

REVIEW

Open Access



Use of gadolinium-based contrast agents in head and neck cancer diagnosis, staging, and monitoring: current applications and future perspectives

Marco Parillo^{1,2*} , Federica Vaccarino², Andrea Falzone², Elena Salvador³, Fabio M. Doniselli⁴, Carlo C. Quattrocchi^{2,5} and Àlex Rovira⁶

Abstract

Gadolinium-based contrast agents (GBCAs) have been fundamental to head and neck cancer (HNC) imaging, enabling effective detection, characterization, treatment response assessment, and disease progression monitoring of lesions. Additionally, perfusion-weighted imaging (PWI) utilizing dynamic contrast enhancement (DCE) has been evaluated for its ability to provide insights into microvascular parameters concerning blood flow within tumor tissue. Nevertheless, increasing worries regarding gadolinium accumulation within the central nervous system and its effects on the environment have led to a reconsideration of its application. This narrative review explores the current role of GBCAs in HNC imaging, the primary sequences used after GBCA administration, their interpretation, and potential alternative imaging approaches. Currently, GBCA administration is a cornerstone of multiparametric MRI for the diagnosis, staging, and monitoring of HNCs, commonly involving a 3D T1-weighted sequence with fat saturation during the equilibrium phase. While PWI shows potential for clinical application in HNCs, its broader clinical adoption requires further standardization. Notably, DCE can visually aid in detecting subtle tumors, and its application in the differential diagnosis of solid parotid lesions is yielding promising results. Arterial spin labeling is emerging as a compelling alternative for PWI, eliminating the need for GBCA administration. Other promising strategies for reducing or even avoiding GBCA use include hybrid PET/MRI examinations, the development of novel contrast agents (including high-relaxivity GBCAs and gadolinium-free contrast agents), and the implementation of artificial intelligence tools.

Key Points

Question *When should GBCAs be administered to patients undergoing MRI for HNCs?*

Findings *GBCA injection is a cornerstone of multiparametric MRI for the diagnosis, staging, and monitoring of HNCs.*

Clinical relevance *GBCAs are recommended for HNC MRIs, with a possible exception for patients with no clinical or radiological evidence of recurrence after 27 months of follow-up. DCE is useful for identifying small carcinomas and characterizing parotid lesions.*

Keywords Gadolinium, Contrast media, Magnetic resonance imaging, Head and neck neoplasms, Practice guideline

*Correspondence:

Marco Parillo

marco.parillo@univr.it

Full list of author information is available at the end of the article



© The Author(s) 2025. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

Graphical Abstract

Use of gadolinium-based contrast agents in head and neck cancer diagnosis, staging and monitoring: current applications and emerging perspectives

What is the role of gadolinium-based contrast agents (GBCAs) administration in patients undergoing MRI for head and neck cancers (HNCs)?

- **Narrative review**
- **GBCAs for diagnosis, staging and monitoring of HNCs**
- **Strategies for reducing or avoiding GBCAs**



	GBCA dose and injection	Sequences		
		Suggested	Optional	Not suggested
Diagnosis	Dose: 0.1 mmol/Kg for Gadoterate, Gadobutrol, and Gadoteridol; 0.05 mmol/Kg for Gadopidrenol; 0.04 mmol/Kg for Gadoquatran Injection: power injector (rate between 2-4 ml/s) is mandatory if dynamic contrast-enhancement is performed, and it is recommended over manual injection in all other cases	3D isotropic T1-weighted with fat-saturation	DCE for identification of HNC of unknown primary and characterization of parotid lesions 2D T1-weighted with fat-saturation	Arterial spin labeling
Staging and monitoring		3D isotropic T1-weighted with fat-saturation	2D T1-weighted with fat-saturation	DCE Arterial spin labeling

GBCAs are usually required in HNCs, unless recurrence is absent after 27 months. Dynamic contrast enhancement (DCE) is valuable for identifying small tumors and characterizing parotid lesions

Eur Radiol (2025) Parillo M, Vaccarino F, Falzone A, et al.; DOI:10.1007/s00330-025-12165-0



Introduction

Head and neck cancers (HNCs) comprise a heterogeneous group of malignant diseases that can arise in different locations, such as the oral cavity, pharynx, larynx, paranasal sinuses, and salivary glands [1–4]. The estimated percentages of new cancer cases in the United States in 2025 for the main neck sites relative to all other cancers are: 1.86% for the oral cavity, 1.06% for the pharynx, and 0.64% for the larynx [5]. Over 90% of HNCs originate from epithelial cells, with squamous cell carcinoma (SCC) being the most prevalent type within this group [6].

MRI has become a crucial imaging technique for staging and monitoring HNCs, offering superior soft tissue differentiation compared to computed tomography (CT) [7]. Standard MRI protocols for HNCs include the intravenous injection of gadolinium-based contrast agents (GBCAs), which provide essential information on tumor vascularity and the degree of tumor spread that could not be apparent on unenhanced sequences (e.g., perineural or dural invasion) [8]. Contrast-enhancement significantly improves the delineation of tumor margins, differentiation between different pathological conditions, and evaluation of treatment response. In addition, perfusion-weighted imaging (PWI) allows for the evaluation of microvascular parameters related to blood flow within the tumor tissue [9].

The safety profile of GBCAs has been a subject of evolving clinical understanding. While these agents are generally safe, two key concerns have emerged. Initially, a primary risk was nephrogenic systemic fibrosis, which was linked to the use of linear GBCAs in patients with severe kidney issues. New cases of this rare disease have become very uncommon since the use of GBCAs was restricted in subjects with renal failure [10]. More recently, attention has shifted to gadolinium retention in the brain and other tissues related to dechelation of GBCAs (linear > macrocyclic), also in patients with preserved renal function [11, 12]. Although no negative symptoms have been directly linked to this deposition, regulatory agencies have issued guidelines to address this concern. In fact, the European Commission has restricted the use of linear GBCAs for head and neck MRIs, emphasizing judicious use of the remaining macrocyclic agents (i.e., Gadoterate, Gadobutrol, and Gadoteridol) [13]. According to the latest European Society of Urogenital Radiology guidelines, mandatory renal function testing is not required before administering macrocyclic GBCAs, but they should be used cautiously in patients with an estimated glomerular filtration rate < 30 mL/min/1.73 m² [14]. Additionally, there are increasing environmental worries linked to the mining of gadolinium and its subsequent dispersion into

the ecosystem following renal excretion [15–17]. Given their high stability, GBCAs are not eliminated by standard wastewater treatment, resulting in widespread contamination with gadolinium of freshwater and drinking water. While the long-term effects of this environmental gadolinium on humans and marine life are still being studied, it is important to monitor and evaluate its impact as more information emerges [18, 19].

Based on these concerns, recommendations for the proper use of GBCAs have recently been published regarding patient categories with chronic diseases who undergo repeated contrast-enhanced MRI over their lifetime (e.g., multiple sclerosis and intracranial tumors) [20–22]. However, a comprehensive evaluation regarding GBCA usage in HNCs is still lacking.

In this narrative review, we aim to provide a framework for clinicians on the role of GBCAs in the diagnosis, staging, and monitoring of HNCs, the main sequences employed after GBCA administration, along with their interpretation and potential alternatives. In addition, it discusses future developments in hybrid imaging, novel contrast agents, and artificial intelligence (AI) tools.

MRI sequences

When performing contrast-enhanced T1-weighted MRI of the neck, turbo- or fast spin echo and gradient-echo sequences are typically used. To effectively differentiate enhancing tissues from fat, fat suppression techniques are crucial. These commonly include spectral or frequency-selective fat suppression or chemical shift-based Dixon methods, the latter particularly useful for reducing artifacts related to magnetic field inhomogeneities by dental prostheses [8, 9]. Historically, contrast-enhanced imaging of the neck involved acquiring separate 2D images in the axial and coronal planes. However, 3D post-contrast T1-weighted imaging has now become routine. This approach offers several advantages, including thinner slices with higher spatial resolution, improved vascular assessment, and reduced total scan time [8]. Notably, a 3D post-contrast T1-weighted sequence is typically the only 3D sequence included in head and neck MRI protocols. The possibility of generating multiplanar reconstructions is particularly valuable for evaluating the tumor's relationship with the complex regional anatomy. For example, 3D gradient-echo sequences allow for acquiring high-resolution volumetric imaging while reducing motion artifacts, without sacrificing either image clarity or the detectability of tumors [23–25]. Newer techniques, like radial imaging, now produce higher-resolution images with better overall quality and fewer artifacts than traditional T1-weighted turbo spin echo sequences [26].

PWI in HNCs is commonly obtained through dynamic contrast-enhanced (DCE) MRI. This involves acquiring a

rapid series of T1-weighted gradient-echo images before, during, and after an intravenous injection of a GBCA [27]. Regions of interest are placed within the tumor to calculate various semiquantitative parameters, such as wash-in and wash-out rates (the rate at which the GBCA enters and leaves the tumor), peak enhancement (the highest concentration of contrast reached), time-to-peak enhancement (the time it takes to reach the maximum enhancement), and the area under the curve (the overall amount of enhancement present over time) [9]. Beyond semiquantitative metrics, pharmacokinetic modeling provides quantitative parameters in DCE-MRI, including: K^{trans} (volume transfer coefficient describing the movement between plasma and the extravascular extracellular space), k_{ep} (rate constant indicating the transfer rate from the extravascular extracellular space back to the plasma), v_e (the fractional volume of the extravascular extracellular space, indicating the available space within the tissue interstitium for the accumulation of GBCA), and v_p (the fractional plasma volume, reflecting the blood plasma within a given tissue volume) [9, 28]. Arterial spin labeling (ASL) has emerged as a promising non-contrast alternative to DCE-MRI for perfusion assessment in HNCs [29, 30]. In ASL, a radiofrequency pulse magnetically labels water protons in arterial blood by saturating them. By subtracting the labeled images from the control images, static background signals are removed, isolating a perfusion-dependent signal [31]. Labeling techniques include continuous, pulsed, and velocity-selective [29]; among these, pseudo-continuous ASL (pCASL) is recommended for neck imaging, using a rapid sequence of radiofrequency pulses to achieve labeling at a rate of roughly one pulse per millisecond [32]. In clinical practice, ASL in the neck is typically approximated by visually examining the subtracted images to determine where the tagged spins are distributed. This approach, however, can lead to biased and subjective evaluations because the background signal is not always optimally suppressed [33]. The most effective way to leverage ASL data involves a quantitative approach, particularly for calculating the tumor blood flow (TBF) in neck lesions [34, 35].

Table 1 summarizes the GBCA doses and contrast-enhanced MRI sequences applicable in daily clinical practice in HNCs.

GBCA in the diagnosis of HNCs

From a diagnostic perspective, head and neck masses present a considerable challenge, with a broad differential that includes inflammatory, infectious, and neoplastic conditions [36].

HNCs typically exhibit intermediate T2 signal intensity (higher than muscle), low T1 signal intensity, and less contrast-enhancement than normal mucosa on MRI. GBCAs improve the visibility of small solid lesions (e.g.,

Table 1 Summary of suggested GBCAs doses and contrast-enhanced magnetic resonance imaging (MRI) sequences applicable in daily clinical practice in HNCs

GBCA dose and injection		Sequences	Not suggested
		Suggested	Optional
Diagnosis	<ul style="list-style-type: none"> Dose: 0.1 mmol/Kg for Gadoterate, Gadobutrol, and Gadoteridol; 0.05 mmol/Kg for Gadopiclesol; 0.04 mmol/Kg for Gadoquatrane Injection: power injector (rate between 2 and 4 mL/s) is mandatory if DCE is performed, and it is recommended over manual injection in all other cases 	<ul style="list-style-type: none"> 3D isotropic T1-weighted with fat-saturation 2D T1-weighted with fat-saturation* 	<ul style="list-style-type: none"> ASL
Staging and monitoring*		<ul style="list-style-type: none"> 3D isotropic T1-weighted with fat-saturation 	<ul style="list-style-type: none"> DCE ASL

ASL arterial spin labeling, DCE dynamic contrast-enhancement, GBCA gadolinium-based contrast agents, HNCs head and neck cancers
 *The appropriate plane is selected according to the tumor's site and spread, with a minimum of 2 planes if 3D imaging is not performed
 *For monitoring patients who continue to undergo follow-up MRI scans beyond 27 months post-treatment, with previous radiological scans negative for disease recurrence and no suspicious clinical findings (e.g., endoscopy negative for suspected tumor recurrence, neck palpation negative for new cervical lymphadenopathies or masses, absence of new signs or symptoms attributable to cranial nerve involvement), GBCA may be injected only when interval changes are detected on unenhanced sequences. This conditional suggestion is inferred from the Neck Imaging Reporting and Data System's surveillance algorithm, though it awaits further prospective validation

tongue border and tonsillar regions, Fig. 1) and enable their distinction from lesions with purulent (e.g., abscess) or mainly cystic (e.g., congenital lesion) contents [9]. When examining the sinonasal cavity, contrast-enhanced MRI aids in differentiating tumors from benign secretions (Fig. S1) and is considered superior to CT in distinguishing inflammatory from neoplastic processes [37].

DCE-MRI scans aimed at distinguishing SCC from lymphoma have employed diverse imaging protocols and yielded somewhat inconsistent outcomes. However, a general trend in the literature suggests that SCC exhibits greater tumor perfusion and capillary permeability compared to lymphoma. This is often indicated by higher K^{trans} and v_e values, as well as greater peak contrast-enhancement and a shorter time-to-peak on time-intensity curve analysis [38]. Furthermore, visual evaluation of early-phase DCE-MRI can reveal minute tumors not visible on conventional sequences, which may be crucial in cases of HNC of unknown primary (Fig. S2). DCE-MRI also facilitates differentiation of hypovascular tumors (e.g., schwannomas) from hypervascular tumors (e.g., paraganglioma) [28, 38]. Parotid lesions might be differentiated based on their DCE-MRI patterns using three key parameters: time-to-peak, wash-out ratio, and the shape of the time-intensity curve (Table 2 and Fig. 2) [39]. Finally, preliminary studies suggest that ASL can distinguish SCC from other types of benign or malignant tumors in regions like the paranasal sinuses, upper airway, parotid glands, and orbits by measuring differences in TBF [29, 30].

Therefore, in head and neck MRI, especially in cases where HNCs are suspected, GBCAs provide vital supplementary data for a more conclusive diagnosis. While routine use of PWI is not yet standard [40], DCE-MRI can assist in visually detecting small tumors and characterizing solid parotid lesions. ASL enables the distinction between solid and cystic lesions and allows for the estimation of TBF in neoplasms, making it a compelling alternative to DCE-MRI for evaluating HNCs.

GBCA in the staging of HNCs

From a staging perspective, MRI is generally the preferred technique for suprahypoid disease. Conversely, contrast-enhanced CT is often the first-line choice for infrahyoid sites, which are more susceptible to motion artifacts. However, the use of MRI for the larynx and hypopharynx is becoming more common [41]. Distinguishing between an SCC and its adjacent inflammatory reaction is feasible through a detailed analysis of multiparametric MRI signal characteristics. Notably, inflammatory processes tend to show greater signal intensity on T2 sequences and enhance more robustly following GBCA administration when compared to the tumor [42]. Contrast-enhanced MRI is crucial in evaluating intraorbital or intracranial extension, commonly seen in carcinomas originating from

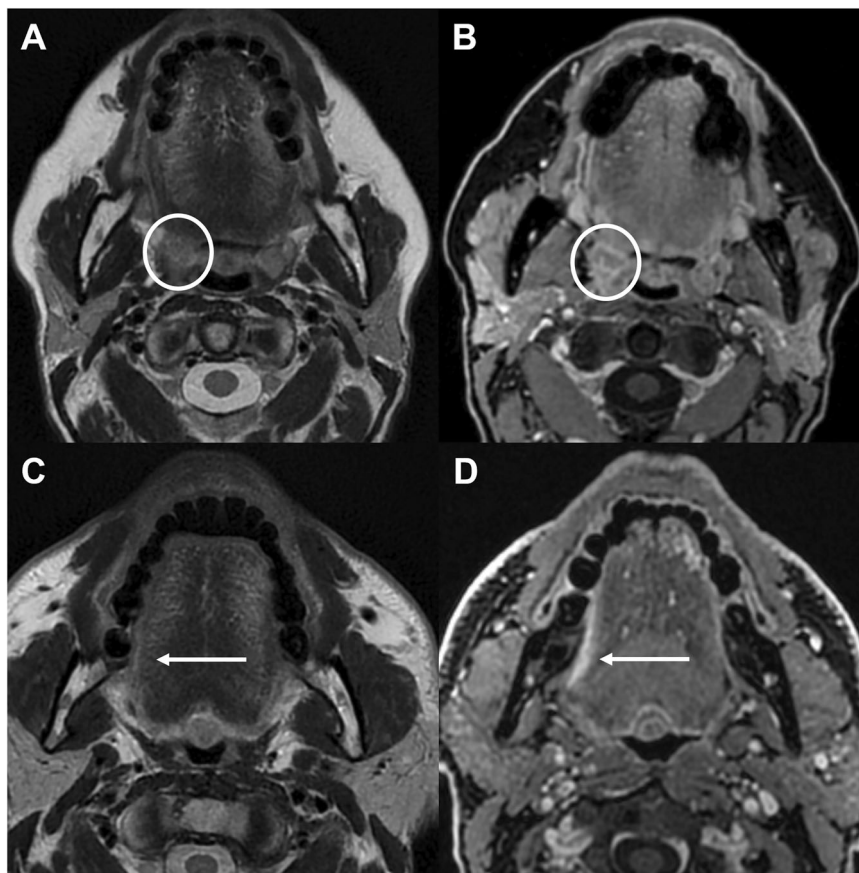


Fig. 1 In a patient with a small right tonsillar tumor (circles in **A, B**) and one with a small right lateral tongue tumor (arrows in **C, D**), fat-saturated T1-weighted images after GBCAs administration (**B, D**) better identify the neoplasm compared to T2-weighted images (**A, C**). GBCA, gadolinium-based contrast agents

Table 2 Key characteristics of parotid tumors detectable by DCE-MRI

Tumor type	Time-intensity curve type	Specification
Pleomorphic adenoma	A	<ul style="list-style-type: none"> • Prolonged time-to-peak (> 120 s) • No wash-out
Warthin tumor	B	<ul style="list-style-type: none"> • Ascending plateau • Short time-to-peak (≤ 120 s)
Malignant tumors	C	<ul style="list-style-type: none"> • Significant wash-out ($\geq 30\%$) • Short time-to-peak (≤ 120 s) • Minimal wash-out (< 30%)
Cyst	D	<ul style="list-style-type: none"> • Descending plateau • No enhancement

DCE dynamic contrast-enhanced

the nasal cavity, paranasal sinuses, and nasopharynx [8]. Contrast-enhanced MRI is also the favored method for visualizing perineural spread (PNS), which is a crucial factor in predicting the outcome of HNCs [43]. PNS appears as asymmetric nerve enhancement, thickening, or obliteration of fat at neural foramina. Advanced PNS may

also show bony erosion of the skull base foramina [44]. Figure 3 shows examples of contrast-enhanced images for defining the locoregional extent of HNCs.

According to the primary tumor location, specific features warrant focus during the evaluation of post-contrast imaging, as they impact accurate T staging [45–49].

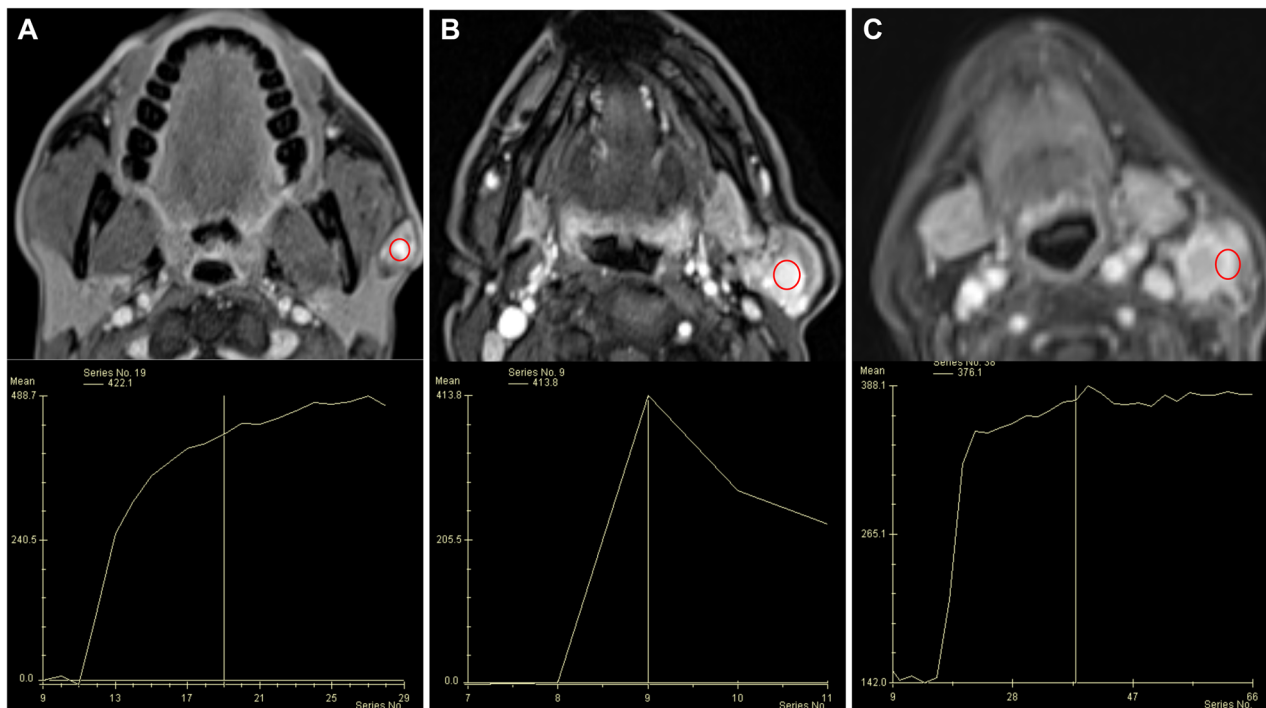


Fig. 2 Differential diagnosis of solid left parotid lesions using dynamic-contrast enhancement and time-intensity curve analysis. The red circles represent the region of interest placed on the vascularized part of the tumor for the generation of intensity-time curves. **A** Pleomorphic adenoma with a Type A curve (prolonged time-to-peak: 150 s). **B** Warthin's tumor with a Type B curve (short time-to-peak: 45 s, significant wash-out: > 30%). **C** Adenocarcinoma with a Type C curve (short time-to-peak: 93 s, minimal wash-out: < 30%)

PWI has been used to predict treatment responses. Despite the intricate biological interplay between tumor perfusion, permeability, and how tumors respond to chemoradiation, current findings generally suggest that tumors exhibiting good baseline perfusion and permeability, as indicated by higher K^{trans} values on DCE-MRI, tend to have less hypoxia. This lower hypoxia is associated with increased sensitivity to chemoradiation treatment and improved survival outcomes [38]. Moreover, as highlighted in the literature and recognized by experts, PWI may also have utility in radiotherapy planning for gross tumor volume delineation [28, 40]. ASL has shown potential for predicting treatment outcomes in HNCs, considering that lower TBF values prior to treatment have been linked to a less favorable prognosis [35].

Finally, it is important to remember that the use of GBCAs helps in delineating the structure of cervical lymph nodes, which, if involved, influence the patient's staging. For example, contrast-enhanced MRI facilitates the identification of cystic lymph nodes, often associated with human papilloma virus-positive oropharyngeal SCC, as well as signs of extra-nodal extension [50, 51]. Irregularities at the lymph node borders may indicate extra-nodal extension, where cancer cells breach the nodal

capsule. This often appears on imaging as capsular enhancement, indistinct margins, and surrounding fat stranding [45, 51]. PWI has also been tested in N staging. Lymph node metastases exhibiting good baseline perfusion and permeability, as indicated by higher K^{trans} values on DCE-MRI, tend to show greater sensitivity to chemoradiation and, consequently, improved survival rates. Conversely, nodal metastases with low K^{trans} values are generally associated with poorer prognoses [38]. Furthermore, DCE-MRI may also aid in distinguishing metastatic from benign lymph nodes by revealing reduced extracellular extravascular space volume and slower contrast transfer, potentially enhancing the accuracy of nodal staging [52].

In summary, GBCA-enhanced imaging represents a key component of multiparametric MRI analysis for T and N assessment in patients with HNCs. Additional evidence is necessary to ascertain the prognostic value of PWI (with DCE-MRI and/or ASL) in the evaluation of both the primary tumor and cervical lymph nodes.

GBCA in the monitoring of HNCs

From a monitoring perspective, imaging is crucial in the management of HNCs to detect any residual disease or

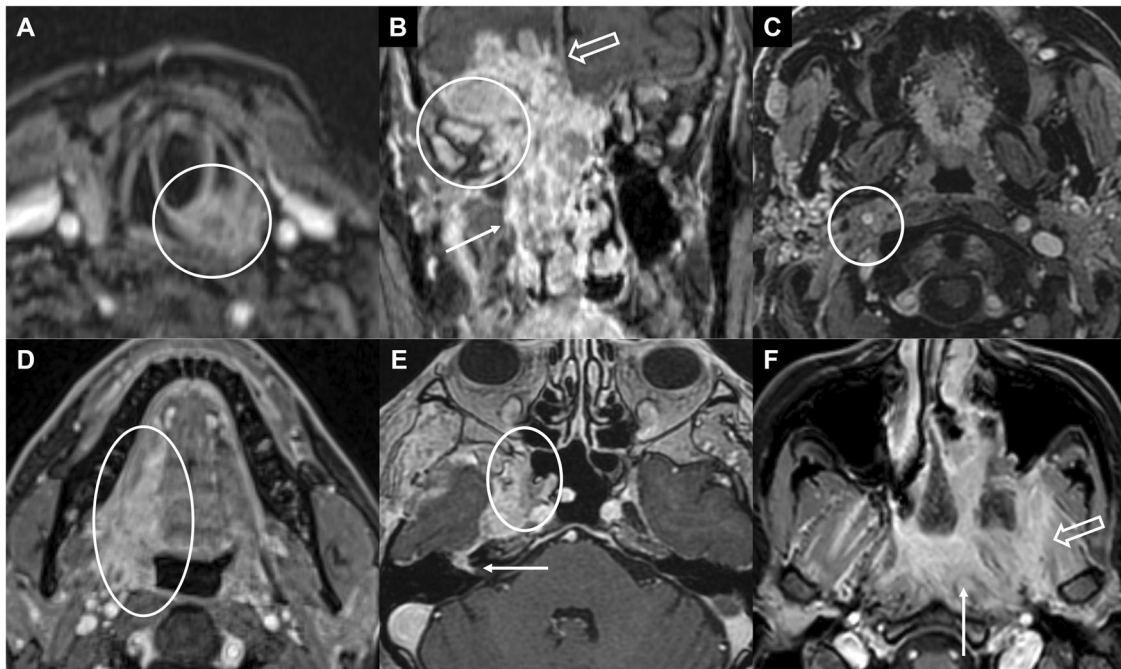


Fig. 3 Examples of T1-weighted images after GBCA administration for defining the locoregional extent of disease. **A** Left laryngeal tumor. Both the cricoid cartilage infiltration and the extra-laryngeal extension are well-delineated (circle). **B** Extensive intracranial (empty arrow), intraorbital (circle), and ethmoido-nasal (arrow) infiltration in a sinonasal undifferentiated carcinoma. The contrast agent makes it possible to clearly distinguish the tumor tissue from the mucosal thickening in the right maxillary sinus and better delineates the infiltration of the ocular muscles and cranial cavity. **C** Encasement of the right carotid space (circle) with

occlusion of the internal jugular vein in a right tonsill carcinoma. **D** Right tonsillar tumor. The extension to the oral floor and tongue is well-delineated (ellipse). **E** Nasopharyngeal carcinoma extending to the right pterygopalatine fossa (circle). The centripetal PNS along the greater superficial petrosal nerve to the geniculate ganglion and along the facial nerve in the internal auditory canal (arrow) is well-delineated. **F** Extensive infiltration of the left prevertebral (arrow) and masticator (empty arrow) spaces in a nasopharyngeal tumor. GBCA, gadolinium-based contrast agents; PNS, perineural spread

recurrence at an early stage, thereby optimizing the potential for effective salvage therapies. Surveillance imaging protocols differ in their recommendations for timing and modality. Generally, a baseline scan is advised 3–6 months post-treatment, depending on therapy type and patient risk [53]. Distinguishing recurrent tumors from the typical tissue changes following surgery, radiation, or chemotherapy poses a major challenge in post-treatment imaging [53–56]. Contrast-enhanced MRI can be especially valuable in addressing this difficulty (Table 3). For instance, post-contrast imaging of larynx or hypopharynx cancer may assist in differentiating laryngeal cartilage invasion from inflammation, considering that inflamed but non-infiltrated cartilage shows more pronounced and earlier enhancement than that of residual tumor [22]. Figure 4 shows examples of contrast-enhanced images during follow-up of HNCs.

To ensure consistent interpretation and reporting of surveillance imaging, the Neck Imaging Reporting and Data System (NI-RADS) was developed by the American College of Radiology [57], originally for CT but now adapted for MRI [58–61]. The use of GBCAs is essential

for evaluating the primary site and the neck (nodal assessment), which is necessary for assigning all NI-RADS categories [62]. Moreover, according to the NI-RADS authors, contrast-enhanced sequences should be acquired in a minimum of 2 planes, and imaging surveillance could conclude 27 months after treatment if all radiological follow-ups have consistently been negative [54, 59].

PWI has been investigated in three distinct scenarios for patients undergoing treatment for HNCs: early response assessment during chemo-radiotherapy, response evaluation after chemo-radiotherapy or induction chemotherapy, and detection of local tumor recurrence. Regarding treatment response evaluation, tumors showing good perfusion on DCE-MRI following initial therapy (high blood volume and high K^{trans} values) that decline during therapy tend to show better local control [28, 38, 40]. For identifying local tumor recurrence, DCE-MRI may offer greater specificity compared to conventional imaging by quantitatively characterizing the changes in enhancement over time. Residual tumor tissue, in particular, enhances earlier and more intensely than benign post-treatment tissue [28, 38, 40]. This is consistent with the common

Table 3 Key post-contrast imaging features to examine during HNC surveillance

High suspicion of recurrence	
Primary site	<ul style="list-style-type: none"> • Nodule or mass at the primary site, particularly if new or growing, with signal characteristics and enhancement patterns consistent with the original tumor • Perineural tumor spread
Lymph nodes	New or growing lymph node that shows necrosis, irregular margins, or heightened enhancement
Main post-operative findings	
Vascularized scar	Contrast-enhancement that fades over time suggests early vascularized scar tissue, whereas chronic scar tissue is indicated by low signal on T2-weighted imaging
Flap	<ul style="list-style-type: none"> • The muscular component of a myocutaneous flap may show different enhancement patterns • Swollen reactive lymph nodes often appear in the mesenteric fat around a free jejunum flap
Normal tissue remnants	Contrast-enhancement in residual tongue or thyroid tissue could be misinterpreted as a recurrence
Main post-operative complications	
Seroma	Fluid collection without contrast-enhancement
Abscess	Fluid collection with diffusion restriction and rim-enhancement
Flap necrosis	Thrombosis of neck vessels and a non-enhancing flap
Main early changes after radiotherapy/chemoradiotherapy	
Mucositis	Swelling of the pharynx and larynx with contrast-enhancement of their walls
Sialadenitis	Swelling of the submandibular and parotid glands with contrast-enhancement
Thyroiditis	Swelling of the thyroid gland with heterogeneous contrast-enhancement
Main post-radiotherapy complications	
Mucosal necrosis	Absence of mucosal enhancement with or without ulceration
Trismus	Pterygoid muscle atrophy with enhancement confined to the radiation field
Osteoradionecrosis	Abnormal marrow signal and widespread, strong contrast-enhancement, with no associated soft tissue mass. Unenhanced areas within the bone point to sequestrum formation in osteomyelitis
Chondroradionecrosis	Contrast-enhancement of collapsed thyroid cartilage
Vascular complications	Internal jugular vein thrombosis or mural thickening of the carotid artery
Neurological complications	<ul style="list-style-type: none"> • Radiation-induced temporal lobe necrosis appears as a ring-enhancing lesion surrounded by perilesional edema • Cranial and brachial neuropathy are characterized by thickening and enhancement of the affected nerves, and their roots and trunks, respectively • Acute radiation-induced spinal cord injury manifests as an enlarged, T2 hyperintense cord with contrast-enhancement

subjective observation that fibrotic tissue shows delayed and less avid contrast-enhancement on routine scans [63]. Finally, ASL has revealed a significant increase in TBF in patients with recurrent HNCs when compared to those without residual tumor or exhibiting post-radiation changes [35, 64, 65].

In summary, GBCA administration is a necessary component of the multiparametric MRI analysis for individuals under surveillance for HNCs. Patients undergoing long-term MRI follow-up (beyond 27 months) with no clinical or radiological signs of recurrence may be considered for non-contrast scans. The use of GBCA could be reserved for cases where unenhanced sequences show new changes. This conditional suggestion is inferred from the NI-RADS' surveillance algorithm, though it awaits further prospective validation. PWI (with DCE-MRI and/or ASL) has shown encouraging results in evaluating therapy response and in distinguishing between disease recurrence and treatment-related effects.

Future directions

Positron emission tomography (PET)/MRI

Contrast-enhanced MRI and PET should not be viewed as mutually exclusive techniques but rather as complementary tools for the comprehensive evaluation of HNCs [66]. MRI offers superior anatomical resolution, while PET allows for whole-body assessment, making it the most effective tool for detecting nodal involvement, distant metastases, and the potential presence of synchronous tumors [67]. Moreover, PET can demonstrate an absence of tumor activity even before morphological changes are evident on MRI. However, PET can show uptake in various benign processes, including benign tumors, inflammatory conditions, post-traumatic changes, and iatrogenic effects [68]. Hybrid PET/MRI systems have emerged, combining the benefits of both PET and MRI. Nevertheless, the precise role of this integrated technique in the diagnosis, treatment planning, and follow-up of HNCs is still being determined [8]. A 2020 study indicated

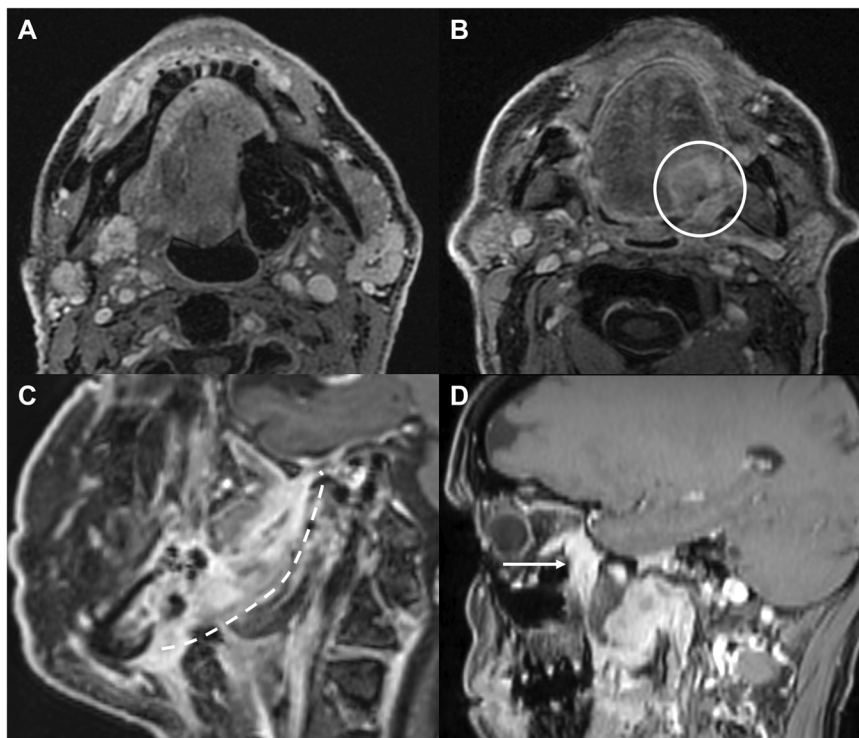


Fig. 4 Examples of findings in T1-weighted images after GBCA administration during follow-up. **A** Left oropharyngeal tumor surgically removed with flap reconstruction, showing no evidence of macroscopic recurrence. **B** Recurrence of a left base tongue tumor (circle). **C** Recurrence of a retromolar trigone tumor along the pterygomandibular raphe (dashed line). **D** Recurrent nasopharyngeal carcinoma reaching the pterygopalatine fossa (arrow) with centrifugal PNS along the infraorbital and greater palatine nerves. GBCA, gadolinium-based contrast agents; PNS, perineural spread

that GBCA administration might not offer additional benefits for local tumor staging in PET/MRI for HNCs. However, the authors recommended retaining post-contrast sequences for salivary gland malignancy work-ups due to their high propensity for nerve invasion, or when PNS is clinically suspected [69].

Novel contrast agents

Considering the concerns about gadolinium deposits and their environmental impact, there is increasing momentum toward developing alternative contrast agents and implementing dose-reduction strategies [11, 12].

Gadopicolenol [70, 71] and gadoquatrane [72–76] are novel macrocyclic GBCA, distinguished by their high relaxivity. They can produce diagnostic images comparable in quality to other GBCAs but with a lower dose (0.05 mmol/kg for gadopicolenol, 0.04 mmol/kg for gadoquatrane). Gadopicolenol was approved by the European Medicines Agency in December 2023 [70, 71], while gadoquatrane was recently submitted to the Food and Drug Administration for approval.

RVP-001 is a novel contrast agent for MRI in clinical development in patients with lesions of the central nervous system. Its mechanism of action harnesses

manganese, an element essential for life and naturally present in the human body, suggesting a superior safety profile compared to gadolinium [77].

Molecular imaging of HNCs employs contrast agents across multiple modalities that bind specific tumor-associated molecular markers, all currently in preclinical development. Examples for MRI include peptide-guided gadolinium compounds (e.g., MT218 targeting extracellular matrix fibronectin, overexpressed in aggressive tumors) for precise localization and characterization; manganese-based probes (e.g., AZA-TA-Mn targeting carbonic anhydrase IX in hypoxic regions) for detailed hypoxia imaging critical to assessing tumor aggressiveness and treatment resistance; and functionalized nanoparticles (e.g., biotin/PEG-UCNPs, single-atom Gd nanospheres, NaHoF₄) [78].

AI tools

AI models, including convolutional neural networks and deep learning algorithms, are showing great promise in interpreting MRI scans for HNC diagnosis [79].

One key application is radiomics, where AI models can extract vast amounts of quantitative data from MRI sequences (like contrast-enhanced T1-weighted images) to predict treatment responses, such as the effectiveness

of induction chemotherapy in nasopharyngeal carcinoma [80, 81]. AI also holds potential for early detection of treatment toxicity, like predicting radiation-induced brain injury before symptoms appear [82]. Despite these exciting advancements, prospective, multicenter validation and standardization of radiomic methods are advised before they can be used routinely in clinics.

AI algorithms can simplify GBCA PWI of HNCs by automating post-processing and initially quantifying parameters, such as those used to differentiate tumor from treatment effects [83].

Finally, machine learning methods have been applied to reduce or eliminate GBCA administration, achieving diagnostic performance comparable to true gadolinium-enhanced scans [84, 85]. Although most work to date has focused on brain, spine, and abdominal imaging, these “virtual contrast” approaches could eventually be adapted for head and neck MRI, obviating or dramatically reducing gadolinium use. Future research should therefore prioritize multicentre validation of AI tools, specifically tailored to the anatomical challenges of HNCs.

In summary, future research needs to investigate PET/MRI to determine in which cases GBCA administration can be avoided. New GBCAs promise to provide the same diagnostic information as macrocyclic agents but at lower doses, while novel gadolinium-free contrast agents are also being explored specifically for use in HNCs. Finally, AI tools could be beneficial in extracting radiomic information, in enhancing the analysis of PWI, and in reducing the dose of GBCA injected.

Conclusion

GBCA-enhanced imaging remains a fundamental component of multiparametric MRI for diagnosis, staging, and monitoring HNCs, typically using a 3D T1-weighted sequence with fat saturation at equilibrium. PWI shows promise for managing HNCs, but it needs further standardization before it can be routinely used in clinical practice. ASL is proving to be an interesting alternative for this purpose, offering the significant benefit of not requiring GBCA administration. Hybrid imaging with PET/MRI, the use of novel contrast agents, and AI tools are all promising options for reducing or eliminating GBCA use. More research is needed to improve these alternative methods and make them more suitable for widespread use in clinics.

Abbreviations

AI	Artificial intelligence
ASL	Arterial spin labeling
CT	Computed tomography
DCE	Dynamic contrast-enhanced
GBCA	Gadolinium-based contrast agent
HNC	Head and neck cancer

MRI	Magnetic resonance imaging
NI-RADS	Neck imaging reporting and data system
PET	Positron emission tomography
PNS	Perineural spread
PWI	Perfusion-weighted imaging
SCC	Squamous cell carcinoma
TBF	Tumor blood flow

Supplementary information

The online version contains supplementary material available at <https://doi.org/10.1007/s00330-025-12165-0>.

Acknowledgements

Marco Parillo, Fabio M. Doniselli, Carlo Cosimo Quattrocchi, and Àlex Rovira are members of the ESMRMB-GREC (European Society for Magnetic Resonance in Medicine and Biology-Gadolinium Research & Education Committee) working group. This is a multidisciplinary group, including clinicians, scientists, chemists, physicists, pathologists, and clinical epidemiologists who all share a common interest in the study of GBCAs in a wide variety of clinical and preclinical conditions. The authors thank all the members for their insights, which were informed by their specific competence and experience. The list of authors includes the radiologist members who developed the project and other external radiologists with expertise on head and neck imaging who revised and approved the final version of the manuscript.

Funding

Open access funding provided by Università degli Studi di Verona within the CRUI-CARE Agreement.

Compliance with ethical standards

Guarantor

The scientific guarantor of this publication is Àlex Rovira.

Conflict of interest

Àlex Rovira serves on scientific advisory boards for Novartis, Sanofi-Genzyme, Synthetic MR, TensorMedical, Roche, Bayer, and Biogen, and has received speaker honoraria from Bayer, Sanofi-Genzyme, Merck-Serono, Teva Pharmaceutical Industries Ltd, Novartis, Roche, Bristol-Myers, and Biogen, and is CMO and co-founder of TensorMedical. Carlo Cosimo Quattrocchi has signed speaker contracts with Bayer Healthcare, Bracco Imaging, and Guerbet. He is co-chair of the ESMRMB-GREC Working Group, whose yearly meetings have received unconditional support from Bayer Healthcare, Bracco Imaging, GE HealthCare, and Guerbet. He is a member of the ESUR-CMSC, whose 2-yearly meetings have received support from Bayer Healthcare, Bracco Imaging, GE HealthCare, and Guerbet. The other authors do not have any conflicts of interest to disclose.

Statistics and biometry

No complex statistical methods were necessary for this paper.

Informed consent

Written informed consent was not required for this study because this is a review.

Ethical approval

Institutional Review Board approval was not required because this is a review.

Methodology

- Narrative review

Author details

¹University of Verona, Verona, Italy. ²Radiology, Multizonal Unit of Rovereto and Arco, APSS Provincia Autonoma Di Trento, Trento, Italy. ³Department of Radiology, Hospital Universitario 12 de Octubre, Madrid, Spain.

⁴Neuroradiology Department, Fondazione IRCCS Istituto Neurologico Carlo Besta, Milan, Italy. ⁵Centre for Medical Sciences-CISMed, University of Trento, Trento, Italy. ⁶Section of Neuroradiology, Department of Radiology, Hospital

Universitari Vall d'Hebron, Autonomous University of Barcelona, Barcelona, Spain.

Received: 7 July 2025 Revised: 10 October 2025 Accepted: 2 November 2025

Published online: 13 December 2025

References

- Machiels J-P, Leemans CR, Golusinski W et al (2020) Squamous cell carcinoma of the oral cavity, larynx, oropharynx and hypopharynx: EHNS-ESMO-ESTRO clinical practice guidelines for diagnosis, treatment and follow-up. *Ann Oncol* 31:1462–1475. <https://doi.org/10.1016/j.annonc.2020.07.011>
- Herpen C van, Poorten W, Skalova A et al (2022) Salivary gland cancer: ESMO-European Reference Network on Rare Adult Solid Cancers (EUR-ACAN) clinical practice guideline for diagnosis, treatment and follow-up. *ESMO Open*. <https://doi.org/10.1016/j.esmoop.2022.100602>
- Bossi P, Chan AT, Licitra L et al (2021) Nasopharyngeal carcinoma: ESMO-EURACAN Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol* 32:452–465. <https://doi.org/10.1016/j.annonc.2020.12.007>
- Resteghini C, Baujat B, Bossi P et al (2025) Sinonasal malignancy: ESMO-EURACAN clinical practice guideline for diagnosis, treatment and follow-up. *ESMO Open*. <https://doi.org/10.1016/j.esmoop.2024.104121>
- Siegel RL, Kratzer TB, Giaquinto AN, Sung H, Jemal A (2025) Cancer statistics, 2025. *CA J Clin* 75:10–45. <https://doi.org/10.3322/caac.21871>
- Gilyoma JM, Rambau PF, Masalu N, Kayange NM, Chalya PL (2015) Head and neck cancers: a clinico-pathological profile and management challenges in a resource-limited setting. *BMC Res Notes* 8:772. <https://doi.org/10.1186/s13104-015-1773-9>
- Junn JC, Soderlund KA, Glastonbury CM (2021) Imaging of head and neck cancer with CT, MRI, and US. *Semin Nucl Med* 51:3–12. <https://doi.org/10.1053/j.semnuclmed.2020.07.005>
- Mukherjee S, Fischbein NJ, Baugnon KL, Policeni BA, Raghaven P (2023) Contemporary imaging and reporting strategies for head and neck cancer: MRI, FDG PET/MRI, NI-RADS, and carcinoma of unknown primary—AJR expert panel narrative review. *AJR Am J Roentgenol* 220:160–172. <https://doi.org/10.2214/AJR.22.28120>
- Widmann G, Henninger B, Kremser C, Jaschke W (2017) MRI sequences in head & neck radiology—state of the art. *Rofo* 189:413–422. <https://doi.org/10.1055/s-0043-103280>
- Endrikat J, Dohanish S, Schleyer N, Schwenke S, Agarwal S, Balzer T (2018) 10 Years of Nephrogenic Systemic Fibrosis: A Comprehensive Analysis of Nephrogenic Systemic Fibrosis Reports Received by a Pharmaceutical Company from 2006 to 2016. *Invest Radiol* 53:541–550. <https://doi.org/10.1097/RLI.0000000000000462>
- van der Molen AJ, Quattrocchi CC, Mallio CA et al (2024) Ten years of gadolinium retention and deposition: ESMO-GREC looks backward and forward. *Eur Radiol* 34:600–611. <https://doi.org/10.1007/s00330-023-10281-3>
- Quattrocchi CC, Parillo M, Spani F et al (2023) Skin thickening of the scalp and high signal intensity of dentate nucleus in multiple sclerosis: association with linear versus macrocyclic gadolinium-based contrast agents administration. *Invest Radiol* 58:223–230. <https://doi.org/10.1097/RLI.0000000000000929>
- EMA (2017) EMA's final opinion confirms restrictions on use of linear gadolinium agents in body scans. https://www.ema.europa.eu/en/documents/referral/gadolinium-article-31-referral-emas-final-opinion-confirms-restrictions-use-linear-gadolinium-agents-body-scans_en.pdf.0. Accessed 16 Aug 2025
- European Society of Urogenital Radiology (2025) ESUR guidelines on contrast agents. https://www.esur.org/wp-content/uploads/2022/03/ESUR-Guidelines-10_0-Final-Version.pdf. Accessed 16 Aug 2025
- Kumasaka S, Kartamihardja AAP, Kumasaka Y, kameo S, Koyama H, Tsushima Y (2024) Anthropogenic gadolinium in the Tone River (Japan): an update showing a 7.7-fold increase from 1996 to 2020. *Eur Radiol Exp* 8:64. <https://doi.org/10.1186/s41747-024-00460-2>
- Lenkinski RE, Rofsky NM (2024) Contrast media-driven anthropogenic gadolinium: knowns and unknowns. *Radiology* 311:e240020. <https://doi.org/10.1148/radiol.240020>
- Ognard J, Barrat J-A, Cotton F et al (2021) A roadmap towards pollution prevention and sustainable development of Gadolinium. *J Neuroradiol* 48:409–411. <https://doi.org/10.1016/j.neurad.2021.08.002>
- Rovira À, Ben Salem D, Geraldo AF et al (2024) Go green in neuroradiology: towards reducing the environmental impact of its practice. *Neuroradiology* 66:463–476. <https://doi.org/10.1007/s00234-024-03305-2>
- Rovira À, Quattrocchi CC (2024) Safe and optimized use of gadolinium-based contrast agents in neuroimaging. *Eur Radiol* 34:4567–4569. <https://doi.org/10.1007/s00330-023-10456-y>
- Rovira À, Doniselli FM, Auger C et al (2024) Use of gadolinium-based contrast agents in multiple sclerosis: a review by the ESMO-GREC and ESNR Multiple Sclerosis Working Group. *Eur Radiol* 34:1726–1735. <https://doi.org/10.1007/s00330-023-10151-y>
- Quattrocchi CC, Rovira À, van der Molen AJ, Mallio CA (2024) ESR essentials: gadolinium-wise MRI—practice recommendations by the European Society for Magnetic Resonance in Medicine and Biology. *Eur Radiol*. <https://doi.org/10.1007/s00330-024-11214-4>
- Doniselli FM, Ramos J, Hilario A et al (2025) Recommendations on the use of gadolinium-based contrast agents in the diagnosis and monitoring of common adult intracranial tumours. *Eur Radiol*. <https://doi.org/10.1007/s00330-025-11646-6>
- Parillo M, Vertulli D, Vaccarino F, Mallio CA, Zobel BB, Quattrocchi CC (2024) The sensitivity of MIPs of 3D contrast-enhanced VIBE T1-weighted imaging for the detection of small brain metastases (≤ 5 mm) on 1.5 tesla MRI. *Neuroradiol J* 37:744–750. <https://doi.org/10.1177/19714009241260802>
- Qu J, Su T, Pan B et al (2023) Free-breathing StarVIBE sequence for the detection of extranodal extension in head and neck cancer: an image quality and diagnostic performance study. *Cancers (Basel)* 15:4992. <https://doi.org/10.3390/cancers15204992>
- Kataoka M, Ueda H, Koyama T et al (2005) Contrast-enhanced volumetric interpolated breath-hold examination compared with spin-echo T1-weighted imaging of head and neck tumors. *AJR Am J Roentgenol* 184:313–319. <https://doi.org/10.2214/ajr.184.1.01840313>
- Hur S-J, Choi Y, Yoon J et al (2021) Intraindividual comparison between the contrast-enhanced golden-angle radial sparse parallel sequence and the conventional fat-suppressed contrast-enhanced T1-weighted spin-echo sequence for head and neck MRI. *AJNR Am J Neuroradiol* 42:2009–2015. <https://doi.org/10.3174/ajnr.A7285>
- Gaddikeri S, Gaddikeri RS, Tailor T, Anzai Y (2016) Dynamic contrast-enhanced MR imaging in head and neck cancer: techniques and clinical applications. *AJNR Am J Neuroradiol* 37:588–595. <https://doi.org/10.3174/ajnr.A4458>
- El Beltagi AH, Elstouhy AH, Own AM, Abdelfattah W, Nair K, Vattoto S (2019) Functional magnetic resonance imaging of head and neck cancer: performance and potential. *Neuroradiol J* 32:36–52. <https://doi.org/10.1177/1971400918808546>
- Martin-Noguerol T, Kirsch CFE, Montesinos P, Luna A (2021) Arterial spin labeling for head and neck lesion assessment: technical adjustments and clinical applications. *Neuroradiology* 63:1969–1983. <https://doi.org/10.1007/s00234-021-02772-1>
- Tanaka F, Umino M, Maeda M et al (2022) Pseudocontinuous arterial spin labeling: clinical applications and usefulness in head and neck entities. *Cancers (Basel)* 14:3872. <https://doi.org/10.3390/cancers14163872>
- Alsop DC, Detre JA, Golay X et al (2015) Recommended implementation of arterial spin-labeled perfusion MRI for clinical applications: a consensus of the ISMRM perfusion study group and the European consortium for ASL in dementia. *Magn Reson Med* 73:102–116. <https://doi.org/10.1002/mrm.25197>
- Abdel Razek AAK, Talaat M, El-Serougy L, Gaballa G, Abdelsalam M (2019) Clinical Applications of Arterial Spin Labeling in Brain Tumors. *J Comput Assist Tomogr* 43:525–532. <https://doi.org/10.1097/RCT.0000000000000873>
- Fallatah SM, Pizzini FB, Gomez-Anson B et al (2018) A visual quality control scale for clinical arterial spin labeling images. *Eur Radiol Exp* 2:45. <https://doi.org/10.1186/s41747-018-0073-2>
- Abdel Razek AAK, Nada N (2018) Arterial spin labeling perfusion-weighted MR imaging: correlation of tumor blood flow with pathological degree of tumor differentiation, clinical stage and nodal metastasis of head and neck squamous cell carcinoma. *Eur Arch Otorhinolaryngol* 275:1301–1307. <https://doi.org/10.1007/s00405-018-4950-3>

35. Fujima N, Yoshida D, Sakashita T et al (2016) Usefulness of pseudo-continuous arterial spin-labeling for the assessment of patients with head and neck squamous cell carcinoma by measuring tumor blood flow in the pretreatment and early treatment period. *AJR Am J Neuroradiol* 37:342–348. <https://doi.org/10.3174/ajnr.A4513>
36. Chowdhury R, Turkdogan S, Alsayegh R et al (2024) Comprehensive diagnostic approach to head and neck masses. *J Otorhinolaryngol Hear Bal Med* 5:17. <https://doi.org/10.3390/ohbm5020017>
37. Pulickal GG, Navaratnam AV, Nguyen T, Dragan AD, Dziedzic M, Lingam RK (2018) Imaging sinonasal disease with MRI: providing insight over and above CT. *Eur J Radiol* 102:157–168. <https://doi.org/10.1016/j.ejrad.2018.02.033>
38. Kabadi SJ, Fatterpekar GM, Anzai Y, Mogen J, Hagiwara M, Patel SH (2018) Dynamic contrast-enhanced MR imaging in head and neck cancer. *Magn Reson Imaging Clin N Am* 26:135–149. <https://doi.org/10.1016/j.mric.2017.08.008>
39. Coudert H, Mirafzal S, Dissard A, Boyer L, Montoriol P-F (2021) Multiparametric magnetic resonance imaging of parotid tumors: a systematic review. *Diagn Interv Imaging* 102:121–130. <https://doi.org/10.1016/j.diii.2020.08.002>
40. Romeo V, Stanzione A, Ugga L et al (2022) Clinical indications and acquisition protocol for the use of dynamic contrast-enhanced MRI in head and neck cancer squamous cell carcinoma: recommendations from an expert panel. *Insights Imaging* 13:198. <https://doi.org/10.1186/s13244-022-01317-1>
41. Homer JJ, Winter SC, Abbey EC et al (2024) Head and neck cancer: United Kingdom National Multidisciplinary Guidelines, sixth edition. *J Laryngol Otol* 138:S1–S224. <https://doi.org/10.1017/S0022215123001615>
42. Becker M, Monnier Y, de Vito C (2022) MR imaging of laryngeal and hypopharyngeal cancer. *Magn Reson Imaging Clin N Am* 30:53–72. <https://doi.org/10.1016/j.mric.2021.08.002>
43. Ong CK, Chong VF-H (2010) Imaging of perineural spread in head and neck tumours. *Cancer Imaging* 10:S92–S98. <https://doi.org/10.1102/1470-7330.2010.9033>
44. Abdullaeva U, Pape B, Hirvonen J (2024) Diagnostic accuracy of MRI in detecting the perineural spread of head and neck tumors: a systematic review and meta-analysis. *Diagnostics (Basel)* 14:113. <https://doi.org/10.3390/diagnostics14010113>
45. Kang H, Kennedy TA, Yu E (2024) Head and neck squamous cell cancer: approach to staging and surveillance. In: Hodler J, Kubik-Huch RA, Roos JE (eds) *Diseases of the brain, head and neck, spine 2024–2027: diagnostic imaging*. Springer Nature Switzerland, Cham, pp 251–264
46. Weimar EAM, Huang SH, Lu L et al (2018) Radiologic-pathologic correlation of tumor thickness and its prognostic importance in squamous cell carcinoma of the oral cavity: implications for the eighth edition tumor, node, metastasis classification. *AJNR Am J Neuroradiol* 39:1896–1902. <https://doi.org/10.3174/ajnr.A5782>
47. Haraguchi K, Yoshida D, Oda M et al (2021) Depth of invasion determined by magnetic resonance imaging in tongue cancer can be a predictor of cervical lymph node metastasis. *Oral Surg Oral Med Oral Pathol Oral Radiol* 131:231–240. <https://doi.org/10.1016/j.oooo.2020.07.005>
48. Zanon DK, Patel SG, Shah JP (2019) Changes in the 8th Edition of the American Joint Committee on Cancer (AJCC) staging of head and neck cancer: rationale and implications. *Curr Oncol Rep* 21: 52. <https://doi.org/10.1007/s11912-019-0799-x>
49. Pan J-J, Mai H-Q, Ng WT et al (2024) Ninth version of the AJCC and UICC nasopharyngeal cancer TNM staging classification. *JAMA Oncol* 10:1627–1635. <https://doi.org/10.1001/jamaoncol.2024.4354>
50. Goldenberg D, Begum S, Westra WH et al (2008) Cystic lymph node metastasis in patients with head and neck cancer: an HPV-associated phenomenon. *Head Neck* 30:898–903. <https://doi.org/10.1002/hed.20796>
51. Henson C, Abou-Foul AK, Yu E et al (2024) Criteria for the diagnosis of extranodal extension detected on radiological imaging in head and neck cancer: Head and Neck Cancer International Group consensus recommendations. *Lancet Oncol* 25:e297–e307. [https://doi.org/10.1016/S1470-2045\(24\)00066-4](https://doi.org/10.1016/S1470-2045(24)00066-4)
52. Fischbein NJ, Noworolski SM, Henry RG, Kaplan MJ, Dillon WP, Nelson SJ (2003) Assessment of metastatic cervical adenopathy using dynamic contrast-enhanced MR imaging. *AJNR Am J Neuroradiol* 24:301–311
53. Hiyama T, Miyasaka Y, Kuno H et al (2024) Posttreatment head and neck cancer imaging: anatomic considerations based on cancer subsites. *Radiographics* 44:e230099. <https://doi.org/10.1148/rg.230099>
54. Aiken AH, Hudgins PA (2018) Neck imaging reporting and data system. *Magn Reson Imaging Clin N Am* 26:51–62. <https://doi.org/10.1016/j.mric.2017.08.004>
55. Chakrabarty N, Mahajan A, Agrawal A, Prabhaskar K, D'Cruz AK (2024) Comprehensive review of post-treatment imaging in head and neck cancers: from expected to unexpected and beyond. *Br J Radiol* 97:1898–1914. <https://doi.org/10.1093/bjr/tqae207>
56. Saito N, Nadgir RN, Nakahira M et al (2012) Posttreatment CT and MR imaging in head and neck cancer: what the radiologist needs to know. *Radiographics* 32:1261–1282. <https://doi.org/10.1148/rg.325115160> discussion 1282–1284
57. Aiken AH, Farley A, Baugnon KL et al (2016) Implementation of a novel surveillance template for head and neck cancer: neck imaging reporting and data system (NI-RADS). *J Am Coll Radiol* 13:743–746.e1. <https://doi.org/10.1016/j.jacr.2015.09.032>
58. American College of Radiology Committee on NI-RADS™ (2018) ACR neck imaging reporting and data systems (NI-RADS™): a white paper of the ACR NI-RADS Committee. <https://edge.sitecorecloud.io/americancoldf5f-acrorgf92a-productioncb02-3650/media/ACR/Files/RADS/NI-RADS/NIRADS-MRI-2025-Assessment-Categories.pdf>. Accessed 16 Aug 2025
59. Bunch PM, Aiken AH, Baugnon KL et al (2025) ACR neck imaging reporting and data system (NI-RADS) for magnetic resonance imaging v2025. *J Am Coll Radiol* S1546-1440:00442–00449. <https://doi.org/10.1016/j.jacr.2025.07.023>
60. Vertulli D, Parillo M, Mallio CA (2025) The role of neck imaging reporting and data system (NI-RADS) in the management of head and neck cancers. *Bioengineering* 12:398. <https://doi.org/10.3390/bioengineering12040398>
61. Falzone A, Parillo M, Neri M et al (2025) Interrater reliability of MRI neck imaging reporting and data system (NI-RADS) in the follow-up of nasopharyngeal carcinoma after radiation therapy. *Radiol Med*. <https://doi.org/10.1007/s11547-025-01982-4>
62. Parillo M, Mallio CA, Van der Molen AJ et al (2024) The role of gadolinium-based contrast agents in magnetic resonance imaging structured reporting and data systems (RADS). *MAGMA* 37:15–25. <https://doi.org/10.1007/s10334-023-01113-y>
63. Parillo M, Mallio CA, Dekkers IA et al (2024) Late/delayed gadolinium enhancement in MRI after intravenous administration of extracellular gadolinium-based contrast agents: Is it worth waiting?. *MAGMA* 37:151–168. <https://doi.org/10.1007/s10334-024-01151-0>
64. Razeq AAKA (2018) Arterial spin labelling and diffusion-weighted magnetic resonance imaging in differentiation of recurrent head and neck cancer from post-radiation changes. *J Laryngol Otol* 132:923–928. <https://doi.org/10.1017/S0022215118001743>
65. Fujima N, Kudo K, Yoshida D et al (2014) Arterial spin labeling to determine tumor viability in head and neck cancer before and after treatment. *J Magn Reson Imaging* 40:920–928. <https://doi.org/10.1002/jmri.24421>
66. Becker M, Zaidi H (2014) Imaging in head and neck squamous cell carcinoma: the potential role of PET/MRI. *Br J Radiol* 87:20130677. <https://doi.org/10.1259/bjr.20130677>
67. Alsibani A, Alqahtani A, Almohammadi R et al (2024) Comparing the efficacy of CT, MRI, PET-CT, and US in the detection of cervical lymph node metastases in head and neck squamous cell carcinoma with clinically negative neck lymph node: a systematic review and meta-analysis. *J Clin Med* 13:7622. <https://doi.org/10.3390/jcm13247622>
68. Purohit BS, Ailianou A, Dulguerov N, Becker CD, Ratib O, Becker M (2014) FDG-PET/CT pitfalls in oncological head and neck imaging. *Insights Imaging* 5:585–602. <https://doi.org/10.1007/s13244-014-0349-x>
69. Pyatigorskaya N, De Laroche R, Bera G et al (2020) Are gadolinium-enhanced MR sequences needed in simultaneous 18F-FDG-PET/MRI for tumor delineation in head and neck cancer?. *AJNR Am J Neuroradiol* 41:1888–1896. <https://doi.org/10.3174/ajnr.A6764>
70. European Medicines Agency (EMA) (2023) Elucirem. <https://www.ema.europa.eu/en/medicines/human/EPAR/elucirem>. Accessed 11 May 2025
71. European Medicines Agency (EMA) (2023) Vueway. <https://www.ema.europa.eu/en/medicines/human/EPAR/vueway>. Accessed 11 May 2025

72. Lohrke J, Berger M, Frenzel T et al (2022) Preclinical profile of gadoquatrane: a novel tetrameric, macrocyclic high relaxivity gadolinium-based contrast agent. *Invest Radiol* 57:629–638. <https://doi.org/10.1097/RLI.0000000000000889>
73. Hofmann BM, Riecke K, Klein S et al (2024) Pharmacokinetics, safety, and tolerability of the novel tetrameric, high-relaxivity, macrocyclic gadolinium-based contrast agent gadoquatrane in healthy adults. *Invest Radiol* 59:140–149. <https://doi.org/10.1097/RLI.0000000000001043>
74. Bayer (2024) A multicenter, prospective, open-label study to evaluate the pharmacokinetics and safety of gadoquatrane in pediatric participants (from birth to <18 years) undergoing contrast-enhanced magnetic resonance imaging (CE-MRI). <https://clinicaltrials.gov/study/NCT05915026>. Accessed 12 May 2025
75. Bayer (2025) A study to compare how well gadoquatrane works and its safety with an already available contrast agent for MRI in people with any known or suspected problems of the body (except brain or spinal cord-related problems) (Quanti OBR). <https://clinicaltrials.gov/study/NCT05915728>. Accessed 12 May 2025
76. Bayer (2025) A study to compare how well gadoquatrane works and its safety with an already available contrast agent for MRI in people with known or suspected brain or spinal cord-related problems (Quanti CNS). <https://clinicaltrials.gov/study/NCT05915702>. Accessed 12 May 2025
77. Reveal Pharmaceuticals Inc. (2025) First in Human Study of RVP-001, a new manganese based MRI contrast agent. <https://clinicaltrials.gov/study/NCT05413668>. Accessed 12 May 2025
78. Neillio JM, Ginat DT (2024) Emerging head and neck tumor targeting contrast agents for the purpose of CT, MRI, and multimodal diagnostic imaging: a molecular review. *Diagnostics* (Basel) 14:1666. <https://doi.org/10.3390/diagnostics14151666>
79. Broggi G, Maniaci A, Lentini M et al (2024) Artificial intelligence in head and neck cancer diagnosis: a comprehensive review with emphasis on radiomics, histopathological, and molecular applications. *Cancers* (Basel) 16:3623. <https://doi.org/10.3390/cancers16213623>
80. Wang G, He L, Yuan C, Huang Y, Liu Z, Liang C (2018) Pretreatment MR imaging radiomics signatures for response prediction to induction chemotherapy in patients with nasopharyngeal carcinoma. *Eur J Radiol* 98:100–106. <https://doi.org/10.1016/j.ejrad.2017.11.007>
81. Zhao L, Gong J, Xi Y et al (2020) MRI-based radiomics nomogram may predict the response to induction chemotherapy and survival in locally advanced nasopharyngeal carcinoma. *Eur Radiol* 30:537–546. <https://doi.org/10.1007/s00330-019-06211-x>
82. Zhang B, Lian Z, Zhong L et al (2020) Machine-learning based MRI radiomics models for early detection of radiation-induced brain injury in nasopharyngeal carcinoma. *BMC Cancer* 20:502. <https://doi.org/10.1186/s12885-020-06957-4>
83. Bruixola G, Remacha E, Jiménez-Pastor A et al (2021) Radiomics and radiogenomics in head and neck squamous cell carcinoma: Potential contribution to patient management and challenges. *Cancer Treat Rev* 99:102263. <https://doi.org/10.1016/j.ctrv.2021.102263>
84. Mallio CA, Radbruch A, Deike-Hofmann K et al (2023) Artificial intelligence to reduce or eliminate the need for gadolinium-based contrast agents in brain and cardiac MRI: a literature review. *Invest Radiol* 58:746–753. <https://doi.org/10.1097/RLI.0000000000000983>
85. Tsui B, Calabrese E, Zaharchuk G, Rauschecker AM (2024) Reducing gadolinium contrast with artificial intelligence. *J Magn Reson Imaging* 60:848–859. <https://doi.org/10.1002/jmri.29095>

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.