

1 **Role of fruit flesh cell morphology and MdPG1 allelotype in**
2 **influencing juiciness and texture properties in apple.**

3 Poles L^{a‡}, Gentile A^{bc‡}, Giuffrida A^b, Valentini L^d, Endrizzi I^e, Aprea E^e, Gasperi F^e, Distefano G^b, Artioli
4 G^d, La Malfa S^b, Costa F^{e,f}, Lovatti L^a, Di Guardo M^b

5 ^a Innovation Fruit Consortium (CIF), via Mach 1, 38010 San Michele all'Adige, Trento, Italy

6 ^b Dipartimento di Agricoltura, Alimentazione e Ambiente, University of Catania, via Valdisavoia 5, 95123,
7 Catania, Italy

8 ^c National Center for Citrus Improvement, College of Horticulture and Landscape, Hunan Agricultural
9 University, Changsha, China

10 ^d Department of Geosciences, University of Padua, Via Gradenigo 6, 35131 Padua, Italy

11 ^e Center Agriculture Food Environment University of Trento, Via E. Mach 1, 38010 San Michele all'Adige, Italy

12 ^f Research and Innovation Centre, Fondazione Edmund Mach, San Michele all' Adige, Trento, Italy

13 ‡: these authors equally contributed to this work

14

15 Keywords: X-ray computed micro-tomography; *Malus domestica*; sensory analysis; cellular shape; cellular
16 size; molecular marker

17

18 Corresponding author: mario.diguardo@unict.it;

19

20

21 ABSTRACT

22 Apple fruit quality is strongly influenced by the interplay between juiciness and texture. To better decipher
23 the complexity underneath the control of such quality traits, a multidisciplinary approach combining the
24 mechanic and acoustic profiling of texture, juice analysis, cell morphology, sensory and genetic analysis was
25 carried out. The analyses were conducted after 1.5 months of cold storage on fourteen accessions employed
26 in novel breeding schemes for texture and juiciness. The food matrix structure was exploited focusing on
27 both the cell morphology (employing an optical microscope) and the intercellular space (using an X-ray
28 computed micro-tomography scanner). The mechanical and acoustic properties of texture were profiled with
29 a texture analyser, while the juice was extracted using a mechanical press. In parallel to the analytical
30 assessments, fruit texture, juiciness and flavour were also evaluated by sensory analysis. The results
31 highlighted a positive correlation between cell shape and the intercellular volume. Apple accessions
32 distinguished by round cells were characterized by a reduced intercellular space, while cell with an angular
33 cell shape had a higher intercellular space. While the cell shape was associated with juiciness, the firmness
34 response was more influenced by cell size. The interplay between cellular morphology and juiciness was also
35 investigated together with the allelotype variability of a genetic marker designed for *MdPG1*, a
36 polygalacturonase gene known to control the regulation of fruit texture in apple. The highest juiciness was
37 found in apples with both a high fraction of round cells and the presence of the *MdPG1* allele associated with
38 low softening rates. The elucidation of the role of cellular morphology in the control of fruit texture and
39 juiciness, and their association with the *MdPG1* alleles, provided valuable information for a more detailed
40 and informative analysis of fruit quality, enabling a more precise characterization and selection of superior
41 apple accessions.

42

43

44

45

46

47 1 INTRODUCTION

48 Fruit quality is affected by appearance, texture, juiciness and nutritional attributes (Abbott, 1999; Cappellin
49 et al., 2015; Corollaro et al., 2014b; Endrizzi et al., 2015). Juiciness and texture have dominant roles in the
50 determination of fruit quality in apple, since these are the two most appreciated characteristics by consumers
51 (Dailliant-Spinnler et al., 1996; King et al., 2000; Bourne, 2002). The relative importance of each quality trait
52 component varies greatly among species. These features develop and change during fruit development and
53 ripening processes rendering the fruit desirable and physiologically prone to seed dispersion (Giovannoni,
54 2004). In apple, the most important changes after colour are fruit softening and increased juiciness. The loss
55 of firmness relies on the depolymerisation of the middle lamellae (a pectin-rich layer adhering cells)-cell wall
56 architectural structure by the action of several cell wall modifying enzymes (CWME) (Brummell and Harpster,
57 2001). The different types of texture among apple cultivars, depend on the different genetically programmed
58 dismantling events of this polysaccharide structure (King et al., 2000; Waldron et al., 2003). While dry and
59 mealy texture is related to high rate of middle lamellae-cell wall depolymerisation, firm and crispy fruit types
60 are associated with more structured integrity of the cell wall (Longhi et al., 2013b). Fruit texture and juiciness
61 are also related with the types of cell morphology, as documented in tomato (*Solanum lycopersicum*) (Bertin
62 et al., 2001), sweet cherry (*Prunus avium*) (Olmstead et al., 2007) and peach (*Prunus persica*) (Quilot and
63 Genard, 2008). In apple, a direct correlation between cell size, firmness and juiciness was observed in
64 cultivars showing larger cells, which were furthermore characterized by higher levels of juiciness and texture
65 (Allan-Wojtas et al., 2003; Mann et al., 2005; McAtee et al., 2009; Ng et al., 2013). Other work suggests a
66 more complex mechanism of texture and juiciness regulation in which the size and numbers of cells do not
67 have any effect on such quality traits (Charles et al., 2018). To investigate the role of cell morphology in
68 controlling fruit quality aspects, several approaches have been employed, such as light microscopy
69 (Schotsmans et al., 2004; McAtee et al., 2009), scanning electron microscopy (SEM) (Seymour et al., 2002;
70 McAtee et al., 2009; Ng et al., 2013) and X-ray computed micro-tomography scanner (Mendoza et al., 2007;
71 Ting et al., 2013).

72 Juiciness is the amount of juice released by cell during the breakdown of the cell wall through mechanical
73 compression, and its perception is generally associated with fruit freshness (Corollaro et al., 2014a; Harker
74 et al., 2003). Juiciness can be directly measured as the amount of juice released during either mechanical
75 compression (Corollaro et al., 2014a) or homogenization (Chen and Borgic, 1985; Lill and Mespel, 1988) or
76 indirectly, through the measurement of the juice absorption by a tissue paper, or the weighing of a portion
77 of fruit flesh forced through a Lauer syringe before and after centrifugation (Harker et al., 1997). Fruit texture,
78 is instead considered a multi-trait feature, involving the interplay between mechanical and acoustic
79 components (Costa et al., 2011). Fruit texture has been investigated using texture analysers (Costa et al.,
80 2012). Sensory analysis indicate that fruit texture and juiciness are tightly correlated (Corollaro et al., 2014;
81 Nybom et al., 2003) explaining why texture is often employed as an indirect indicator of juiciness (Allan-
82 Wojtas et al., 2003). Fruit texture and juiciness can be also strongly affected by storage conditions (Corollaro
83 et al., 2013). During storage, in fact, important loss of fruit firmness can occur, severely limiting the storability
84 of particular cultivar and consequently their marketability. For this reason, texture and juiciness are
85 considered as fundamental traits in breeding programs for cultivars with superior fruit quality attributes. The
86 selection process for firmness can now be assisted by molecular markers associated with these traits. For the
87 advanced DNA-informed selection of texture in apple, a gene-based marker related to *MdPG1* has been
88 developed and validated (Longhi et al., 2012, 2013a; Farneti et al., 2017; Di Guardo et al., 2017). This gene is
89 a member of the polygalacturonase family located in chromosome 10 of the apple genome, known to play a
90 pivotal role in the cell wall complex dismantling process (Wakasa et al., 2006).

91 The objective of this study was to investigate the interactions between cell morphology and texture-juiciness
92 properties, including the *MdPG1* marker allelotype, in fruit of fourteen apple cultivars and selections.

93

94 2 MATERIALS AND METHODS

95 2.1 Plant material

96 The cultivars and advanced selections used in this study are listed in Table 1 together with their pedigree
 97 information. We used cultivars with a history of cultivation and commercialization (e.g. 'Golden Delicious',
 98 'Fuji') and novel accessions chosen for their superior characteristics in terms of fruit quality, production and
 99 storability. Five of the accessions included in the list were also used as parental cultivars. Plants were grown
 100 in two experimental orchards of the Fruit Innovation Consortium (CIF) located in the province of Trento
 101 (north of Italy) and maintained following standard pruning and agronomical practices. Fruit were harvested
 102 at the commercial ripening stage, established through the assessment of the starch content on 20 fruit per
 103 accession with the Lugol's test (mean value of 7 on the Starch Conversion Chart, CTIFL, Paris, France). For
 104 each accession, a minimum of 50 fruits with homogeneous size were collected and stored in air for 1.5
 105 months at 2 °C and 90% relative humidity). After storage, fruit were maintained at room temperature for 24
 106 hours prior the destructive analysis.

107

108 Table 1: Cultivars and selections employed in the study. For each accession the parentage is specified; if
 109 available, the name of the clone is reported in parentheses next to the cultivar name. The genetic
 110 configuration at the MDPG1_{SSR}10Kd locus is reported in the last column both specifying the size of the
 111 amplicons (bp) and the favourable (A) or unfavourable (a) allelic effect on texture.

Accession	Maternal line	Paternal line	SSR-PG
CIV323	Royal Gala	A3-7	313-317 (AA)
FEM16	Cripps Pink	Caudle	317-317 (AA)
Fuji (Fubrax)	Red Delicious	Ralls Janet	313-313 (AA)
Golden Delicious (Clone B)	Grimes Golden	OP	317-324 (Aa)
Gradisca	Fuji	Cripps Pink	317-317 (AA)
Kizuri	Golden Delicious	NY75413-30	313-317 (AA)
Lumaga	Resy	Delbard Jubilee	317-317 (AA)
Minneiska	Honeycrisp	Minnewashta	313-317 (AA)
MN55	Honeycrisp	MonArk	317-317 (AA)
Red Delicious (Jeromine)	NA	NA	313-324 (Aa)
Royal Gala (Baigent)	Kidd's Orange Red	Golden Delicious	317-324 (Aa)
UEB32642	Golden Delicious	Topaz	317-317 (AA)
UEB6581	Fuji	UEB32642	313-317 (AA)
Y102	Golden Delicious	SJ109	317-324 (Aa)

112

113 2.2 Phenotyping of the apple juiciness

114 For each accession, 10 fruit were used to measure the extractable juice following the protocol described by
115 Corollaro et al. (2014). For each fruit, three disks were isolated from different sides of the fruit
116 (Supplementary Table 1). Extractable juice ('juiciness') was assessed by weighting the liquid expressed from
117 mechanical compression of the disks.

118

119 2.3 Texture profiling analysis

120 Phenotyping of fruit texture was carried-out with a TA-XT texture analyzer (Stable MicroSystem Ltd.,
121 Godalming, UK) equipped with an acoustic envelope device (AED) as described by Costa et al. (2011).
122 Mechanical measurements were carried out with a 4 mm flat probe, at a speed of 100 mm min⁻¹ and an auto-
123 force trigger at 5 g. The AED connected to the instrument allowed the simultaneous assessment of the
124 acoustic response of the sample during fracturing. Texture properties were measured on three disks/fruit on
125 a minimum of five apples per accession, (Supplementary Table 1). For the combined mechanical-acoustic
126 textural profile, 12 parameters were digitally identified through the use of an *ad hoc* macro (Table 2), as
127 detailed by Costa et al. (2011, 2012).

128

129 2.4 Cell isolation and morphological analysis

130 Three fruits for each accession were selected for cell extraction following the protocol described by McAtee
131 et al. (2009). For each apple, a block with a volume of approximately 1 cm³ was sampled from the central
132 portion of the fruit cortex, avoiding the cells (normally smaller) located in the proximity of the peel (Allan-
133 Wojtas et al., 2003) (Supplementary Table 1). Sampling was accurately performed by selecting disks at the
134 same depth of the fruit cortex, but the side (sunny and shaded) of the fruit was not taken into account as it
135 does not affect cell size (McAtee et al., 2009). The initial block was further sectioned into smaller cubes of
136 approximately 2 mm³ using a fine edge scalpel and gently boiled for 25 minutes in a 40 mL solution of 0.05
137 M Na₂CO₃ in 0.3 M mannitol (McAtee et al., 2009). Mannitol was added to the solution to stabilize the osmotic

138 pressure, inside and outside the cells, while Na₂CO₃ was added to solubilize the pectin matrix, facilitating the
139 cell separation. This procedure resulted in the dismantling of the ordered cell wall structures while preserving
140 cell integrity and shape. After boiling, single cells formed a homogenate in which cells were suspended in the
141 solution allowing a direct observation or after storage at 4 °C. Cell observation was carried out using a Leica
142 DM 2500 optical microscope (Heidelbergh, Germany) equipped with a Leica DFC 320 digital camera and a
143 10x magnifying glass under bright field. Cell images were elaborated and analysed using the Leica application
144 suite software (v. 2.5.0). Each cell contour was manually highlighted enabling the software to automatically
145 compute the corresponding cell area (CA), perimeter, height and width. Cellular shape (CS) was instead
146 indirectly determined through the analysis of the ratio between height and width, following the assumption
147 that more similar these two parameters are, more round the shape of the cell will be, as already proposed
148 by Smedt et al. (1998). Both raw and processed photos are available upon request.

149

150 **2.5 DNA extraction and analysis of *MdPG1***

151 For each accession included in this survey, total genomic DNA was isolated from young leaf tissue using the
152 Qiagen DNeasy Plant mini kit (Qiagen) following the manufacturer's protocol. DNA quality was assessed using
153 a Nanodrop ND-8000[®] spectrophotometer (Thermo Scientific, USA). For each sample, the genetic
154 configuration at the *MdPG1*_{SSR}10kd (the molecular marker designed to target the *MdPG1* gene) locus was
155 assessed using the primer sequences and PCR conditions described by Longhi et al. (2013a). *MdPG1*_{SSR}10kd
156 was amplified using the following pair of primers: forward:_5'-50-TTCCTCCTGGGTTTTTGG-3' and
157 reverse_5'-ACTCGTGCGCCAGATAGC-3' The PCR amplification thermal conditions were: 94 °C for 2 min, 32
158 cycles of 94 °C for 30 s, 58 °C for 30 s and 72 °C for 45 s, followed by a final extension of 72 °C for 5 min. PCR
159 products were separated using an ABI Prism 3730 capillary sequencer (Applied Biosystem by Life
160 Technologies) while the size of the amplicons was assessed using the GeneMapper[®] software (Applied
161 Biosystem by Life Technologies).

162

163 2.6 X-ray computed micro-tomography

164 3D imaging was carried out on a restricted dataset encompassing the two MDPG1_{SSR}10Kd genotypes in
165 analysis and, within each genetic class. 'Fuji', 'Kizuri', 'Gala', 'Golden Delicious', 'Gradisca', 'Lumaga' and
166 'Minneiska' were selected on the basis of their divergent juiciness and texture responses. The measurements
167 were performed using a Skyscan 1172 micro-CT scanner. Cylindrical cores of approximately 6 mm diameter
168 and 20 mm height were extracted and mounted on a plastic sample holder (Supplementary Table 1). The
169 samples were irradiated using a cone-shaped X-ray beam (W source) with 44 kV voltage and 222 μ A current.
170 Each projection was acquired with an exposure time of 265 ms and an angular step of 0.3°. Tomographic
171 reconstruction was performed using the FDK algorithm (Feldkamp et al., 1984). After this latter step, the
172 dataset associated with each cultivar consisted of a sequence of 1,200 cross sections having a pixel size of 5
173 μ m. These images were cropped at a final diameter of 4 mm in order to remove any possible artefact deriving
174 from sample extraction. Binary images were obtained using an iterative thresholding method (Ridler and
175 Calvard, 1978) and analysed using the Fiji software (Schindelin et al., 2012) to measure the volume of the
176 individual intercellular spaces (IS).

177

178 2.7 Sensory analysis

179 The sensory profiles of apples were assessed using the descriptive analysis method with a consensus lexicon
180 of 34 sensory descriptors developed by Corollaro et al. (2013). We considered nine descriptors related to
181 texture, juiciness aroma and flavour. The latter two were included in the analysis in light of their relationship
182 with texture and juiciness characteristics (Supplementary Table 2). The intensity of each descriptor was
183 expressed as a score on a 100 mm linear scale, ranging from 0 (absence) to 100 (extremely intense), and with
184 50 as middle point. The analyses were carried out by a trained panel composed of 15 panellists (8 males and
185 7 females) who had between 2 and 7 years of experience in descriptive analysis of apple fruit quality aspects.
186 Fruit were analysed in five panel sessions according to the different harvest dates. Panellists evaluated 3
187 apple samples per session in duplicate according to a balanced order of presentation over the panel. Each

188 panellist received eight apple disks per apple sample: flesh cylinders (1.8 cm diameter; ± 2.5 g) cut from 8
189 different fruit (Supplementary Table 1), treated with an antioxidant solution (0.2 % citric acid, 0.2 % ascorbic
190 acid, 0.5 % calcium chloride), and presented in a clear plastic cup encoded with a random three-digit code.
191 The panel evaluated the samples under red light (to avoid bias due to the external appearance of the sample)
192 in a sensory laboratory equipped with 22 individual booths. Refer to Corollaro et al. (2013) for further details
193 regarding the selection of the panel and its performance monitoring, general lexicon development, and
194 evaluation procedures.

195

196 **2.8 Statistical analysis**

197 The outputs of the different analyses were processed, integrated and visualised using the R software (R Core
198 Team, 2016). Tables of correlation were visualized using 'psych' package (Revelle, 2017), principal component
199 analysis (PCA) were calculated using the 'prcomp' function of the 'stat' package; outputs were displayed using
200 the 'factoextra' package (Kassambara and Mundt, 2016). Heatmaps were produced using the packages
201 'ggplot2' and 'Deducer' (Fellows, 2012; Wickham, 2016) and the 'cor' function of the 'stat' package.

202

203 3 RESULTS

204 3.1 Phenotypic analysis of juiciness and fruit texture in apple

205 'Juiciness' was assessed on 10 fruit (3 disks/fruit) per accession through mechanical extraction. The 'juiciness'
206 measured across the 14 accessions showed a normal distribution (Shapiro-Wilk test: $W = 0.99$, p value = 0.52),
207 with a mean and median value of 3.12 and 3.13 g, respectively. Significant differences were observed among
208 accessions (ANOVA test: F value = 12.5, p value < $2.2 \cdot 10^{-16}$; Figure 1A), with 'Lumaga' and 'MN55' showing the
209 lowest (2.75 g) and highest (3.42 g) juice mean value, respectively (Figure 1A, Table 2).

210 The fruit texture of the 14 accessions was phenotypically analysed through the identification of 12
211 parameters related to both the mechanical and acoustic component of texture (Supplementary Figure 1,
212 Table 2). All traits showed a quantitative distribution and the pairwise correlations between parameters
213 ranged from 0 (absence of correlation, as observed for 'Yield force' and 'N. Force Peak') to 1 (perfect
214 correlation, for 'Mean force' and 'Area') (Supplementary Figure 1). For group of parameters with a correlation
215 higher than 0.97, only one was considered for further analyses, thereby reducing data redundancy.
216 Therefore, 'Area' and 'Mean force' were not further considered since both had correlations of 0.97 with 'Max
217 force' (included in the analysis) (Supplementary figure 1).

218 The 10 remaining parameters were used for principal component analysis (PCA) to get insights on the texture
219 differences between samples (Supplementary Figure 2A-B). The combination of the first two PCA dimensions
220 (Dim1 and Dim2) explained a total phenotypic variability of 93.1 % (Dim1 = 73.1 %, Supplementary Figure 2C)
221 providing an accurate overview of the different texture performances among accessions. Dim1 was linked to
222 the overall textural performance and allowed the identification of two main groups, with samples showing
223 low or high textural properties characterized by negative or positive Dim1 values, respectively
224 (Supplementary Figure 2A); to this extent 'Gala' and 'FEM16' showed the highest (4.33) and lowest (-4.09)
225 PC1 values respectively (Supplementary Table 3). These insights were complemented by Dim2 (20 % of the
226 total variability) that allowed a clear distinction of accessions according to the acoustic or mechanical
227 components of texture (respectively red and blue arrows in Supplementary Figure 2B). Cultivars showing

228 positive Dim2 values ('Lumaga' PC2 = 2.94) were characterized by firmer texture while a negative Dim2
229 ('Kizuri' PC2 = -1.79) identified accessions with a high acoustic response (Supplementary Figure 2A-B;
230 Supplementary table 3). The two traits showing the highest divergence in terms of Dim2 loading scores were
231 the 'Final force', representative of the mechanical component, and the 'N. Force Peak' (supplementary Figure
232 2B). Even though the latter was formally considered a mechanical parameter, previous reports have
233 highlighted its strict correlation with acoustical parameters (Costa et al., 2011). The comparison of the
234 phenotypic distribution of the two divergent traits (Supplementary Figure 2D-E) showed substantial
235 differences in the mechanical and acoustic behaviour of the different accessions with 'Lumaga', for instance,
236 showing the highest mean value for 'Final force' and the second lowest value for the 'N. Force Peak' (Table
237 2).

238

239 Table 2: Instrumental measurements of juiciness, texture and cellular morphology for the fourteen accessions in analysis. The phenotyping of texture involved
 240 the analysis of twelve traits either related to the mechanical or acoustic component of texture. Cell morphology was analysed focusing both on cell area (CA) and
 241 cell shape (CS). Measure units are specified in brackets (g = grams, N = Newton, dB = decibel, m = meters).

242

Accession	'Juiciness' [g]	Yield		Max Force [N]	Final Force [N]	Mean Force [N]	Area [N *Strain]	Force		Young's Module [N*Strain]	N. Force Peak	Max Acoustic Pres. [dB]	Mean Acoust. Pres. [dB]	Acoust.		Acoust. Peak	CA [μm^2]	CS
		Force [N]	Force [N]					Lin. Dist.	Lin. Dist.									
Lumaga	2.75	12.4	16.6	14.0	13.6	1,148	106.2	1.6	26.4	58.1	32,858	5,203	626.7	43,858	1.309			
FEM16	2.81	11.1	15.7	12.4	12.9	1,106	122.4	1.9	35.6	64.4	32,897	7,858	830.0	41,562	1.302			
Gala	2.99	7.6	9.2	6.9	7.6	656	99.9	1.2	23.3	54.2	32,833	2,716	281.1	36,051	1.269			
UEB32642	3.00	10.1	11.4	8.5	9.7	822	104.4	1.2	26.8	59.4	32,844	5,334	528.0	48,528	1.334			
Golden Delicious	3.02	6.4	7.9	5.9	6.5	566	100.5	1.3	27.9	57.5	32,840	4,420	477.1	34,642	1.262			
Red Delicious	3.11	5.9	8.0	6.3	6.6	568	102.0	1.2	31.9	57.4	32,860	5,057	612.4	34,719	1.233			
Gradisca	3.12	9.1	12.6	8.7	10.5	900	114.4	1.8	34.7	63.2	32,885	7,475	823.8	34,318	1.272			
CIV323	3.15	10.5	13.3	11.1	11.2	954	107.0	1.4	27.9	58.1	32,847	4,170	480.6	33,137	1.310			
Y102	3.17	10.8	13.3	8.9	11.0	945	111.5	1.7	31.7	61.2	32,864	5,794	658.3	35,356	1.186			
UEB6581	3.19	12.7	15.5	12.8	13.2	1,124	111.9	1.5	31.5	61.9	32,869	6,109	702.3	43,171	1.271			
Minneiska	3.28	5.8	7.2	5.4	6.0	517	99.5	1.0	28.3	56.0	32,848	3,172	425.0	38,465	1.235			
Fuji	3.31	8.7	11.1	8.4	9.4	807	107.5	1.5	31.6	61.9	32,880	6,630	764.9	41,109	1.272			
Kizuri	3.39	10.4	12.6	8.0	10.3	878	123.2	1.5	35.4	66.6	32,899	8,988	824.8	38,731	1.184			
MN55	3.42	10.6	13.0	10.7	11.2	965	110.8	1.7	32.9	64.5	32,886	7,333	792.1	44,457	1.206			

243

244

245

246

247 **3.2 Histological analysis of the flesh tissue in different apple varieties**

248 The analysis of the cell morphology (CA, perimeter, width and height) was conducted on all accessions. Since
249 CA and perimeter showed a high positive correlation ($\text{cor} = 0.9$, $p \text{ value} = < 2^{-16}$), only CA was further
250 considered for analysis. A lower, though significant, correlation was instead observed between CA and Width
251 ($\text{cor} = 0.68$, $p \text{ value} = < 2^{-16}$) as well as between CA and Height ($\text{cor} = 0.66$, $p \text{ value} = < 2^{-16}$), while no correlation
252 was instead observed for Width and Height ($\text{cor} = 0.09$, $p \text{ value} = 9.272 \cdot 10^{-10}$) (Supplementary Figure 3).

253 The number of cells analysed varied from 199 for 'UEB6581' to 343 for 'Gradisca' (mean and median equal
254 to 271 and 272 cells per accessions respectively) according to the different efficiency in the cell extraction
255 procedure (Supplementary Figure 4). To assess if the number of cells analysed was sufficient for robust
256 estimates, 1000 subsets (represented by 10 different subset sizes with 100 repetitions each) for each
257 accession were randomly sampled from the total number of cells and the relative mean CA calculated. The
258 subset sizes were established by dividing the range between 10 cells (arbitrarily set as the minimum sample
259 size to calculate a mean value) and the total number of cells in ten uniform intervals. For each subset size,
260 100 random samplings were performed, and the respective mean CA value calculated. As expected, the CA
261 mean variability within the 100 repetitions decreased with the increase in size of the subset. For all
262 accessions, 100 randomly chosen cells provided already a robust estimate of the overall CA (Supplementary
263 Figure 4). The same process was also performed for perimeter, width, height and CS giving similar results
264 (data not shown).

265 Overall, the CA distribution showed a quantitative, slightly skewed, distribution, with mean and median
266 values equal to 39,653 and 38,390 μm^2 respectively (Table 2). Significant differences between accessions
267 were observed (ANOVA test: $F \text{ value} = 51.45$, $p \text{ value} < 2^{-16}$). Among the fourteen accessions, 'CIV323' and
268 'UEB32642' showed the lowest and highest median CA values (33,137 and 48,528 μm^2), respectively (Figure
269 2A, Table 2). The cultivars standard deviation was normally distributed (Shapiro-Wilk test: $W = 0.97$, $p \text{ value}$
270 $= 0.88$) with extreme values ranging from 9,912 ('Y102') to 15,505 ('Minneiska') (Figure 2A). CS showed

271 instead a skewed distribution with mean and median value equal to 1.35 and 1.25, respectively (Table 2).
272 Differences in CS among cultivars were observed (ANOVA test: F value = 8.75, p value < 2⁻¹⁶), with 'Kizuri' and
273 'UEB32642' showing the lowest and highest median value respectively (1.184 and 1.334 respectively, Figure
274 2B). As for the CA, the cultivar standard deviation was normally distributed (Shapiro-Wilk test: W = 0.92, p
275 value = 0.22). The standard deviation for CS was larger in cultivar showing higher CS (thus a higher fraction
276 of angular cells), with 'Kizuri' and 'UEB32642' showing not only the most extreme CS values but also the
277 lowest and highest standard deviations (Figure 2B).

278

279 **3.3 Combined analysis of 'juiciness', textural properties and cellular characteristics**

280 The texture parameters, together with juiciness and cell morphology related traits, were distributed over a
281 2D-PCA plot (Figure 3, Supplementary Table 3). The first two dimensions explained a cumulative phenotypic
282 variability of 80.9 %. Dim1 (57.3 % of the total variability) was predominantly influenced by individuals with
283 poor texture, such as 'Gala' (PC1 = 4.48) and 'Minneiska' (PC1 = 3.82) plotted in the Dim1 positive area, as
284 well as accessions showing desirable textural features like 'FEM16' and 'Kizuri', distinguished instead by
285 negative Dim1 values (PC1 = -3.97 and -3.53 respectively) (Figure 3A, Supplementary Table 3). PC1 scores
286 were associated with all texture parameters, with 'Max Acoustic Pressure' (cor = -0.95, p values = 0.51⁻⁷)
287 showing the highest association, while neither CS, CA nor 'juiciness' showing significant associations with
288 PC1. Similarly to what observed on supplementary Figure 2B, the mechanical and acoustic properties of
289 texture can be efficiently discriminated through the examination of the Dim2, with individuals showing high
290 firmness ('Lumaga', PC2 = 3.65) or favourable crispness ('Kizuri', PC2 = -2.55) and characterized by the highest
291 positive and negative Dim2 values, respectively (Figure 3A). In addition, the orientation of the accessions
292 along the y-axis was greatly influenced by 'juiciness' and CS (Figure 3A). Both traits, although in opposite
293 directions, showed a projection over the y-axis. However, while the loading representing 'juiciness' directed
294 towards the lower-left quadrant, the CS loading was oriented towards the specular upper-right quadrant
295 (Figure 3A). The visual inspection of the 2D-PCA plot underlined that juiciness and round shape type of cells
296 were distinguished by negative values of Dim2, while elongated type of cells and more dried fruit were

297 characterized by positive values of Dim2. PC2 scores showed the highest correlation with 'juiciness' (corr = -
298 0.76, p value = 0.001) and 'CS' (corr = 0.81, p value = 0.0004). The opposite projection of 'juiciness' and CS
299 loadings underlined an inverse correlation between these two traits, suggesting a direct role of the cellular
300 shape in modulating the juiciness response. This result was further confirmed by the heatmap depicted in
301 Figure 3B, with CS and 'juiciness' showing an inverse correlation (cor = -0.68, p value = 0.007) (Figure 3B). In
302 contrast to what was observed for CS, CA showed a direct correlation with firmness parameters (and not
303 with 'Juiciness') as indicated by its loading orientation with regards to 'Final force', 'Max force' and 'Yield
304 force' (Figure 3A) and correlation values ranging from 0.41 ('Max force') to 0.49 ('Yield force') (Figure 3B).

305

306 **3.4 Interplay between 'juiciness', CS and the MDPG1 SSR10Kd marker**

307 Previous work have highlighted the central role of the cell wall modifying enzyme *MdPG1* in modulating
308 textural characteristics (Longhi et al., 2012, 2013a; Di Guardo et al., 2017) but associations between juiciness
309 and *MdPG1* has not been investigated. To test this hypothesis, the apple accessions were genotyped with
310 the *MDPG1_{SSR}10Kd*, an SSR marker closely linked to *MdPG1* (Longhi et al., 2013a). The *MDPG1_{SSR}10Kd*
311 genotypes were distinguished by three alleles, consistent with the findings previously reported by Longhi
312 et al. (2013a). The two alleles 313 and 317 were previously associated to a favourable textural characteristics
313 (A), while the 324 bp-allele was instead associated with a poor textural performance (a) (Longhi et al., 2013a,
314 2013b; Baumgartner et al., 2016). The presence of two A and one a alleles allowed the distinction of the apple
315 accessions among three genotypic classes (AA, Aa and aa). The apple accessions considered in this work were
316 represented by 10 AA and 4 Aa genotypes (Table 1). *MDPG1_{SSR}10Kd* was associated with all 10 textural
317 parameters (Welch Two Sample t-test, p values ranging from 4.52^{-5} to $< 2.2^{-16}$, Supplementary figure 5) while
318 no significant association was found for *MDPG1_{SSR}10Kd* allelism and 'juiciness' (Welch Two Sample t-test, p
319 value = 0.11), implying that *MDPG1_{SSR}10Kd* alone could not be associated with juiciness (Supplementary
320 Figure 6).

321 The role of *MdPG1_{SSR}10kd* in controlling juiciness was also analysed together with CS, the only parameter
322 showing a significant correlation with 'juiciness' (Figure 4). 'MN55' and 'Kizuri' had the highest 'juiciness' and
323 the lowest CS values (thus the highest fraction of round cells), while 'Lumaga' and 'FEM16' were characterized
324 by a specular phenotype, with lower 'juiciness' and higher CS values (Figure 4). The fitted line (grey line in
325 Figure 4) represented the optimal regression line linking CS with 'juiciness'. All apples plotted below or above
326 the fitted line were characterized by lower or higher 'juiciness' responses respectively, with regards to their
327 cellular morphology. It is interesting to notice that none of the accessions plotted above the fitted line (higher
328 'juiciness' compared to CS) showed the *Aa MdPG1_{SSR}10kd* allelic state. The relationship between the marker
329 allelism and juiciness was further supported by the comparison between the 'juiciness' of accessions showing
330 similar CS value but different *MDPG1_{SSR}10Kd* genotype. Despite 'Y102' (*Aa*) and 'Kizuri' (*AA*) showed a similar
331 CS value (1.186, 1.184), 'Kizuri' was characterized by a significant increase in 'juiciness' (6.9%) compared to
332 'Y102' (Welch Two Sample t-test, p value = 0.01). Similarly, the other cultivars showing an *Aa* genotype were
333 less juicy compared with those characterized by similar CS values and *AA* genotype, with an increase in
334 juiciness ranging from 5.4 % for 'Minneiska' (*AA*) compared to 'Red Delicious' (*Aa*) to 10.7 % for 'Fuji' (*Aa*)
335 compared to 'Gala' (*AA*) (Figure 4). 'Juiciness' can be therefore associated to the combination of CS and the
336 genotype at the *MdPG1 locus*. Similarly, 'juiciness' can result from the combination of either favourable
337 *MDPG1_{SSR}10KD* genetic configuration and high CS values (angular cells) or from samples showing *Aa* genotype
338 and low CS values (round cells). Even though 'Gala' and 'UEB32642' showed similar 'juiciness' (2.99 and 3 g
339 respectively, Table 2), the first cultivar is characterized by *Aa* genotype and a CS value of 1.269, while
340 'UEB32642' showed a favourable *AA* genotype and a less favourable CS of 1.334. Similar patterns were also
341 observed for 'Y102' and 'UEB6581' or for 'Red Delicious' and 'Gradisca'.

342

343 **3.5 Sensory analysis**

344 Fruit quality parameters related to texture, juiciness, flavour and aroma were also assessed using a sensory
345 panel (Table 3). The standard deviations of the nine sensory descriptors showed a normal distribution
346 (Shapiro-Wilk test: $W = 0.91$, p value = 0.37). 'MN55' was characterized by the highest sensory juiciness score

347 (79.5), in agreement with the correspondent instrumental measurement (Table 2). As for the sensory traits
348 related to texture, 'Lumaga' showed the highest response in terms of crispness and crunchiness (67.2 and
349 71.5 respectively), while 'FEM16' and 'UEB6581' were characterized by the highest fibrousness and hardness
350 (73.5 and 74.7, respectively) (Table 3). As for the textural parameters that are generally negatively perceived
351 by consumers, the apples showing the highest mealiness and granularity were 'Golden Delicious' and
352 'CIV323' (67.7 and 60.2 respectively). 'Gradisca' and 'Kizuri' were instead characterized by the highest flavour
353 (62.4) and aroma (65.1) respectively (Table 3).

354 Results of both instrumental and sensory analysis were integrated in a PCA analysis (Figure 5A-B). The first
355 two PCs explained the 76.6 % of the total phenotypic variability. The distribution of the cultivars along the
356 two axes was in agreement with the results depicted in Figure 3, implying that the sensory measurements
357 showed a high (direct or inverse) correlation with at least one of the traits measured instrumentally. The
358 loadings related to 'Mealiness' and 'Granularity' pointed to the lower-left PCA quadrant, and both traits
359 resulted inversely correlated with the mechanical components of texture (Figure 5; Supplementary Table 3).
360 Sensory evaluated parameters, such as 'Flavour', 'Hardiness', 'Fibrousness', 'Crispness' and 'Crunchiness'
361 were inversely correlated to 'Mealiness' and 'Granularity', as shown by the opposite loading projections and
362 the correlation matrix, and positively correlated with the instrumentally measured mechanical parameters
363 related to texture (Figure 5). 'Aroma' and 'Juiciness Sens.' showed orthogonal loadings projection compared
364 with all the other sensory traits, implying a low correlation between these two sensory parameters and all
365 the others (Figure 5). 'Sensory Juiciness' showed instead significant, positive, correlations, ranging from 0.6
366 ('Force Lin. Dis') to 0.8 ('Max Acoust. Pres.') with the instrumentally measured acoustic components of
367 texture (Figures 5), with 'Kizuri' (PC1 = 2.91, PC2 -3.44), 'Gradisca' (PC1 = 2.29, PC2 = -1.07) and 'MN55' (PC1
368 = 4.59, PC2 = -1.42) showing the highest sensory juiciness values and optimal acoustic responses (Table 3,
369 Supplementary Table 3). Both sensory and instrumental juiciness loadings pointed to the lower-right
370 quadrant of the PCA (Figure 5A) with the two parameters showing a correlation of 0.54 (p value = 0.045)
371 (Figure 5B). As for the cellular morphology parameters, CS resulted negatively correlated with the 'aroma'
372 and 'juiciness' (cor = -0.37 and -0.31, respectively), while CA showed a direct correlation with both traits (cor

373 = 0.27 and 0.45, respectively). CA showed a different degree of association with the two type of juiciness
374 assessed (instrumental and sensory), showing a weak indirect correlation and a direct correlation
375 respectively. CS showed instead a more consistent behaviour, being inversely correlated to both
376 measurements of juiciness. CA showed a high direct correlation with both the sensory traits related to
377 texture, and the instrumentally measured mechanical components of texture. Overall, CS was inversely
378 correlated with the instrumentally measured acoustic parameters, and a weak correlation with the
379 mechanical parameters and the sensorially assessed texture (Figure 5 A-B).

380

381 Table 3: Sensory descriptors related to juiciness, texture, aroma and flavour for the fourteen accessions in analysis. Texture descriptors are either related to the
 382 physical (Hardiness, Mealiness, Granularity and Fibrousness) or acoustic response (Crispness, Crunchiness) of the fruit during mastication. The intensity of each
 383 descriptor was expressed as a score on a 100 mm linear scale, ranging from 0 (absence) to 100 (extremely intense).

384

Accession	Juiciness Sens.	Aroma	Crispness	Hardiness	Mealiness	Crunchiness	Granularity	Fibrousness	Flavour
Golden Delicious	32.3	45.2	20.9	11.5	67.7	12.9	60.1	8	45.9
Gala	34.2	47.8	29.6	25.4	56.3	17.6	53.4	12	40.3
CIV323	43.8	59.1	29.8	21	65.4	14.7	60.2	13.5	47.5
Red Delicious	45.7	60.1	44.8	23.9	61.5	24.7	48.8	26.3	37.9
Lumaga	48.3	55	65.5	71.8	8.4	53.4	14.1	68.9	45.4
Y102	52.3	50.9	57.3	68.1	10.6	53.6	26.3	57	49.1
UEB32642	53	53.7	58.4	51.1	29.2	48.8	35.6	37.8	56.1
Minneiska	54	56.8	25.8	17.8	56.2	14.6	57.9	11.7	39.7
FEM16	57.6	47.3	63.3	72.8	7.2	56.6	25.3	73.5	54.2
UEB6581	59.1	62	62.3	74.7	6	55.1	18	68	45.8
Fuji	62.5	50	53.8	46.9	18.6	38.3	34.8	47.6	43.2
Kizuri	63.2	65.1	50.9	43.7	19.2	43.1	32.3	39.4	47.2
Gradisca	66.3	45.8	55	58.5	7.1	52.1	22.5	54.1	62.4
MN55	79.5	65	67.2	72	5.6	71.5	16.2	72.7	49.8

385

386

387 3.6 Validation using X-ray computed micro-tomography

388 The tight correlation between cellular morphology and 'juiciness' was further confirmed by X-ray computed
389 micro-tomography analysis. Example images are displayed in Figure 6 and Supplementary Figure 7. All
390 cultivars showed a bimodal distribution of the log transformed volume of IS, with 'Golden Delicious', 'Fuji'
391 and 'Kizuri' characterized by an increasing relative frequency of small over large IS, while 'Lumaga' showed a
392 more uniform distribution of the IS spaces (Figure 7). This analysis supported and complemented the results
393 obtained with the optical microscopy, confirming the interplay between CS and *MdPG1* allelism on the overall
394 'juiciness'. The seven cultivars depicted in Figure 7 were ordered according to their 'juiciness' in descending
395 order and coloured according to their *MDPG1_{SSR10Kd}* genotype (AA = green, Aa = yellow). The elaboration of
396 the 3D images analysis shows that

397 'juiciness' was influenced by the relative frequency of small IS within individuals showing the same
398 *MDPG1_{SSR10Kd}* genetic background. Among the samples showing an AA genotype, cultivars like 'Kizuri' and
399 'Fuji' were characterized by both the highest fraction of small IS and the highest 'juiciness' values. In contrast,
400 'Lumaga' showed the lowest relative frequency of small IS and the lowest 'juiciness' while 'Minneiska' and
401 'Gradisca' were characterized by intermediate phenotypes for both traits. The same pattern was also
402 observed within the two cultivars showing an Aa genotype with the higher juiciness of 'Golden Delicious'
403 being accompanied by a higher fraction of small IS compared to 'Gala'. As observed in Figure 4, 'Fuji' and
404 'Golden Delicious' showed a similar cellular morphology but a substantial difference in 'juiciness' response
405 (Figure 7, Table 2).

406

407 4 DISCUSSION

408 4.1 Analysis of the cellular morphology

409 Two complementary approaches were adopted to study cellular morphology focusing either on the cell
410 (measuring CA and CS) or on the IS (measured by volume). While CA was slightly correlated with IS (cor =
411 0.24), the latter showed a higher positive correlation with CS (cor = 0.67).

412 The range of CA ($33,137 \mu\text{m}^2 - 48,528 \mu\text{m}^2$) was similar to what observed in previous work using the same
413 extraction protocol (McAtee et al., 2009), but was higher when compared to that using different cell
414 extraction and observation methods (Allan-Wojtas et al., 2003; Schotsmans et al., 2004) To this end it is in
415 fact interesting to underline that our study was based on observation made on whole cells, while most of the
416 works published to date were on the contrary based on light microscopy analysis carried out on fruit slices,
417 leading, therefore, to wrong cell size estimations, as already pointed out by McAtee et al. (2009).

418 The examination of the CS relied on the fact that previous analysis using confocal microscopy confirmed that
419 cell shape was not altered mechanically or osmotically during the cell isolation process (McAtee et al., 2009).
420 Interestingly, apples showing the highest fraction of round cells ('Kizuri', 'Y102') were also those
421 characterized by the lowest CS standard deviations and vice-versa (Figure 2B). The absence of cultivars
422 showing high CS (thus high fraction of elongated cells) and low standard deviation could be either due to the
423 sampling of elite germplasm or to the relative frequency of round cells that could not be lower than a certain
424 amount. Further studies on wider germplasm collections could better clarify this aspect.

425

426 **4.2 Comparison between instrumental and sensory data**

427 The fruit quality features were analysed across the accessions using instrumental and sensorial approaches.
428 Analytically measured juiciness was correlated with its sensory evaluation, validating the phenotyping
429 protocol employed in the study. Moreover, sensory juiciness showed the highest correlations with the
430 instrumentally measured acoustic parameters ('max acoustic pressure' and 'mean acoustic pressure'),
431 confirming the high relationship between acoustic response and juiciness (Daillant-Spinnler et al., 1996;
432 Allan-Wojtas et al., 2003; Corollaro et al., 2014a). However, the analytically measured juiciness showed the
433 highest correlation (-0.68) with CS, suggesting a specific role of cellular shape in influencing the apple
434 juiciness response. We believe that this is the first report of cellular shape influencing the juiciness of apple.
435 Even though the two measurements of juiciness were correlated, that between CS and sensory juiciness was
436 below the significance threshold. Further studies with larger germplasm collections and/or employing an

437 increased number of panellists, could help to elucidate the slightly different behaviour of the two
438 measurements of juiciness. As for the correlation between cell size and the instrumentally measured texture
439 parameters, CA was highly correlated with 'Yield force', which indicates the point of transition from the
440 elastic (reversible) to the plastic phase (irreversible crushing) of the apple fruit flesh compression (Costa et
441 al., 2011) (Figure 5B). Among the sensory parameters, CA, instead, showed the highest correlation (cor =
442 0.66, p value = 0.01) with 'Crispness', which represents the acoustic response of the fruit during the first bite.
443 Combining both results, CA was more related to the instrumental and sensory parameters characterizing the
444 first bite suggesting a role of CA in modulating the apple response when consumed fresh. Comparing the two
445 parameters related to cell morphology, CA was directly related to the analytically measured textural
446 parameters, while CS showed a differential pattern for the mechanical and acoustic parameters (being
447 directly and inversely correlated to the two groups of traits, respectively) (Figure 5B). This aspect, coupled
448 with the inverse correlation to 'Juiciness', made CS an ideal candidate to guide the selection of juicy and
449 crispy apple fruit. Juiciness and cellular characteristics have also direct repercussion on apple flavour and
450 aroma (Farneti et al., 2017). The direct correlation between aroma and 'juiciness' (and accordingly the inverse
451 correlation between aroma and CS) can be reconducted to the fact that most of the volatiles that composes
452 the apple aromatic *bouquet* are released (or *de novo* produced) upon cell breakage and consequent juice
453 release. Thus, the selection of apple showing an increased fraction of round cells could result not only in
454 higher juiciness but also in a better response in terms of aromatic production.

455

456 **4.3 Effect of the *MdPG1* allelic variation on cell morphology**

457 The depolymerization of the polysaccharide structure in the middle lamella is one of the major events
458 affecting the texture response in apple (Atkinson et al., 2012; Ben-Arie and Kisleev, 1979; Brummell, 2006).
459 Our results confirmed the important role exerted by *MdPG1* and the direct correlation between the
460 microsatellite marker designed in the proximity of this gene and textural features (supplementary Figure 5).

461 Even though MDPG1_{SSR}10Kd was not statistically associated with differences in juice release, cultivars
462 showing an AA genotype were juicier than those with an Aa genotype (Supplementary Figure 6), suggesting
463 a possible interplay between the physiological mechanisms governing the cell wall metabolism and the
464 overall juiciness. Both analytical and sensory evaluation of fruit quality indicate that CS within each genotype
465 categories affected overall apple juiciness (Figures 4, 7). Our hypothesis is that neither the cellular shape (or
466 equally the size of the IS) nor MDPG1_{SSR}10Kd alone could explain the different response of accessions in terms
467 of juiciness. The analysis of CS showed that similar values of 'juiciness' could be achieved, either with a high
468 fraction of round cell and Aa or with a relatively lower fraction of round cell and AA (Figure 4). Thus, if not
469 adequately considered as a cofactor, *MdPG1* could ultimately act as a confounding variable masking a real
470 correlation between cell morphology and texture or juiciness. This is confirmed by the fact that the best
471 performances in terms of juiciness were registered for accessions combining low CS value and AA genotype
472 as shown for 'Kizuri' and 'MN55' (Table 2).

473

474 5 CONCLUSIONS

475 Economic success of an apple cultivar is determined by its juiciness and texture performances at harvest and
476 after storage. Our results highlight that these factors are influenced by the combination of cellular
477 morphology and the activity of *MdPG1* after storage. The role of cell shape (rather than cell size) influences
478 juiciness, especially when the accessions are grouped according to their genotype at the MDPG1_{SSR}10Kd locus
479 Nevertheless, further studies on wider germplasm is necessary to better elucidate the complex physiological
480 regulation of juiciness and crispness. Taking the results presented here as a proof of concept, the
481 establishment of an automated pipeline for an accurate phenotyping of the cell morphology could enable, in
482 a close future, a valuable large screening tool for this feature.

483 Breeding programmes for juiciness often rely on the use of this marker for marker-assisted seedling selection
484 (MASS) in light of the correlation between textural parameters and juiciness (King et al., 2000; Ulrich et al.,
485 2014). Since the fruit texture is considered a fundamental trait in modern breeding programs, most of the

486 advanced selections are now homozygous for the favourable allele at this locus. Despite this favourable
487 genetic configuration, breeding material showing a fixed AA genotype is still characterized by important
488 differences in terms of texture and juiciness attributes, due to the polygenic regulation of such features.
489 Furthermore, the use of this marker in breeding schemes for texture did not allow the selection of crispy over
490 firm apple since *MdPG1* is linked to both components of texture. The identification of molecular markers
491 linked to CS could greatly support the breeding for juiciness and aroma. The use of CS to guide the seedling
492 selection could also allow a more precise analysis of the two textural components since CS showed a different
493 pattern of correlation between the acoustic (prevalence of round cells) and mechanical (prevalence of
494 elongated cells) texture parameters.

495

496 **6. ACKNOWLEDGMENT**

497 This work has been funded under the Program LP 6/99 art. 5 Autonomous Province of Trento by the CIF
498 Project 'Apple & Berry'. The work was carried out in the frame of the PON "AIM: Attrazione e Mobilità
499 Internazionale" No. 1808223-2 project.

500

501 **7. REFERENCES**

- 502 Abbott, J.A. 1999. Quality measurement of fruits and vegetables. *Postharvest Biology and Technology* 15:
503 207–225.
- 504 Allan-Wojtas, P.; Sanford, K.A.; Mcrae, K.B.; Carbyn, S. 2003. An integrated microstructural and sensory
505 approach to describe apple texture. *Journal of the American Society for Horticultural Science* 128: 381–
506 390.
- 507 Atkinson, R.G.; Sutherland, P.W.; Johnston, S.L.; Gunaseelan, K.; Hallett, I.C.; Mitra, D.; Brummell, D.A.;
508 Schröder, R.; Johnston, J.W.; Schaffer, R.J. 2012. Down-regulation of POLYGALACTURONASE1 alters
509 firmness , tensile strength and water loss in apple (*Malus x domestica*) fruit. *BMC Plant Biology*: 11–21.
- 510 Baumgartner, I.O.; Kellerhals, M.; Costa, F.; Dondini, L.; Pagliarani, G.; Gregori, R.; Tartarini, S.; Leumann, L.;
511 Laurens, F.; Patocchi, A. 2016. Development of SNP-based assays for disease resistance and fruit quality
512 traits in apple (*Malus × domestica* Borkh.) and validation in breeding pilot studies. *Tree Genetics and*
513 *Genomes* 12: 12–35.
- 514 Ben-arie, R.; Kislev, N. 1979. Ultrastructural changes in the cell walls of ripening apple and pear fruit. *Plant*
515 *Physiology* 64: 197–202.
- 516 Bertin, N.; Gautier, H.; Roche, C. 2001. Number of cells in tomato fruit depending on fruit position and source-
517 sink balance during plant development. *Plant Growth Regulation* 57221: 1–8.
- 518 Bourne, M. 2002. *Food texture and viscosity: concept and measurement*. Elsevier.
- 519 Brummell, D.A. 2006. Cell wall disassembly in ripening fruit. *Functional Plant Biology* 33: 103–119.
- 520 Brummell, D.A.; Harpster, M.H. 2001. Cell wall metabolism in fruit softening and quality and its manipulation
521 in transgenic plants. *Plant Molecular Biology* 47: 311–340.
- 522 Cappellin, L.; Farneti, B.; Di Guardo, M.; Busatto, N.; Khomenko, I.; Romano, A.; Velasco, R.; Costa, G.; Biasioli,
523 F.; Costa, F. 2015. QTL analysis coupled with PTR-ToF-MS and candidate gene-based association
524 mapping validate the role of Md-AAT1 as a major gene in the control of flavor in apple fruit. *Plant*

525 Molecular Biology Reporter 33: 239–252.

526 Charles, M.; Corollaro, M.L.; Manfrini, L.; Endrizzi, I.; Aprea, E.; Zanella, A.; Corelli Grappadelli, L.; Gasperi, F.
527 2018. Application of a sensory–instrumental tool to study apple texture characteristics shaped by
528 altitude and time of harvest. Journal of the Science of Food and Agriculture 98: 1095–1104.

529 Chen, P.M.; Borgic, D.M. 1985. Changes in water soluble polyuronides in the pulp tissue of ripening “Bosc”
530 pears following cold storage in air or in 1% oxygen. Journal of the American Society for Horticultural
531 Science 110.

532 Corollaro, M.L.; Aprea, E.; Endrizzi, I.; Betta, E.; Demattè, M.L.; Charles, M.; Bergamaschi, M.; Costa, F.;
533 Biasioli, F.; Corelli, L.; Gasperi, F. 2014a. A combined sensory-instrumental tool for apple quality
534 evaluation. Postharvest Biology and Technology 96: 135–144.

535 Corollaro, M.L.; Endrizzi, I.; Bertolini, A.; Aprea, E.; Demattè, M.L.; Costa, F.; Biasioli, F.; Gasperi, F. 2013.
536 Sensory profiling of apple: methodological aspects, cultivar characterisation and postharvest changes.
537 Postharvest Biology and Technology 77: 111–120.

538 Corollaro, M.L.; Gasperi, F.; Corelli Grappadelli, L. 2014b. An overview of sensory quality of apple fruit. Journal
539 of the American Pomological Society 68: 141–157.

540 Costa, F.; Cappellin, L.; Fontanari, M.; Longhi, S.; Guerra, W.; Magnago, P.; Gasperi, F.; Biasioli, F. 2012.
541 Texture dynamics during postharvest cold storage ripening in apple (*Malus × domestica* Borkh.).
542 Postharvest Biology and Technology 69: 54–63.

543 Costa, F.; Cappellin, L.; Longhi, S.; Guerra, W.; Magnago, P.; Porro, D.; Soukoulis, C.; Salvi, S.; Velasco, R.;
544 Biasioli, F.; Gasperi, F. 2011. Assessment of apple (*Malus × domestica* Borkh.) fruit texture by a
545 combined acoustic-mechanical profiling strategy. Postharvest Biology and Technology 61: 21–28.

546 Daillant-Spinnler, B.; MacFie, H.J.H.; Beyts, P.K.; Hedderley, D. 1996. Relationship between perceived sensory
547 properties and major preference directions of 12 varieties of apples from the southern hemisphere.
548 Food Quality and Preference 7: 113–126.

549 Endrizzi, I.; Torri, L.; Corollaro, M.L.; Demattè, M.L.; Aprea, E.; Charles, M.; Biasioli, F.; Gasperi, F. 2015. A
550 conjoint study on apple acceptability: sensory characteristics and nutritional information. *Food Quality*
551 *and Preference* 40: 39–48.

552 Farneti, B.; Di Guardo, M.; Khomenko, I.; Cappellin, L.; Biasioli, F.; Velasco, R.; Costa, F. 2017. Genome-wide
553 association study unravels the genetic control of the apple volatilome and its interplay with fruit
554 texture. *Journal of Experimental Botany* 68: 1467–1478.

555 Feldkamp, L.A.; Davis, L.C.; Kress, J.W. 1984. Practical cone-beam algorithm. *Journal of the Optical Society of*
556 *America A* 1: 612–619.

557 Fellows, I. 2012. A Data Analysis GUI for R. *Journal of Statistical Software* 49: 1–15.

558 Giovannoni, J. 2004. Genetic regulation of fruit development and ripening. *The Plant Cell* 16: 170–181.

559 Di Guardo, M.; Bink, M.C.A.M.; Guerra, W.; Letschka, T.; Lozano, L.; Busatto, N.; Poles, L.; Tadiello, A.; Bianco,
560 L.; Visser, R.G.F.; van de Weg, E.; Costa, F. 2017. Deciphering the genetic control of fruit texture in apple
561 by multiple family-based analysis and genome-wide association. *Journal of Experimental Botany* 68:
562 1451–1466.

563 Harker, F.R.; Gunson, F.A.; Jaeger, S.R. 2003. The case for fruit quality: an interpretive review of consumer
564 attitudes, and preferences for apples. *Postharvest Biology and Technology* 28: 333–347.

565 Harker, F.R.; Stec, G.H.; Hallett, I.C.; Bennett, C.L. 1997. Texture of parenchymatous plant tissue : a
566 comparison between tensile and other instrumental and sensory measurements of tksue strength and
567 juiciness. *Postharvest biology and technology* 5214: 63–72.

568 Kassambara, A.; Mundt, F. 2016. Factoextra: extract and visualize the results of multivariate data analyses. R
569 package version 1.

570 King, G.J.; Maliepaard, C.; Lynn, J.R.; Alston, F.H.; Durel, C.E.; Evans, K.M.; Griffon, B.; Laurens, F.; Manganaris,
571 A.G.; Schrevens, E.; Tartarini, S.; Verhaegh, J. 2000. Quantitative genetic analysis and comparison of
572 physical and sensory descriptors relating to fruit flesh firmness in apple (*Malus pumila* Mill.). *Theoretical*

573 and Applied Genetics 100: 1074–1084.

574 Lill, R.E.; Mespel, G.J. Van Der 1988. A method for measuring the juice content of mealy nectarines. Scientia
575 Horticulturae 36: 267–271.

576 Longhi, S.; Moretto, M.; Viola, R.; Velasco, R.; Costa, F. 2012. Comprehensive QTL mapping survey dissects
577 the complex fruit texture physiology in apple (*Malus x domestica* Borkh.). Journal of Experimental
578 Botany 63: 1107–1121.

579 Longhi, S.; Cappellin, L.; Guerra, W.; Costa, F. 2013a. Validation of a functional molecular marker suitable for
580 marker-assisted breeding for fruit texture in apple (*Malus x domestica* Borkh.). Molecular Breeding 32:
581 841–852.

582 Longhi, S.; Hamblin, M.T.; Trainotti, L.; Peace, C.P.; Velasco, R.; Costa, F. 2013b. A candidate gene based
583 approach validates Md-PG1 as the main responsible for a QTL impacting fruit texture in apple (*Malus x*
584 *domestica* Borkh). BMC plant biology 13: 37.

585 Mann, H.; Bedford, D.; Luby, J.; Tong, C. 2005. Relationship of instrumental and sensory texture
586 measurements of fresh and stored apples to cell number and size. HortScience 40: 1815–1820.

587 McAtee, P.A.; Hallett, I.C.; Johnston, J.W.; Schaffer, R.J. 2009. A rapid method of fruit cell isolation for cell
588 size and shape measurements. Plant Methods 5: 1–7.

589 Mendoza, F.; Verboven, P.; Mebatsion, H.K.; Kerckhofs, G. 2007. Three-dimensional pore space quantification
590 of apple tissue using X-ray computed microtomography. Planta: 559–570.

591 Ng, J.K.T.; Schröder, R.; Sutherland, P.W.; Hallett, I.C.; Hall, M.I.; Prakash, R.; Smith, B.G.; Melton, L.D.;
592 Johnston, J.W. 2013. Cell wall structures leading to cultivar differences in softening rates develop early
593 during apple (*Malus x domestica*) fruit growth. BMC Plant Biology 13: 183.

594 Nybom, H.; Esselink, G.D.; Werlemark, G.; Vosman, B. 2003. Microsatellite DNA marker inheritance indicates
595 preferential pairing between two highly homologous genomes in polyploid and hemisexual dog-roses,
596 *Rosa L. Sect. Caninae* DC. Heredity 92: 139–150.

597 Olmstead, J.W.; Iezzoni, A.F.; Whiting, M.D. 2007. Genotypic differences in sweet cherry fruit size are
598 primarily a function of cell number. *Journal of the American Society for Horticultural Science* 132.

599 Quilot, B.; Genard, M. 2008. Is competition between mesocarp cells of peach fruits affected by the
600 percentage of wild species (*Prunus davidiana*) genome? *Journal of Plant Research* 121:55: 55–63.

601 Revelle, W.R. 2017. psych: Procedures for personality and psychological research title. [Online] Available:
602 <https://cran.r-project.org/package=psych>.

603 Ridler, T.; Calvard, S. 1978. Picture Thresholding Using an Iterative Selection Method. *IEEE Transactions on*
604 *Systems, Man, and Cybernetics* 8: 630–632.

605 Schindelin, J.; Arganda-Carreras, I.; Frise, E.; Kaynig, V.; Longair, M.; Pietzsch, T.; Preibisch, S.; Rueden, C.;
606 Saalfeld, S.; Schmid, B.; Tinevez, J.-Y.; White, D.J.; Hartenstein, V.; Eliceiri, K.; Tomancak, P.; Cardona, A.
607 2012. Fiji: an open-source platform for biological-image analysis. *Nature Methods* 9: 676–682. [Online]
608 Available: <https://doi.org/10.1038/nmeth.2019>.

609 Schotsmans, W.; Verlinden, B.E.; Lammertyn, J.; Nicolai, B.M. 2004. The relationship between gas transport
610 properties and the histology of apple. *Journal of the Science of Food and Agriculture* 84: 1131–1140.

611 Seymour, G.B.; Manning, K.; Eriksson, E.M.; Popovich, A.H.; King, G.J. 2002. Genetic identification and
612 genomic organization of factors affecting fruit texture. *Journal of experimental botany* 53: 2065–2071.

613 Smedt, V. De; Pauwels, E.; Baerdemaeker, J. De; Nicolai, B. 1998. Microscopic observation of mealiness in
614 apples : a quantitative approach. *Postharvest Biology and Technology* 14: 151–158.

615 Ting, V.J.L.; Silcock, P.; Bremer, P.J.; Biasioli, F. 2013. X-ray micro-computer tomographic method to visualize
616 the microstructure of different apple cultivars. *Journal of Food Science* 78.

617 Ulrich, D.; Hoberg, H.; Fisher, C. 2014. Diversity and dynamic of sensory related traits in different apple
618 cultivars. *Journal of Applied Botany and Food Quality* 83: 70–75.

619 Wakasa, Y.; Kudo, H.; Ishikawa, R.; Akada, S.; Senda, M.; Niizeki, M.; Harada, T. 2006. Low expression of an
620 endopolygalacturonase gene in apple fruit with long-term storage potential. *Postharvest Biology and*

621 Technology 39: 193–198.

622 Waldron, K.W.; Parker, M.L.; Smith, A.C. 2003. Plant Cell Walls and Food Quality. Comprehensive reviews in
623 food science and food safety 2.

624 Wickham, H. 2016. ggplot2: Elegant graphics for data analysis. Springer-Verlag New York. [Online] Available:
625 <https://ggplot2.tidyverse.org>.

626 Figure 1: Boxplots of the 'juiciness' distribution within each of the fourteen samples.

627 Figure 2: Boxplot of the cell area (CA) (A) and cell size (CS) (B) distribution within the fourteen cultivars or
628 advances selections.

629 Figure 3: PCA analysis of the instrumentally measured phenotypic data for 'juiciness', texture and cell
630 morphology. (A) PCA scoring and loading plot; (B) heatmap of the pairwise correlations between the traits in
631 analysis, correlations (B). values exceeding the significance threshold level (p value > 0.5) were crossed.

632 Figure 4: biplot representing the correlation between CS and 'juiciness'. Samples are coloured according to
633 their genetic configuration at the MDPG1_{SSR}10KD locus as specified in legend; the overall linear regression
634 linking the two variables is represented by a grey line while the linear regression made on the base of the
635 two MDPG1_{SSR}10KD genotypes were represented in blue (AA) and pink (Aa) respectively.

636 Figure 5: PCA analysis of the instrumentally measured phenotypic data for 'juiciness', texture and cell
637 morphology and the sensory data. (A) PCA scoring and loading plot. (B) Pairwise correlations plot, traits that
638 were evaluated instrumentally or through a sensory panel are labelled in green and violet respectively. Values
639 exceeding the significance threshold level (p value > 0.5) were crossed.

640 Figure 6: Cross-sectional slice obtained after tomographic reconstruction (A). Binary image displaying the
641 segmented intercellular spaces in white (B). 3D rendering of a reconstructed cylindrical sample (C).

642 Figure 7: Histograms of the log transformed volume of the intercellular space. Samples are coloured
643 according to their genetic configuration at the MDPG1 SSR10Kd locus (yellow = Aa, green = AA). For each

644 accession, the density distribution of the three fruits analysed for each cultivar is depicted as blue, green and
645 red continuous lines.

646

647 Supplementary Figure 1: Texture profiling of the 14 apple cultivars stored under normal atmospheric
648 conditions. The distribution of each texture parameter (both as histogram and as density fitted line) is
649 represented on the diagonal. On the lower triangle the bivariate scatter plots and the relative fitted line (red
650 continuous line) are depicted while the respective absolute correlation values are reported on the upper
651 triangle.

652 Supplementary Figure 2: Principal component analysis of the textural parameters; A: Scoreplot of the first
653 two principal components, individuals are coloured according to their qualities of representation, B: loading
654 plots of the ten variables used to compute the principal component analysis, arrows are coloured according
655 to the type of variables (acoustic or mechanic). (C) barplot representing the percentage of the explained
656 variance of all computed dimensions (principal components). The boxplot of the two variables showing the
657 most divergent loading scores and grouped by cultivars are represented in D ('Final force') and E ('Number
658 of force peaks').

659 Supplementary Figure 3: Analysis of the cellular morphology on the germplasm in analysis. The distribution
660 of each of the four parameters (both as histogram and as density fitted line) is represented on the diagonal.
661 On the lower triangle the bivariate scatter plots and the relative fitted line (red continuous line) are depicted
662 while the respective absolute correlation values are reported on the upper triangle.

663 Supplementary Figure 4: For each cultivar, the average cell area (CA, y axis) is calculated for ten subsets of
664 increasing size ranging from 10 cells to the total number of cells analysed (x axis). For each sample sizes, 100
665 random samplings were extracted, the respective average area calculated, and its distribution plotted (grey
666 boxplots). The red, horizontal line represents, for each cultivar, the average value of the cell area when all
667 cells are considered, cultivars are ordered from left to right and from top to bottom according to the quantity
668 of extracted juice.

669 Supplementary Figure 5: Boxplots of the 10 textural parameters analysed. Traits that are related to
670 mechanical or acoustic parameters are coloured in blue or red respectively. For each trait, individuals are
671 grouped according to their genotype at the MDPG1 SSR10Kd *locus* (heterozygous =:Aa, homozygous = AA).

672 Supplementary Figure 6: Boxplots of the 'juiciness' explained by the genotype at the MDPG1 SSR10Kd *locus*
673 (heterozygous = Aa, homozygous = AA).

674 Supplementary Figure 7: Virtual slicing of a sample of the 'Kizuri' cultivar, reconstructed by X-ray
675 microtomography.

676

Figure 1

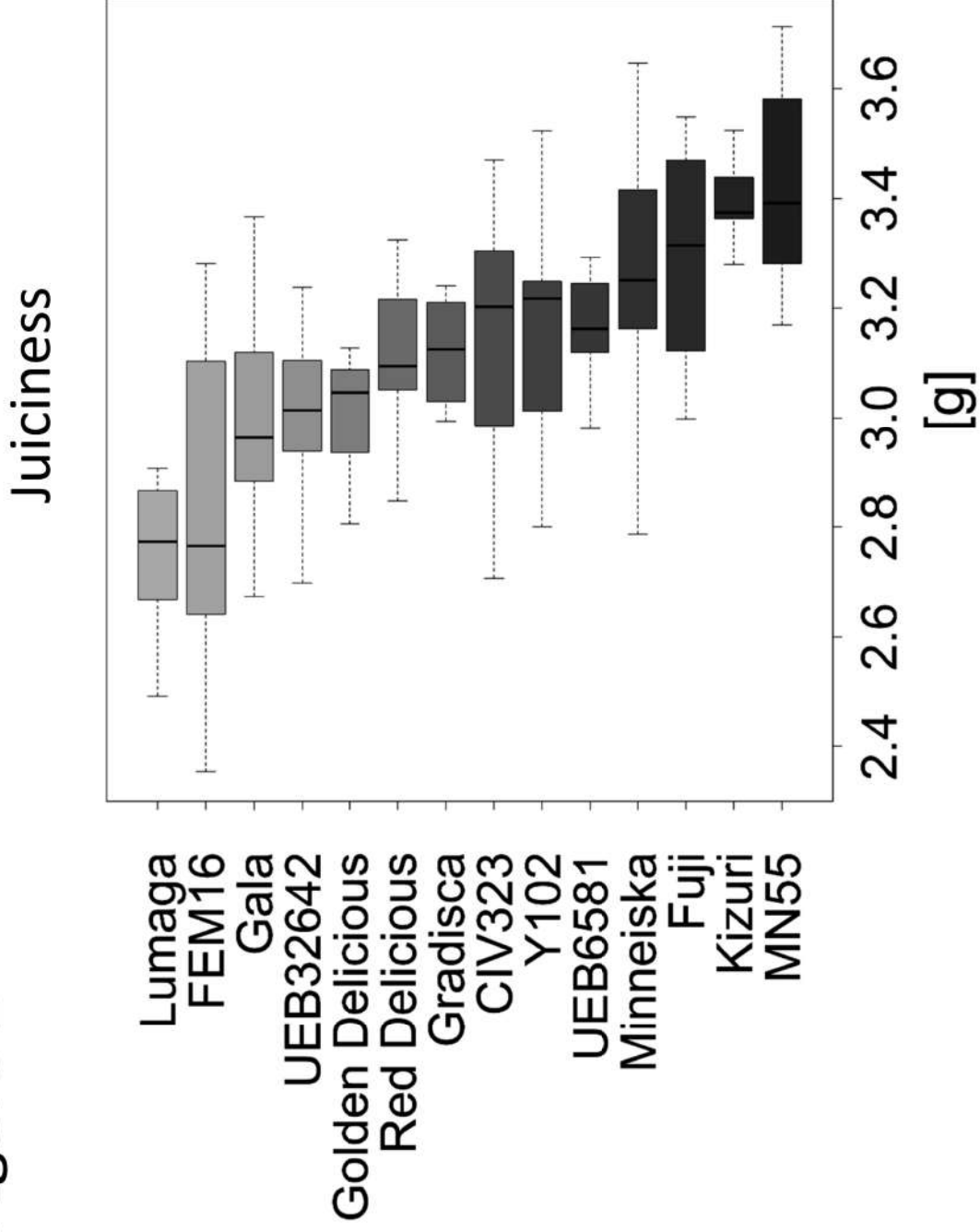


Figure 2

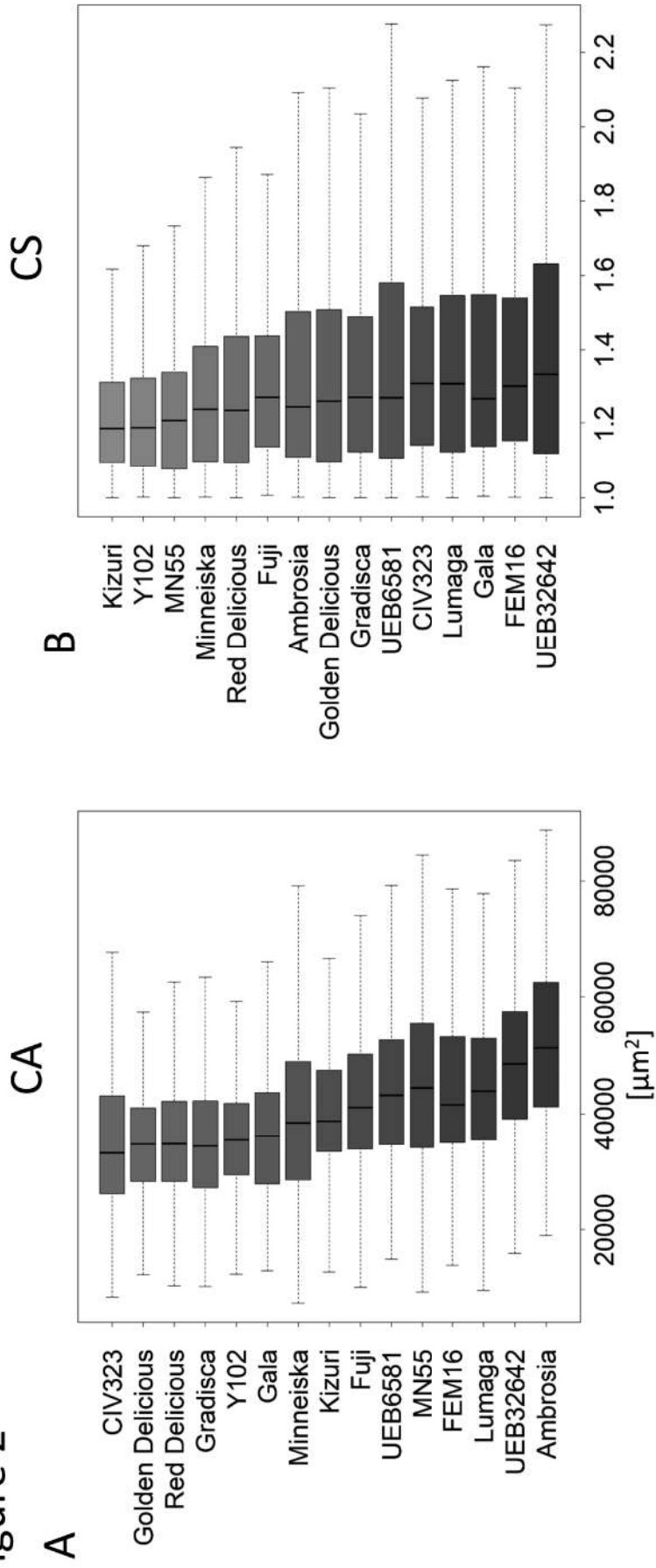
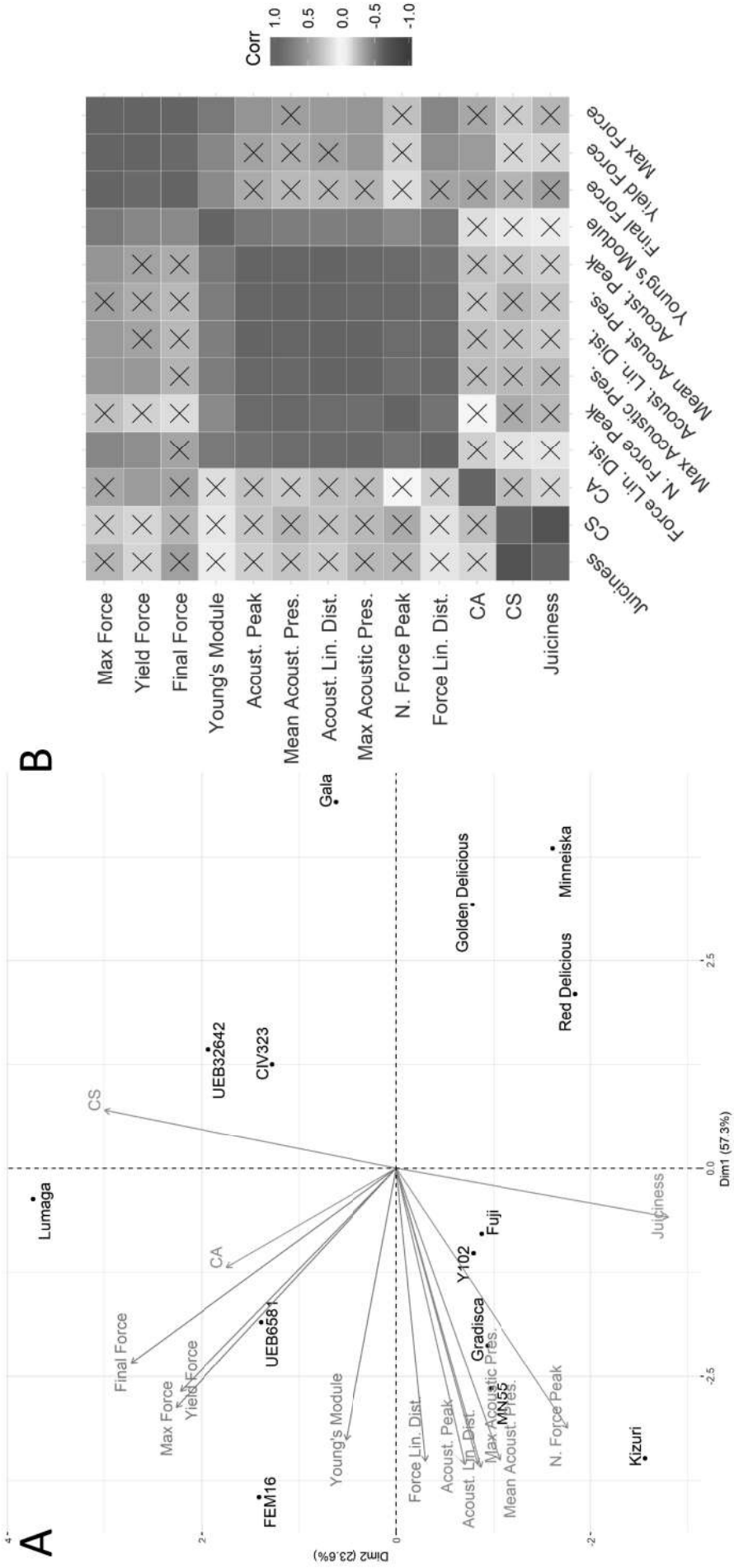


Figure 3



Col. ⇨ Acoustic Parameters ⇨ Cell morphology ⇨ Juiciness ⇨ Mechanic Parameters

Figure 4

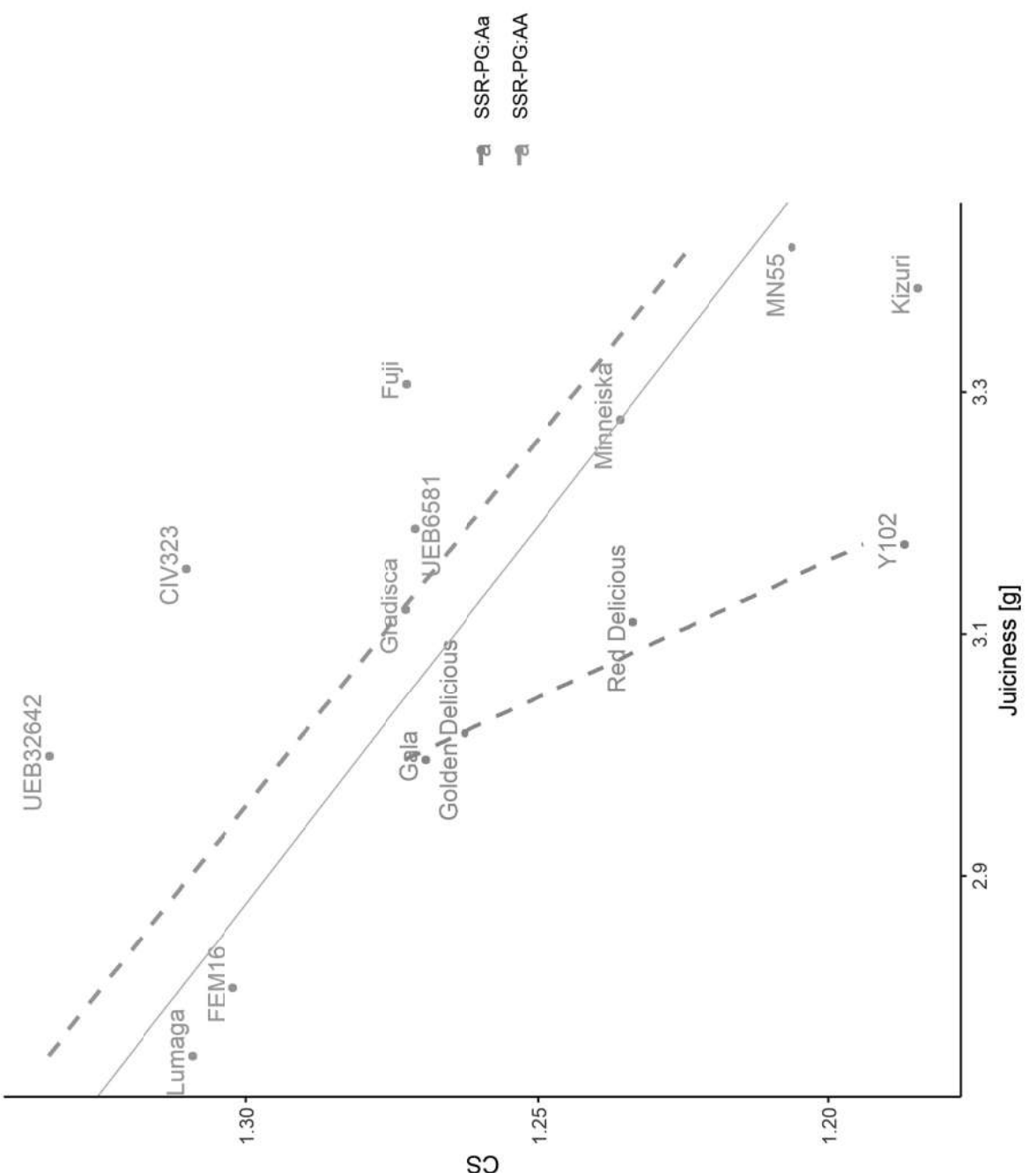
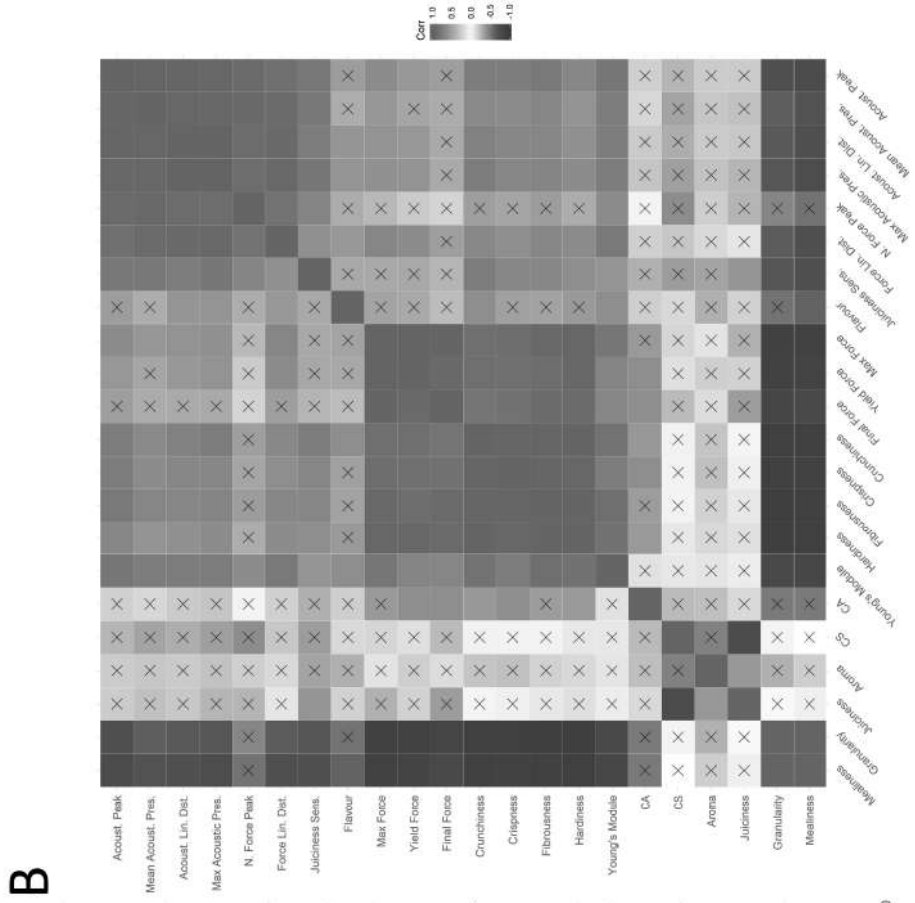
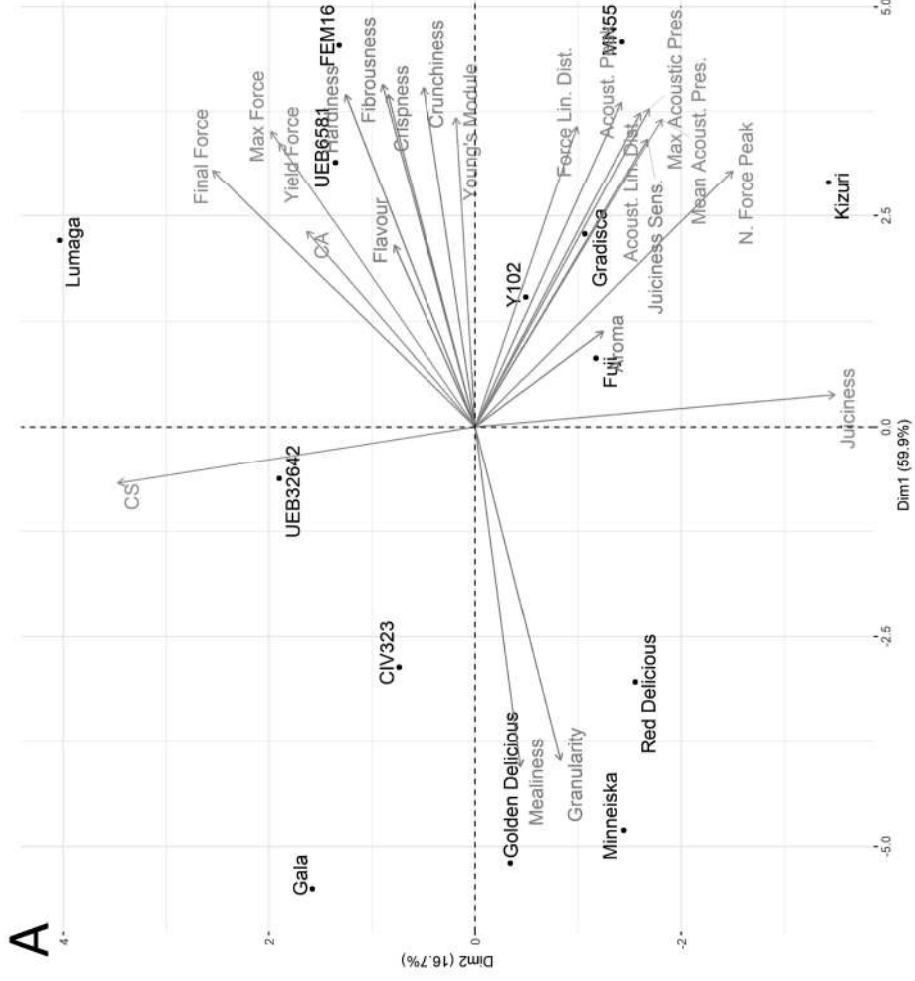


Figure 5



Col. -> Acoustic Parameters -> Cell morphology -> Juiciness -> Mechanical Parameters -> Sensory Parameters

Figure 6

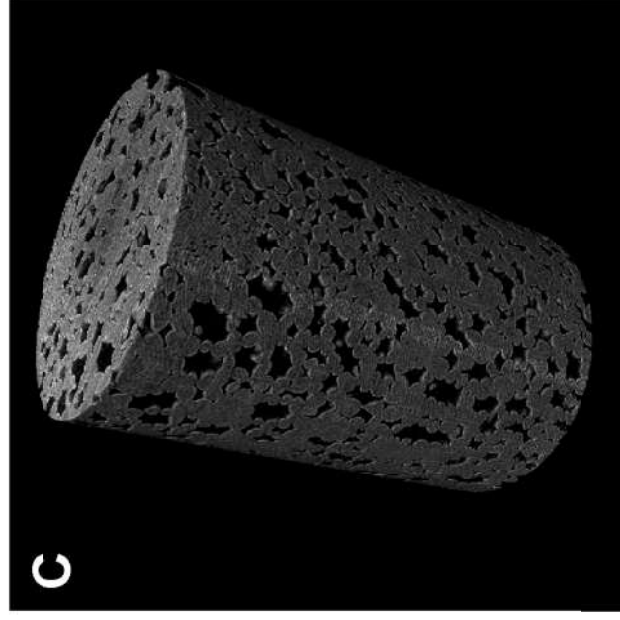
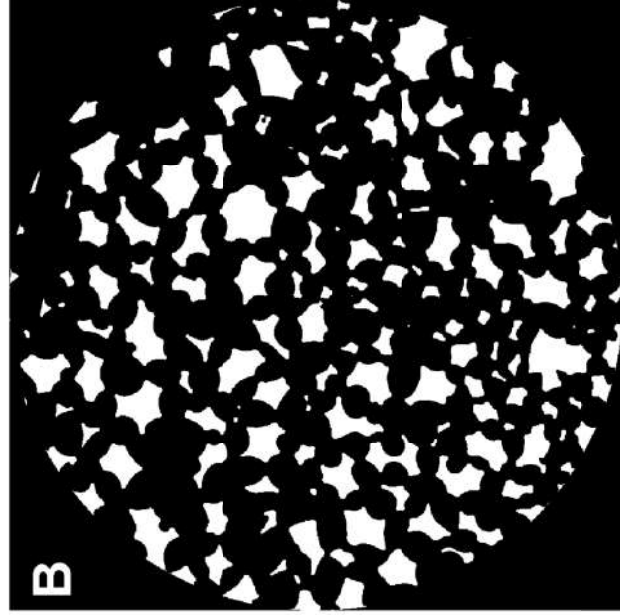
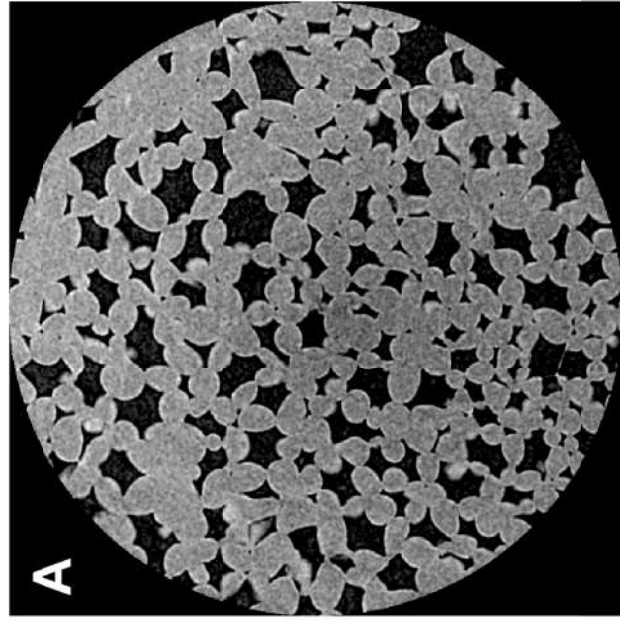
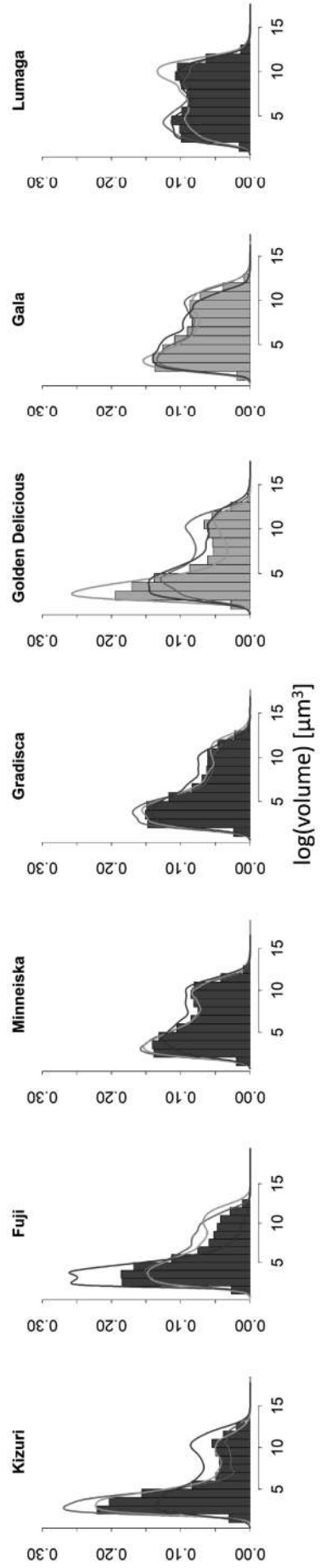
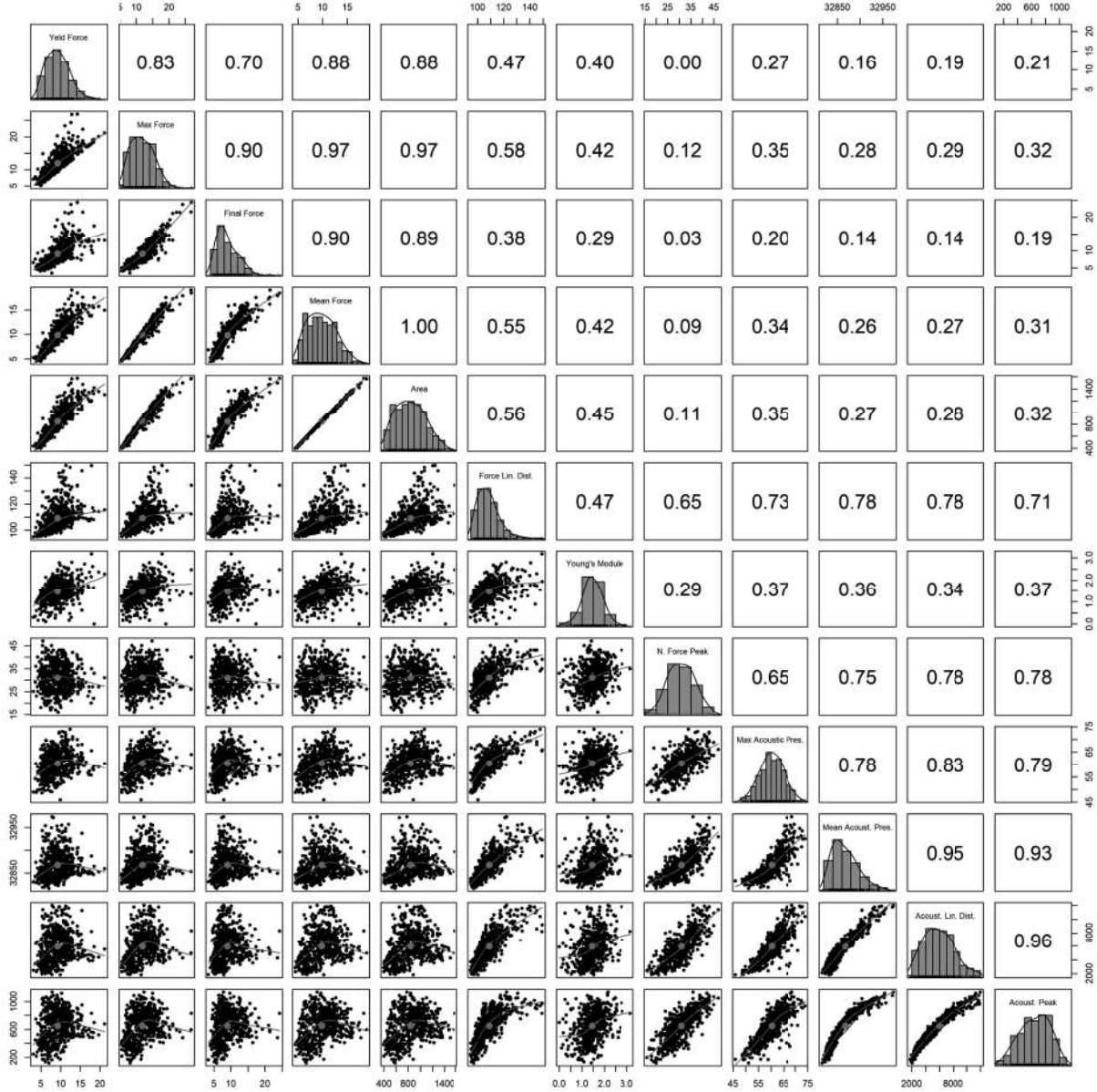


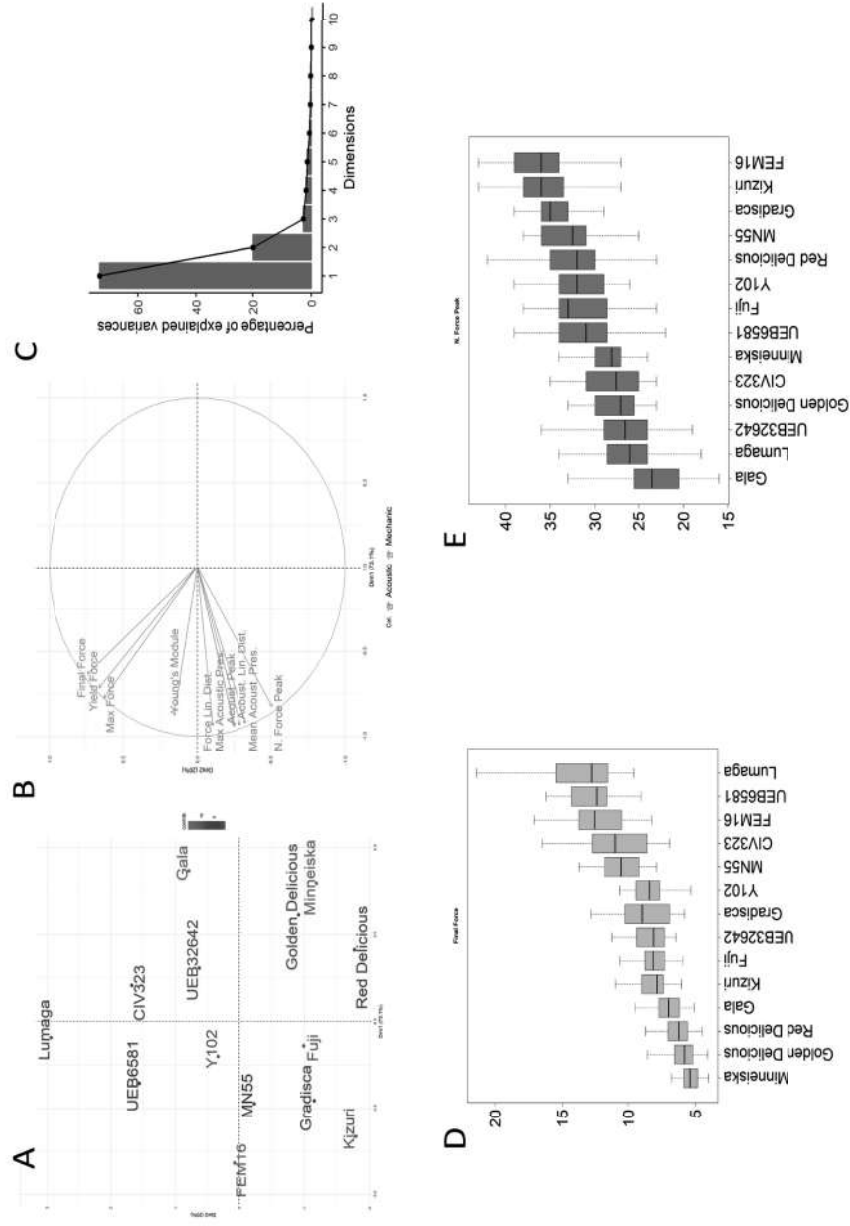
Figure 7



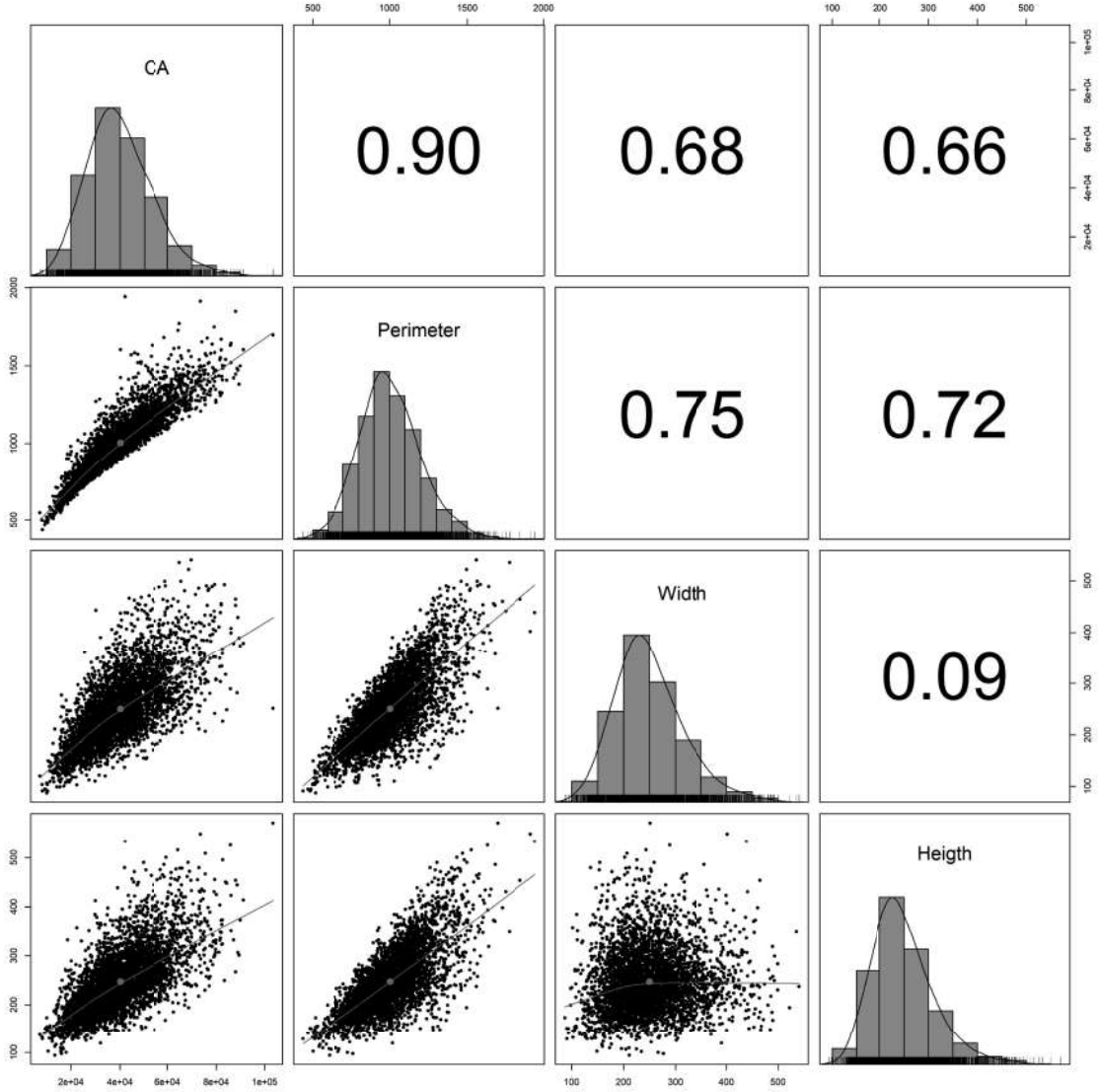
Supplementary Figure 1



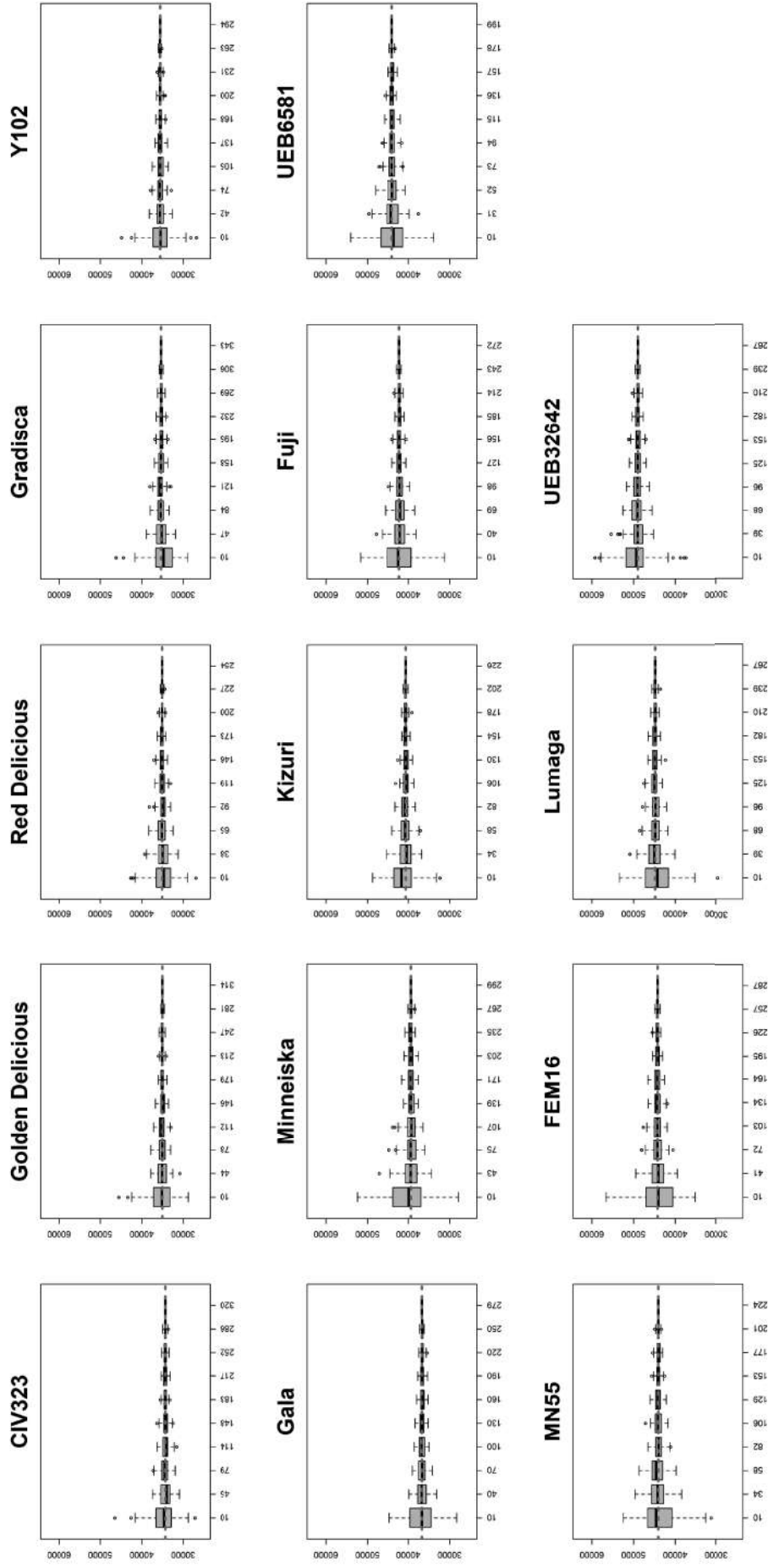
Supplementary Figure 2



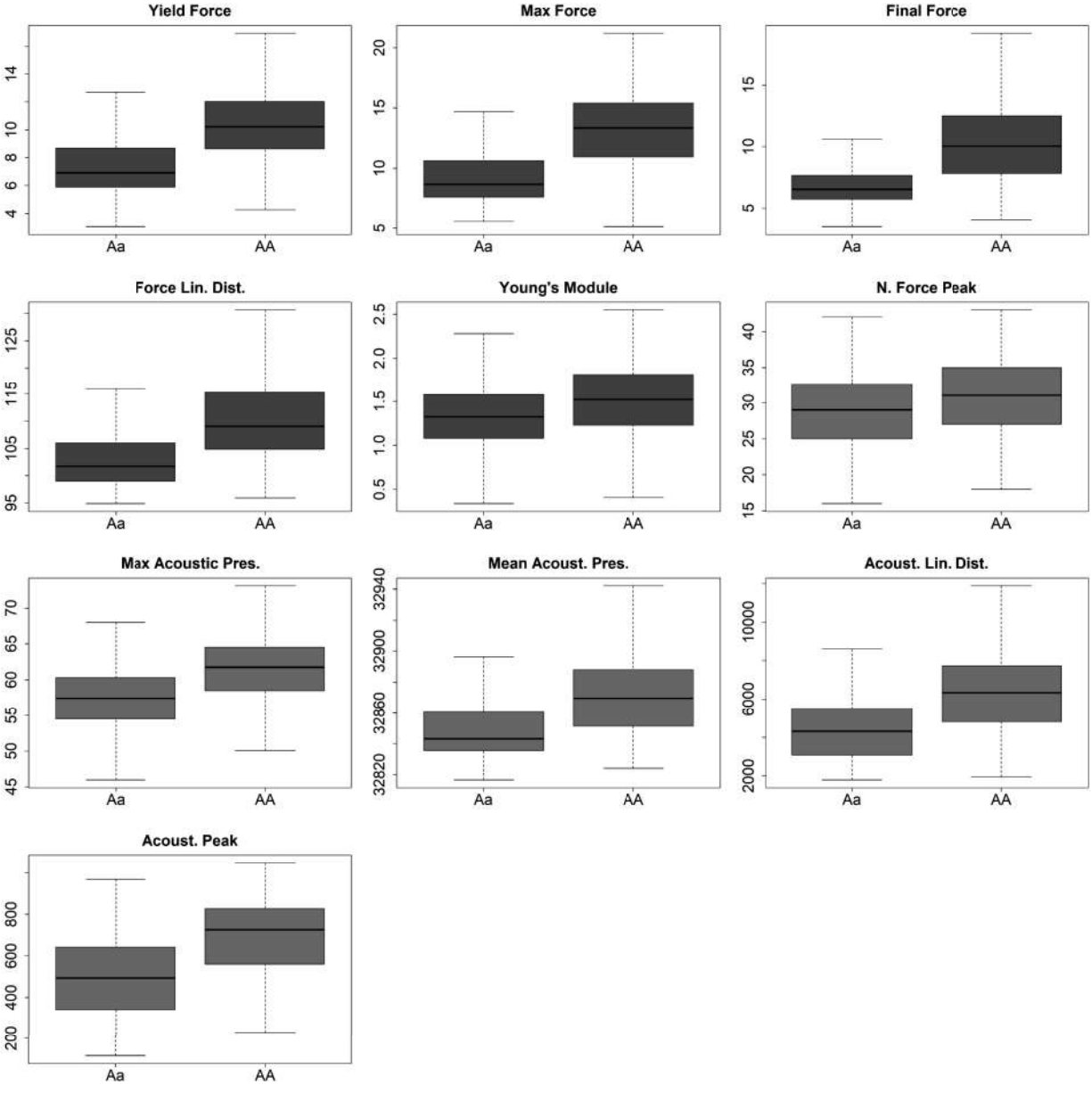
Supplementary Figure 3



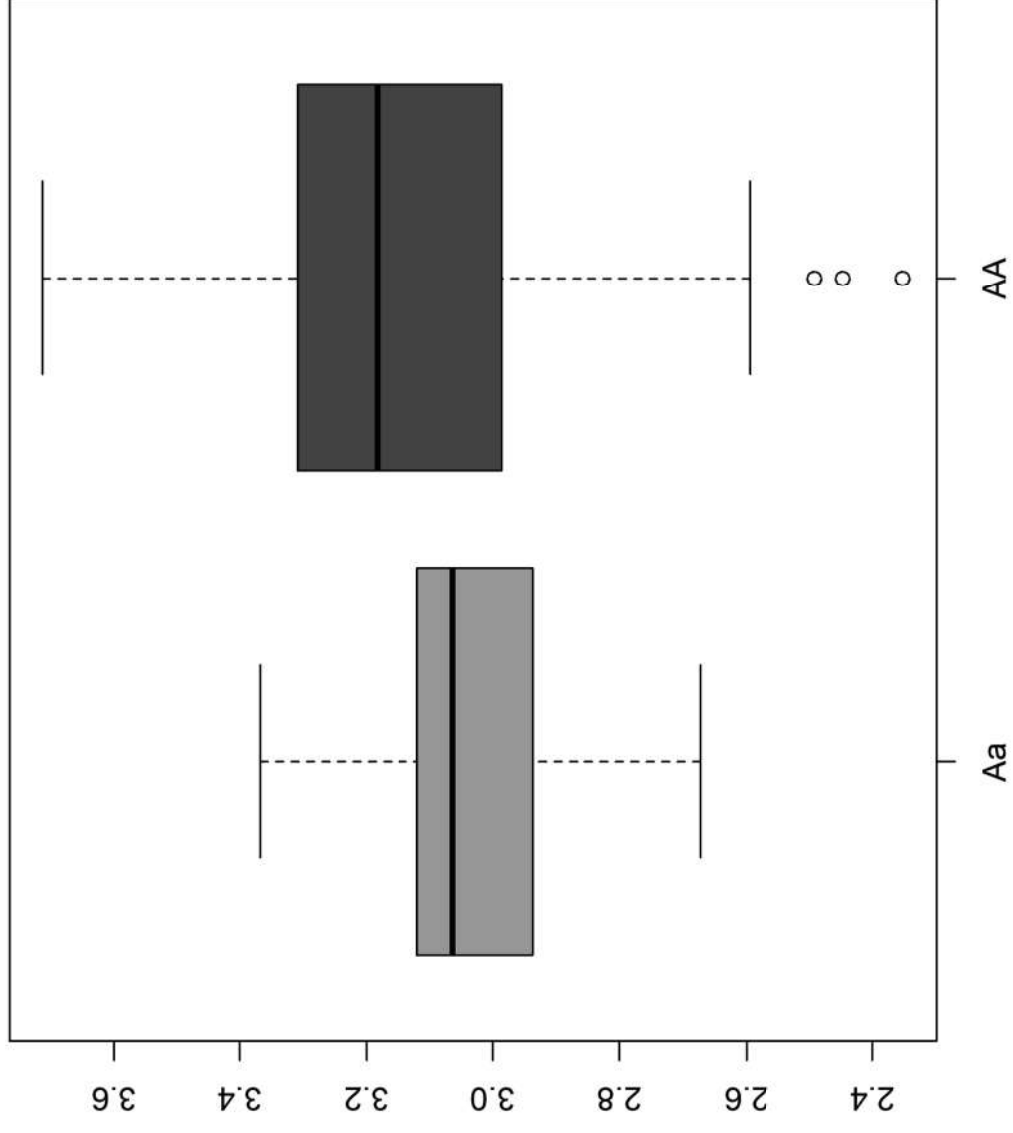
Supplementary Figure 4



Supplementary Figure 5



Supplementary Figure 6



Supplementary Table 1: Description of the size of the disks (or cubes for light microscopy) used for the phenotypic analysis. For each analysis the number of fruits and disk/fruit is reported.

	Diameter-Side [cm]	Volume [cm ³]	Fruits	Disk/fruit
Juiciness	1	0.785	3	3
Texture	1	0.785	≥5	3
Light microscopy	1	1	3	1
Micro tomography	0.6	0.226	3	1
Sensory	1.8	2.544	16	1

Supplementary Table 2: The sensory lexicon developed by the panel for descriptive analysis of apples: For each descriptor, the sensory definition is shown.

Descriptors	Sensory definition
Aroma	Overall odour sensation orto-nasally perceived (by smelling the sample)
Crispness	Sound (pitch/intensity) produced by the sample at the first bite using the incisors
Hardness	Resistance of the sample to the first chews with molars (1-2 chews without breaking it)
Juiciness	Amount of juice released during chewing the sample (first 3 chews)
Mealiness	Degree of flesh breaking in small and dry fragments/granules during chewing
Crunchiness	Sound (pitch/intensity) produced by the sample during 5 molar chews
Granularity	Numbers/size of fragments/granules produced during chewing
Fibrousness	Degree of flesh breaking during chewing in thick and fibrous fragments/granules
Flavour	Overall flavour (odour) sensation retro-nasally perceived (by tasting the sample)

Supplementary table 3: Individuals' scores of the principal component analysis performed: PCA – Texture refers to supplementary Figure 2; PCA - Texture, Juiciness, Cell Morphology refers to Figure 3A, PCA - Texture, Juiciness, Cell Morphology, Sensory analysis refers to Figure 5A

Samples	PCA - Texture		PCA - Texture, Juiciness, Cell Morphology		PCA - Texture, Juiciness, Cell Morphology, Sensory analysis	
	PC1	PC2	PC1	PC2	PC1	PC2
CIV323	1.04	1.68	1.27	1.2	-2.87	0.74
FEM16	-4.09	0.09	-3.97	1.62	4.54	1.32
Fuji	-0.70	-0.99	-0.83	-1.05	0.82	-1.17
Gala	4.33	0.8	4.48	0.95	-5.5	1.58
Golden Delicious	3.07	-0.91	3.19	-0.84	-5.2	-0.34
Gradisca	-2.28	-1.15	-2.09	-0.87	2.29	-1.07
Kizuri	-3.32	-1.79	-3.53	-2.55	2.91	-3.44
Lumaga	-0.46	2.94	-0.4	3.65	2.21	4.03
Minneiska	3.97	-1.19	3.83	-1.67	-4.81	-1.44
MN55	-2.37	-0.23	-2.65	-1.16	4.59	-1.42
Red Delicious	2.06	-1.77	2.13	-1.76	-3.04	-1.56
UEB32642	1.54	0.64	1.37	1.66	-0.62	1.9
UEB6581	-1.77	1.56	-1.81	1.61	3.15	1.36
Y102	-1	0.34	-0.99	-0.79	1.54	-0.49

Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

Poles L: Conceptualization, Methodology, Investigation, Writing - Original Draft; **Gentile A:** Investigation, Writing - Review & Editing; **Giuffrida A:** Investigation; **Valentini L:** Investigation; **Endrizzi I:** Investigation; **Aprea E:** Investigation, **Gaspero F:** Investigation, Writing - Review & Editing, Resources; **Distefano G:** Investigation, Writing - Review & Editing; **Artoli G:** Writing - Review & Editing; **La Malfa S:** Writing - Review & Editing, Resources; **Costa F:** Writing - Review & Editing; **Lovatti F:** Conceptualization, Resources, Writing - Review & Editing, Funding acquisition; **Di Guardo M:** Conceptualization, Formal analysis, Writing - Original Draft, Writing - Review & Editing, Supervision