lam EW, et al. Cell cycle inhibition by FoxO forkhead transcription factors involves downregulation of cyclin D. Mol Cell Biol 2002; 22:7842-52.
- Delpuech O, Griffiths B, East P, Essafi A, Lam EW, Burgering B, et al. Induction of Mxi1-SRalpha by FOXO3a contributes to repression of Myc-dependent gene expression. Mol Cell Biol 2007; 27:4917-30.

doxorubicin (a DNA intercalating agent), imatinib (a multikinase inhibitor), OSU-03012 (a novel PI3K-Akt inhibitor), trastuzumab and cetuximab (EGFR/HER2 inhibitors), ionizing radiation, and even gene therapy (adenoviral delivery of FoxO3A in melanoma cells).<sup>56</sup> AMPK agonists (i.e., metformin) have a great deal of pharmaceutical interest since they have been in clinics for many years now (for type 2 diabetes, insulin resistance, metabolic syndrome) and are well tolerated by patients. Other agonists include phenformin (currently withdrawn from clinical use because of side effects causing fatal lactic acidosis), AICAR and A769662 (a small molecule directly activating AMPK), which have been shown to inhibit the growth of a plethora of tumor cells in vitro, in tumor xenografts and in animal cancer models in vivo.<sup>31,57,58</sup> Given the great number of patients taking metformin every day, epidemiologic studies have evaluated the effects of this drug on cancer incidence reporting statistical effectiveness in cancer prevention and pathological complete response rate, to such an extent that a large phase III clinical trial in breast cancer is in progress.31,59-61

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- Mammucari C, Milan G, Romanello V, Masiero E, Rudolf R, Del Piccolo P, et al. FoxO3 controls autophagy in skeletal muscle in vivo. Cell Metab 2007; 6:458-71.
- Zhao J, Brault JJ, Schild A, Cao P, Sandri M, Schiaffino S, et al. FoxO3 coordinately activates protein degradation by the autophagic/lysosomal and proteasomal pathways in atrophying muscle cells. Cell Metab 2007; 6:472-83.
- Juhász G, Puskás LG, Komonyi O, Erdi B, Maróy P, Neufeld TP, Sass M. Gene expression profiling identifies FKBP39 as an inhibitor of autophagy in larval Drosophila fat body. Cell Death Differ 2007; 14:1181-90.
- Hu MC, Lee DF, Xia W, Golfman LS, Ou-Yang F, Yang JY, et al. IkappaB kinase promotes tumorigenesis through inhibition of forkhead FOXO3a. Cell 2004; 117:225-37.
- Yang JY, Zong CS, Xia W, Yamaguchi H, Ding Q, Xie X, et al. ERK promotes tumorigenesis by inhibiting FOXO3a via MDM2-mediated degradation. Nat Cell Biol 2008; 10:138-48.
- Jagani Z, Singh A, Khosravi-Far R. FoxO tumor suppressors and BCR-ABL-induced leukemia: a matter of evasion of apoptosis. Biochim Biophys Acta 2008; 1785:63-84.
- Trotman LC, Alimonti A, Scaglioni PP, Koutcher JA, Cordon-Cardo C, Pandolfi PP. Identification of a tumour suppressor network opposing nuclear Akt function. Nature 2006; 441:523-7.
- Kikuno N, Shiina H, Urakami S, Kawamoto K, Hirata H, Tanaka Y, et al. Knockdown of astrocyte-elevated gene-1 inhibits prostate cancer progression through upregulation of FOXO3a activity. Oncogene 2007; 26:7647-55.

- Cornforth AN, Davis JS, Khanifar E, Nastiuk KL, Krolewski JJ. FOXO3a mediates the androgen-dependent regulation of FLIP and contributes to TRAILinduced apoptosis of LNCaP cells. Oncogene 2008; 27:4422-33.
- Seoane J, Le HV, Shen L, Anderson SA, Massagué J. Integration of Smad and forkhead pathways in the control of neuroepithelial and glioblastoma cell proliferation. Cell 2004; 117:211-23.
- Greer EL, Oskoui PR, Banko MR, Maniar JM, Gygi MP, Gygi SP, et al. The energy sensor AMPactivated protein kinase directly regulates the mammalian FOXO3 transcription factor. J Biol Chem 2007; 282:30107-19.
- Shackelford DB, Shaw RJ. The LKB1-AMPK pathway: metabolism and growth control in tumour suppression. Nat Rev Cancer 2009; 9:563-75.
- Resta N, Simone C, Mareni C, Montera M, Gentile M, Susca F, et al. STK11 mutations in Peutz-Jeghers syndrome and sporadic colon cancer. Cancer Res 1998; 58:4799-801.
- Zheng B, Jeong JH, Asara JM, Yuan YY, Granter SR, Chin L, et al. Oncogenic B-RAF negatively regulates the tumor suppressor LKB1 to promote melanoma cell proliferation. Mol Cell 2009; 33:237-47.
- Inoki K, Ouyang H, Zhu T, Lindvall C, Wang Y, Zhang X, et al. TSC2 integrates Wnt and energy signals via a coordinated phosphorylation by AMPK and GSK3 to regulate cell growth. Cell 2006; 126:955-68.
- Zhou J, Huang W, Tao R, Ibaragi S, Lan F, Ido Y, et al. Inactivation of AMPK alters gene expression and promotes growth of prostate cancer cells. Oncogene 2009; 28:1993-2002.
- Canto C, Gerhart-Hines Z, Feige JN, Lagouge M, Noriega L, Milne JC, et al. AMPK regulates energy expenditure by modulating NAD\* metabolism and SIRT1 activity. Nature 2009; 458:1056-60.
- Fulco M, Cen Y, Zhao P, Hoffman EP, McBurney MW, Sauve AA, et al. Glucose restriction inhibits skeletal myoblast differentiation by activating SIRT1 through AMPK-mediated regulation of Nampt. Dev Cell 2008; 14:661-73.
- Brunet A, Sweeney LB, Sturgill JF, Chua KF, Greer PL, Lin Y, et al. Stress-dependent regulation of FOXO transcription factors by the SIRT1 deacetylase. Science 2004; 303:2011-5.

- Li XN, Song J, Zhang L, LeMaire SA, Hou X, Zhang C, et al. Activation of the AMPK-FOXO3 pathway reduces fatty acid-induced increase in intracellular reactive oxygen species by upregulating thioredoxin. Diabetes 2009; 58:2246-57.
- Colombo SL, Moncada S. AMPKalpha1 regulates the antioxidant status of vascular endothelial cells. Biochem J 2009; 421:163-9.
- McCay CM, Crowell MF, Maynard LA. The effect of retarded growth upon the length of life span and upon the ultimate body size. J Nutr 1935; 10:63-79.
- 42. Kirkwood TB, Shanley DP. Food restriction, evolution and ageing. Mech Ageing Dev 2005; 126:1011-6.
- Fontana L, Meyer TE, Klein S, Holloszy JO. Longterm calorie restriction is highly effective in reducing the risk for atherosclerosis in humans. Proc Natl Acad Sci USA 2004; 101:6659-63.
- 44. Maswood N, Young J, Tilmont E, Zhang Z, Gash DM, Gerhardt GA, et al. Caloric restriction increases neurotrophic factor levels and attenuates neurochemical and behavioral deficits in a primate model of Parkinson's disease. Proc Natl Acad Sci USA 2004; 101:18171-6.
- Michels KB, Ekbom A. Caloric restriction and incidence of breast cancer. Jama 2004; 291:1226-30.
- Grandison RC, Piper MD, Partridge L. Amino-acid imbalance explains extension of lifespan by dietary restriction in Drosophila. Nature 2009; Epub ahead of print.
- Greer EL, Brunet A. Different dietary restriction regimens extend lifespan by both independent and overlapping genetic pathways in *C. elegans*. Aging Cell 2009; 8:113-27.
- Mair W, Piper MD, Partridge L. Calories do not explain extension of life span by dietary restriction in Drosophila. PLoS Biol 2005; 3:223.
- Piper MD, Selman C, McElwee JJ, Partridge L. Separating cause from effect: how does insulin/IGF signalling control lifespan in worms, flies and mice? J Intern Med 2008; 263:179-91.
- Skorupa DA, Dervisefendic A, Zwiener J, Pletcher SD. Dietary composition specifies consumption, obesity and lifespan in *Drosophila melanogaster*. Aging Cell 2008; 7:478-90.

- Katic M, Kahn CR. The role of insulin and IGF-1 signaling in longevity. Cell Mol Life Sci 2005; 62:320-43.
- Jiang W, Zhu Z, Thompson HJ. Dietary energy restriction modulates the activity of AMP-activated protein kinase, Akt and mammalian target of rapamycin in mammary carcinomas, mammary gland, and liver. Cancer Res 2008; 68:5492-9.
- Greer EL, Banko MR, Brunet A. AMP-activated protein kinase and FoxO transcription factors in dietary restriction-induced longevity. Ann NY Acad Sci 2009; 1170:688-92.
- 54. Shaw RJ. Glucose metabolism and cancer. Curr Opin Cell Biol 2006; 18:598-608.
- Shackelford DB, Vasquez DS, Corbeil J, Wu S, Leblanc M, Wu CL, et al. mTOR and HIF-1alpha-mediated tumor metabolism in an LKB1 mouse model of Peutz-Jeghers syndrome. Proc Natl Acad Sci USA 2009; 106:11137-42.
- Yang JY, Hung MC. A new fork for clinical application: targeting forkhead transcription factors in cancer. Clin Cancer Res 2009; 15:752-7.
- Kisfalvi K, Eibl G, Sinnett-Smith J, Rozengurt E. Metformin disrupts crosstalk between G protein-coupled receptor and insulin receptor signaling systems and inhibits pancreatic cancer growth. Cancer Res 2009; 69:6539-45.
- Huang X, Wullschleger S, Shpiro N, McGuire VA, Sakamoto K, Woods YL, et al. Important role of the LKB1-AMPK pathway in suppressing tumorigenesis in PTEN-deficient mice. Biochem J 2008; 412:211-21.
- Evans JM, Donnelly LA, Emslie-Smith AM, Alessi DR, Morris AD. Metformin and reduced risk of cancer in diabetic patients. BMJ 2005; 330:1304-5.
- Libby G, Donnelly LA, Donnan PT, Alessi DR, Morris AD, Evans JM. New users of metformin are at low risk of incident cancer: a cohort study among people with type 2 diabetes. Diabetes Care 2009; 32:1620-5.
- Memmott RM, Dennis PA. Akt-dependent and -independent mechanisms of mTOR regulation in cancer. Cell Signal 2009; 21:656-64.