

Draft Genome Sequences of 40 *Pseudomonas aeruginosa* Clinical Strains Isolated from the Sputum of a Single Cystic Fibrosis Patient Over an 8-Year Period

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We report draft genome sequences of 40 *Pseudomonas aeruginosa* strains, isolated from the sputum of a single cystic fibrosis patient over eight years. Analyses indicated a correlation between multidrug-resistant phenotypes and population structure. Our data provide new insights into the mechanisms leading to acquisition of antibiotic resistance in *P. aeruginosa*.

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Pseudomonas aeruginosa is the most pervasive of all recognized pathogens in the nosocomial environment, causing pulmonary and bloodstream infection with mortality rates of up to 50% (1). Multi-drug-resistant (MDR) *P. aeruginosa* strains are emerging with increasing frequency and infection rates have tripled over the past two decades (2, 3). Some *P. aeruginosa* strains have been found to be resistant to nearly all or all antibiotics in clinical use (4).

Cystic fibrosis (CF) patients infected with resistant *P. aeruginosa* are exposed to increased mortality and morbidity (5, 6) and estimates indicate that 25 to 45% of adult CF patients are chronically infected with MDR *P. aeruginosa* within their airway (7). The bacterium develops MDR phenotypes during its persistence in a CF patient's airway by accumulating pathoadaptive mutations (8). Whole-genome sequencing (WGS) can help to point out potential molecular mechanisms of resistance and has already proved to be able to predict antimicrobial susceptibility in several pathogens (9, 10). However, despite the fact that several WGS studies on *P. aeruginosa* CF lineages have been published (11–14), their evolutionary trajectories in relation to the development of antimicrobial resistance remain mostly unexplored to date.

To track the pathoadaptive changes leading to the development of MDR in *P. aeruginosa* during its microevolution in a CF patient's airway, we obtained whole-genome sequences of 40 *P. aeruginosa* clinical CF strains isolated at Trentino Regional Support CF Centre (Rovereto, Italy) from the sputum of a single CF patient over an eight-year period (2007 to 2014). Interestingly, despite a high degree of genome sequence conservation, isolates evolved toward the acquisition of an MDR phenotype over time.

Bacteria were grown in Luria-Bertani broth overnight at 37°C in a shaking incubator. Cells were harvested and genomic DNA was extracted using the DNeasy blood and tissue kit (Qiagen, Germany) following the manufacturer's instructions for Gram-negative bacteria. Genomic DNA libraries were prepared using the Nextera XT DNA library preparation kit and protocols (Illumina, USA) and sequenced on the Illumina MiSeq platform at the Next Generation

Sequencing (NGS) Core Facility of the Centre for Integrative Biology, University of Trento. Assembly of draft genomes was carried out using SPAdes version 3.1.0 (15). To improve the assemblies' qualities, raw reads were mapped on the contigs using Bowtie2 v2.2.6 (16) and contigs with less than three reads mapping and/or with coverage below 1 were removed.

Identification of MLST profiles (sequence types) was performed *in silico* from *de novo* assembled genomes using MLST 1.8 (Table 1) (17).

The average number of contigs per genome was 101 with a standard deviation of 56. Draft genomes ranged in size from 6,545 kbp to 6,653 kb with a G+C content of 66.28% (Table 1). The N_{50} of the draft genomes ranged from 30,645 to 378,317 bp with an average of 179,843 bp (Table 1).

Accession number(s). This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank. See Table 1 for accession numbers of the single genomes. The version described in this paper is the first one.

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REFERENCES

- Osmon S, Ward S, Fraser VJ, Kollef MH. 2004. Hospital mortality for patients with bacteremia due to *Staphylococcus aureus* or *Pseudomonas aeruginosa*. *Chest* 125:607–616. <http://dx.doi.org/10.1378/chest.125.2.607>.
- Obritsch MD, Fish DN, MacLaren R, Jung R. 2005. Nosocomial infections due to multidrug-resistant *Pseudomonas aeruginosa*: epidemiology and treatment options. *Pharmacotherapy* 25:1353–1364. <http://dx.doi.org/10.1592/phco.2005.25.10.1353>.
- Lautenbach E, Weiner MG, Nachamkin I, Bilker WB, Sheridan A, Fishman NO. 2006. Imipenem resistance among *Pseudomonas aeruginosa* isolates: risk factors for infection and impact of resistance on clinical and

- economic outcomes. *Infect Control Hosp Epidemiol* 27:893–900. <http://dx.doi.org/10.1086/507274>.
4. Centers for Disease Control and Prevention. 2013. Antibiotic resistance threats in the United States, 2013. CDC, Atlanta, GA.
 5. Lyczak JB, Cannon CL, Pier GB. 2000. Establishment of *Pseudomonas aeruginosa* infection: lessons from a versatile opportunist. *Microbes Infect* 2:1051–1060. [http://dx.doi.org/10.1016/S1286-4579\(00\)01259-4](http://dx.doi.org/10.1016/S1286-4579(00)01259-4).
 6. Chmiel JF, Davis PB. 2003. State of the art: why do the lungs of patients with cystic fibrosis become infected and why can't they clear the infection? *Respir Res* 4:8. <http://dx.doi.org/10.1186/1465-9921-4-8>.
 7. Lechtzin N, John M, Irizarry R, Merlo C, Diette GB, Boyle MP. 2006. Outcomes of adults with cystic fibrosis infected with antibiotic-resistant *Pseudomonas aeruginosa*. *Respiration* 73:27–33. <http://dx.doi.org/10.1159/000087686>.
 8. Breidenstein EBM, de la Fuente-Núñez C, Hancock RE. 2011. *Pseudomonas aeruginosa*: all roads lead to resistance. *Trends Microbiol* 19: 419–426. <http://dx.doi.org/10.1016/j.tim.2011.04.005>.
 9. Stoesser N, Batty EM, Eyre DW, Morgan M, Wyllie DH, Del Ojo Elias C, Johnson JR, Walker AS, Peto TEA, Crook DW. 2013. Predicting antimicrobial susceptibilities for *Escherichia coli* and *Klebsiella pneumoniae* isolates using whole genomic sequence data. *J Antimicrob Chemother* 68:2234–2244. <http://dx.doi.org/10.1093/jac/dkt180>.
 10. Zankari E, Hasman H, Kaas RS, Seyfarth AM, Agersø Y, Lund O, Larsen MV, Aarestrup FM. 2013. Genotyping using whole-genome sequencing is a realistic alternative to surveillance based on phenotypic antimicrobial susceptibility testing. *J Antimicrob Chemother* 68:771–777. <http://dx.doi.org/10.1093/jac/dks496>.
 11. Darch SE, McNally A, Harrison F, Corander J, Barr HL, Paszkiewicz K, Holden S, Fogarty A, Cruz SA, Diggle SP. 2015. Recombination is a key driver of genomic and phenotypic diversity in a *Pseudomonas aeruginosa* population during cystic fibrosis infection. *Sci Rep* 5:7649. <http://dx.doi.org/10.1038/srep07649>.
 12. Marvig RL, Sommer LM, Molin S, Johansen HK. 2015. Convergent evolution and adaptation of *Pseudomonas aeruginosa* within patients with cystic fibrosis. *Nat Genet* 47:57–64. <http://dx.doi.org/10.1038/ng.3148>.
 13. Williams D, Evans B, Haldenby S, Walshaw MJ, Brockhurst MA, Winstanley C, Paterson S. 2015. Divergent, coexisting *Pseudomonas aeruginosa* lineages in chronic cystic fibrosis lung infections. *Am J Respir Crit Care Med* 191:775–785. <http://dx.doi.org/10.1164/rccm.201409-1646OC>.
 14. Caballero JD, Clark ST, Coburn B, Zhang Y, Wang PW, Donaldson SL, Elizabeth Tullis D, Yau YCW, Waters VJ, Hwang DM, Guttman DS. 2015. Selective sweeps and parallel pathoadaptation drive *Pseudomonas aeruginosa* evolution in the cystic fibrosis lung. *mBio* 6:e00981-15.
 15. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <http://dx.doi.org/10.1089/cmb.2012.0021>.
 16. Langmead B, Salzberg SL. 2012. Fast gapped-read alignment with Bowtie 2. *Nat Methods* 9:357–359. <http://dx.doi.org/10.1038/nmeth.1923>.
 17. Larsen MV, Cosentino S, Rasmussen S, Friis C, Hasman H, Marvig RL, Jelsbak L, Sicheritz-Pontén T, Ussery DW, Aarestrup FM, Lund O. 2012. Multilocus sequence typing of total-genome-sequenced bacteria. *J Clin Microbiol* 50:135–1361.

TABLE 1 Draft genome sequences and global statistics of the 40 *P. aeruginosa* CF isolates

| Accession no. | Isolate name | Yr of isolation | Sequence type | No. of contigs | Genome size (kb) | N_{50} (kb) | G+C content (%) |
|---------------|--------------|-----------------|---------------|----------------|------------------|---------------|-----------------|
| MAUO00000000 | TNCF_3 | 2007 | 390 | 139 | 6,636 | 92 | 66.28 |
| MAUP00000000 | TNCF_4M | 2007 | 390 | 161 | 6,630 | 78 | 66.29 |
| MAUQ00000000 | TNCF_6 | 2007 | 390 | 356 | 6,618 | 31 | 66.28 |
| MAUR00000000 | TNCF_7M | 2007 | 390 | 259 | 6,623 | 47 | 66.28 |
| MAUS00000000 | TNCF_10 | 2007 | 390 | 101 | 6,643 | 143 | 66.28 |
| MAUT00000000 | TNCF_10M | 2007 | 390 | 107 | 6,633 | 111 | 66.29 |
| MAZG00000000 | TNCF_12 | 2007 | 390 | 102 | 6,545 | 177 | 66.36 |
| MAZI00000000 | TNCF_13 | 2007 | 390 | 75 | 6,637 | 195 | 66.27 |
| MAZH00000000 | TNCF_14 | 2007 | 390 | 89 | 6,633 | 158 | 66.28 |
| MAKL00000000 | TNCF_16 | 2007 | 1864 | 59 | 6,638 | 269 | 66.28 |
| MAZJ00000000 | TNCF_23 | 2007 | 390 | 71 | 6,635 | 228 | 66.28 |
| MAZK00000000 | TNCF_23M | 2007 | 390 | 64 | 6,636 | 228 | 66.28 |
| MAKM00000000 | TNCF_32 | 2007 | 390 | 67 | 6,639 | 229 | 66.28 |
| MAZL00000000 | TNCF_32M | 2007 | 390 | 138 | 6,627 | 93 | 66.28 |
| MAZM00000000 | TNCF_42 | 2008 | 390 | 70 | 6,639 | 228 | 66.28 |
| MAZN00000000 | TNCF_42M | 2008 | 390 | 71 | 6,640 | 228 | 66.28 |
| MAZO00000000 | TNCF_49M | 2008 | 390 | 76 | 6,635 | 177 | 66.29 |
| MAZP00000000 | TNCF_68 | 2010 | 390 | 82 | 6,633 | 162 | 66.28 |
| MAZQ00000000 | TNCF_69 | 2010 | 1863 | 88 | 6,639 | 150 | 66.28 |
| MAZR00000000 | TNCF_76 | 2010 | 390 | 61 | 6,634 | 281 | 66.28 |
| MAZS00000000 | TNCF_85 | 2010 | 1864 | 101 | 6,644 | 124 | 66.29 |
| MAZT00000000 | TNCF_88M | 2010 | 1864 | 65 | 6,636 | 229 | 66.28 |
| MAZU00000000 | TNCF_101 | 2011 | 1864 | 142 | 6,653 | 92 | 66.28 |
| MAZV00000000 | TNCF_105 | 2011 | 390 | 92 | 6,644 | 191 | 66.28 |
| MAZW00000000 | TNCF_106 | 2011 | 390 | 77 | 6,634 | 205 | 66.28 |
| MAZX00000000 | TNCF_109 | 2011 | 390 | 69 | 6,634 | 205 | 66.28 |
| MAZD00000000 | TNCF_130 | 2012 | 390 | 157 | 6,625 | 76 | 66.28 |
| MAZF00000000 | TNCF_133 | 2012 | 390 | 82 | 6,637 | 154 | 66.29 |
| MAZE00000000 | TNCF_133_1 | 2012 | 1864 | 87 | 6,641 | 269 | 66.28 |
| MAKK00000000 | TNCF_151 | 2013 | 390 | 53 | 6,629 | 378 | 66.28 |
| MBM00000000 | TNCF_151M | 2013 | 1864 | 103 | 6,636 | 143 | 66.28 |
| MBMJ00000000 | TNCF_154 | 2013 | 390 | 86 | 6,635 | 177 | 66.28 |
| MBMK00000000 | TNCF_155 | 2013 | 390 | 62 | 6,634 | 339 | 66.28 |
| MBML00000000 | TNCF_155_1 | 2013 | 1923 | 71 | 6,635 | 221 | 66.28 |
| MBMM00000000 | TNCF_165 | 2013 | 1923 | 119 | 6,634 | 135 | 66.28 |
| MBMN00000000 | TNCF_167 | 2013 | 390 | 73 | 6,634 | 191 | 66.27 |
| MBMO00000000 | TNCF_167_1 | 2013 | 390 | 91 | 6,628 | 143 | 66.28 |
| MBMP00000000 | TNCF_174 | 2014 | 390 | 111 | 6,645 | 143 | 66.29 |
| MBMQ00000000 | TNCF_175 | 2014 | 390 | 118 | 6,642 | 124 | 66.28 |
| MBMR00000000 | TNCF_176 | 2014 | 1923 | 61 | 6,637 | 354 | 66.28 |