



## Article

# Revisiting Multi-Omics Data to Unravel Galectins as Prognostic Factors in Head and Neck Squamous Cell Carcinoma

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**Abstract:** Head and Neck Squamous Cell Carcinoma (HNSCC) is a malignant cancer with a poor prognosis. Galectins (Gal) have been the subject of intensive research, but the comparative prognostic value of each Gal type is not yet understood. Therefore, a literature search for evaluating galectins as prognostic biomarkers in HNSCC was conducted. The relationship between Gal expression in HNSCC with HPV and *TP53* mutational status was assessed using the UALCAN database. The impact of these biomarkers on prognosis was analyzed using ToPP and CPPA web tools. The expression of galectins in the tumor microenvironment and the impact on prognosis depending on the cancer immune subtype were analyzed using single-cell RNA sequencing. *Gal-1* and *Gal-3BP* were shown to be promising biomarkers with a triple function for the prediction of HPV and *TP53* mutational status, stratification of the HNSCC prognosis, and prediction of the response to treatment. In addition, these two galectins have been shown to be most influenced by the tumor microenvironment of HNSCC. *Gal-1* and *Gal-3BP* are the most promising galectins in HNSCC. Furthermore, this study highlights the need for further studies to evaluate galectins in HNSCC and clarify the role of individual Gals in the patient's stratification.

**Keywords:** galectins; prognosis; multi-omics; head and neck squamous cell carcinoma



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## 1. Introduction

Head and neck squamous cell carcinoma (HNSCC) is one of the most common malignant cancers worldwide and originates from the epithelial cells of the oral cavity, pharynx, and larynx. Currently, the prognosis is based on Tumour Node Metastasis (TNM) Cancer Staging. The p16 is the only approved prognostic biomarker applied in oropharyngeal cancer. In other subtypes of HNSCC, p16 is not mandatory [1,2]. Still, there is a lack of diagnosis tools to optimize risk stratification in HNSCC.

Galectins are a family of non-integrin  $\beta$ -galactoside binding lectins encoded by *LGALS* genes. According to their structure and the number of carbohydrate-recognition domains, they can be classified into prototype, chimera type, and tandem repeat type. Galectins are involved in numerous biological processes including the tumor-stroma crosstalk, which is considered a critical hallmark that promotes the progression and metastasis of solid tumors. These lectins are key players in modulating the tumor microenvironment (TME) and in regulating tumor progression, invasion, and metastatization processes. The TME consists

of several cell types, namely, T cells, B cells, natural killer cells, macrophages, neutrophils, dendritic cells, stromal cells, endothelial cells, cancer-associated fibroblasts, adipocytes, and stellate cells. These cells interact with non-cellular components of the TME, such as extracellular matrix components and exosomes that also participate in carcinogenesis, invasion, and tumor metastasis [3,4]. The TME often induces immunosuppressive states that affect the efficacy of conventional chemotherapy treatments. Thus, in recent years, therapies targeting TME constituents have been developed to be administered alone or in combination with conventional chemotherapy treatments to increase their efficacy. To identify patient subsets that may benefit from immunotherapy, it is necessary to identify the predictive biomarkers of treatment response to optimize the therapeutic management of patients with HNSCC [3,4].

Several works have been done to evaluate the role of specific galectins in the prognosis of HNSCC [5–42]. However, there are no studies that make a systematized evaluation of galectins in HNSCC to identify the panel of galectins with greater interest to be validated as a prognostic biomarker. This work aimed to identify the best panel of galectins that can be used as biomarkers of HPV and *TP53* mutational status, prognosis, and response to treatment in patients with HNSCC. To evaluate which galectins could have the capacity to play this triple role, the expression of galectins in HNSCC tissue was analyzed in relation to HPV and *TP53* mutational status. Subsequently, the prognostic potential of each of the galectins in HNSCC was compared to identify the most promising panel of galectins as prognostic biomarkers for HNSCC. Finally, the ability of galectins to predict the response of HNSCC patients to standard treatments for this type of cancer was evaluated. This multi-dimensional approach provides insights into the intricate role of galectins in HNSCC, contributing to improving the prognosis and boosting the development of personalized treatment strategies.

## 2. Methods

### 2.1. Literature Survey of the Galectin Role in HNSCC Prognosis

A literature search was performed using a text-mining approach to ensure a comprehensive analysis of the existing literature. This search was performed in Web of Science, Scopus, and PubMed Databases between February 2022 and October 2023 using the following keywords [(galectin OR gal OR lgals) AND (squamous AND cell AND carcinoma)]. All original papers published in the last ten years were considered. From the literature search, 52 articles were selected. After analyzing the selected articles, we verified that 16 articles compared survival outcomes in HNSCC patients. These studies were integrated in our study. This literature search aimed to assess the state of the art regarding each of the galectins in the prognosis of head and neck cancer.

### 2.2. Galectins Expression and Survival in HNSCC

UALCAN Database ([ualcan.path.uab.edu/home](http://ualcan.path.uab.edu/home), accessed on 30 November 2023), Cancer Proteome and Phosphoproteome Atlas (CPPA) (<http://cppa.site/cppa/>, accessed on 30 November 2023), and ToPP Platform (<http://www.biostatistics.online/topp/index.php>, accessed on 30 November 2023) were assessed to evaluate the expression of galectins in HNSCC and healthy individuals, as well their impact on survival [43,44]. The web resources allow a comprehensive analysis of cancer Omics data (TCGA, MET500, and CPTAC). For these analyses, the “HNSCC-Head and Neck Squamous Cell Carcinoma” Dataset was used. Student’s t-test was used to calculate the *p*-value with a cutoff of 0.05.

### 2.3. Single-Cell RNA seq (scRNA-seq) Analysis of Galectins in HNSCC for Immune Cell Infiltration Analysis

Integration of the expression of each of the galectins in the HNSCC tumor microenvironment was done using the Tumor Immune Single Cell Hub 2 (TISCH2), ToPP, and canSARblack web tools [44–46]. TISCH2 (<http://tisch.comp-genomics.org/>, accessed on 30 November 2023) is a source of single-cell RNA-seq data that combines the information

extracted from 190 tumor datasets across 50 cancer types. In this work, TISCH2 was used to evaluate the expression of galectins in various HNSCC cell types. The datasets that were evaluated are described in Supplementary Table S1. Using canSARblack (<https://cansar.ai>, accessed on 30 November 2023), the expression profile of each of the galectins in HNSCC was studied according to each one of the six immune phenotypes: C1 (wound healing), C2 (IFN- $\gamma$  dominant), C3 (inflammatory), C4 (lymphocyte depleted), C5 (immunologically quiet), and C6 (TGF- $\beta$  dominant). To evaluate the impact of galectin expression of C1 and C2 immune phenotypes of HNSCC, the ToPP platform was used. Survival analyses were conducted only on these two immune subtypes because they were the phenotypes where galectins were shown to be most overexpressed. For a deeper understanding of how single nucleotide variation (SNVs) and copy number variation (CNVs) influence immune infiltration, Gene Set Cancer Analysis (GSCA; <http://bioinfo.life.hust.edu.cn/GSCA/#/>, accessed on 30 November 2023) was used. To assess an association between immune cell infiltrates and galectins, the “Immune” module was used through ImmuCellAI to realize an association of genes with about 24 types of immune cells using the Wilcoxon test (comparison of two groups) or the One-Way ANOVA test (more than two groups). The *p*-value was adjusted by FDR.

#### 2.4. Mutations and Post-Translation Modifications (PTM) for Each Galectin in HNSCC

TCGA-HNSC Dataset was used to identify CNV and SNV, and the methylation profile of the genes encoding galectins was assessed using GSCA and cBioPortal (<https://www.cbioportal.org>, accessed on 30 November 2023). cBioPortal is a web server used as a cancer genomics database based on TCGA. In this study, cBioPortal allowed a better understanding of the PTM associated with each galectin in HNSCC. The results obtained with cBioPortal were complemented with GSCA. In GSCA, the SNV data of 10,234 samples from 33 cancer types and CNV data of 11,495 samples were downloaded from TCGA Database. Methylation data of 14 cancer types were downloaded from Illumina Human Methylation 450k level 3 (TCGA Database) using the methylation module from GSCA. The Kaplan-Meier survival analysis with log-rank test in GSCA was used to assess the correlation between the galectins that present these types of gene alterations with survival. The data were analyzed using R package survival, Cox Proportional-Hazards, and Logrank test.

#### 2.5. Galectin Interaction Networks

GeneMANIA (<https://genemania.org>, accessed on 30 November 2023) is a web tool used for predicting gene function. This tool finds the most related genes to the input set of genes using a large set of functional association data. The gene-gene interaction network was constructed using this tool [47]. STRING database (<https://string-db.org>, accessed on 30 November 2023) was used for the analysis of the protein-protein interaction network. STRING provides associations between proteins using a score based on several sources such as scientific literature, experiments, computational interaction predictions, and systematic transfers of evidence among organisms [48]. REACTOME database (<https://reactome.org>, accessed on 30 November 2023) web tool allows the establishment of functional relationships between gene expression profiles, and it was used as a complementary tool for the functional enrichment of the main galectin interactors [49].

#### 2.6. Galectins as Drug Targets in HNSCC

The potential of each of the galectins as the biomarker of response to the HNSCC treatment was studied using the GSCALite web server. GSCALite (<http://bioinfo.life.hust.edu.cn/web/GSCALite/>, accessed on 30 November 2023) is a web server that allows an integrated and broad approach to a set of genes [50]. In this case, GSCALite was chosen rather than GSCA; since GSCALite provides the values of a much larger number of drugs whereas GSCA only shows the results for the 30 drugs with the highest correlation with the genes of interest. One of the modules allows the assessment of drug sensitivity for genes using the Genomics of Drug Sensitivity in Cancer (GDSC) and Cancer Therapeutics

Response Portal (CTRP) Databases. From GDSC, the inhibitory concentration 50% (IC50) of 265 small molecules in 860 cell lines and their corresponding mRNA gene expression were integrated, whereas the IC50 information of 481 small molecules in 1001 cell lines and their corresponding mRNA gene expression was extracted from CTRP. To establish the correlation between IC50 and corresponding mRNA gene expression, Pearson’s correlation was used and the *p*-value was adjusted by FDR.

### 3. Results

#### 3.1. Identification of Galectin Isoforms with Key Role in HNSCC Pathogenesis

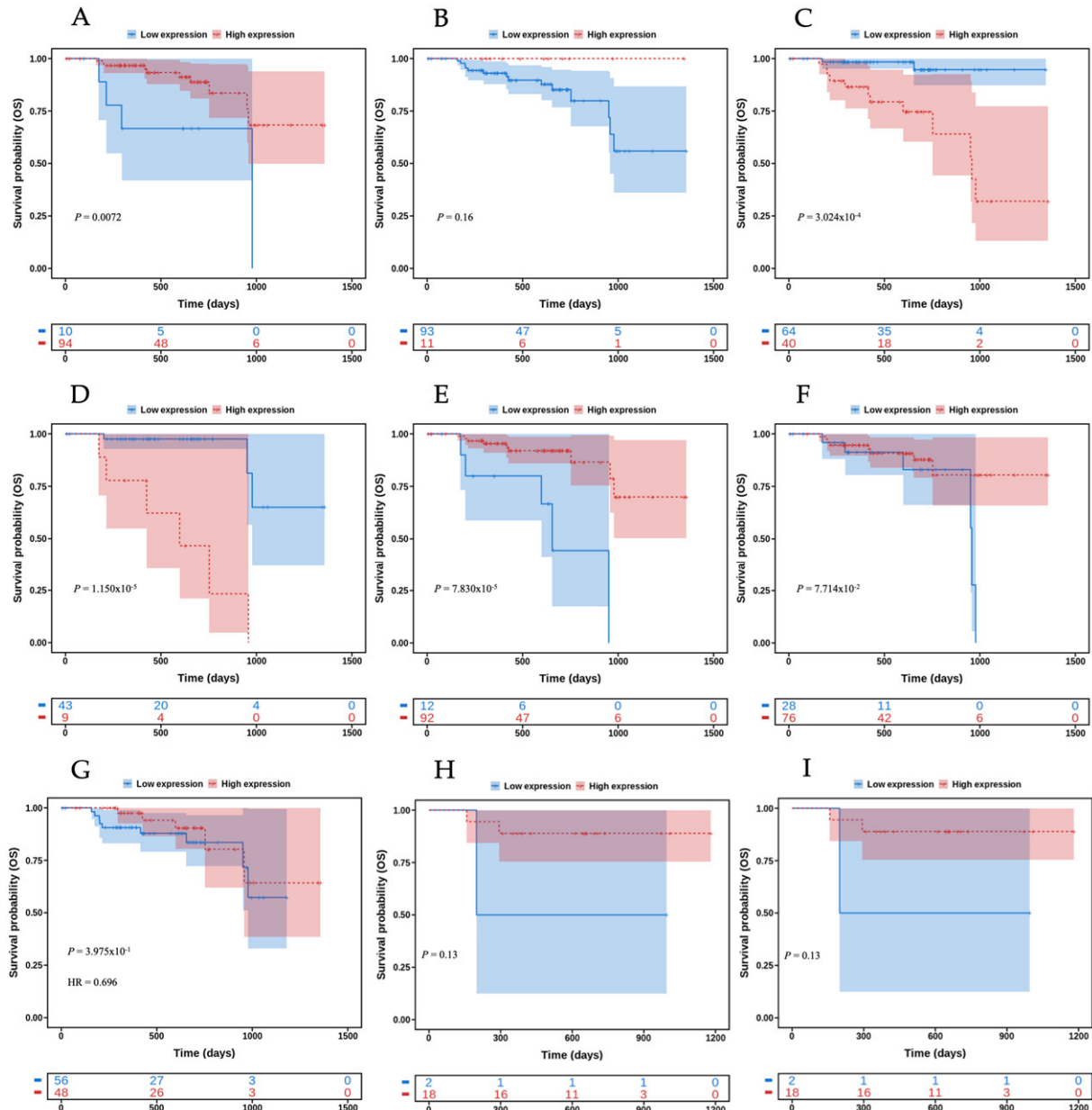
To assess the state of the art regarding the potential of galectins as prognostic biomarkers for head and neck cancer, a literature search was performed and the several survival-related parameters described in the articles were summarized in Table 1. It was possible to observe that in the existing literature, the most studied galectin was *Gal-3*. From the literature, we found a lot of heterogeneity in terms of the survival parameters analyzed. The most studied subtype of HNSCC was OSCC. By analyzing the Kaplan-Meier curves associated with each of the galectins shown in Supplementary Figure S1, we could observe that *Gal-1* and *Gal-2* have the greatest impact on the OS of patients with HNSCC. According to the TCGA analysis in UALCAN, *Gal-1* was shown to be overexpressed in HNSCC. *Gal-2* did not show significantly different expression compared to healthy individuals, as shown in Figure S2 of Figure Supplementary.

**Table 1.** Literature overview of the galectin impact on several parameters related to HNSCC prognosis. Influence of each galectin in tumor size (T), lymph node invasion (N), metastasis (M), invasion pattern (IP), relapse-free survival (RFS), disease-free survival (DFS), recurrence rate (RR), and histological grade malignancy (HGM) in head and neck squamous cell carcinoma (HNSCC). The analyzed HNSCC types are GSCC, gingival squamous cell carcinoma; LSCC, laryngeal squamous cell carcinoma; OSCC, oral squamous cell carcinoma; PSCC, palate squamous cell carcinoma; SS, sample size; TSCC, tongue squamous cell carcinoma. The studies with a statistically significant correlation (*p* < 0.05) with each one of the evaluated parameters are presented in green color and those without a statistically significant impact on survival-related parameters are presented in orange color.

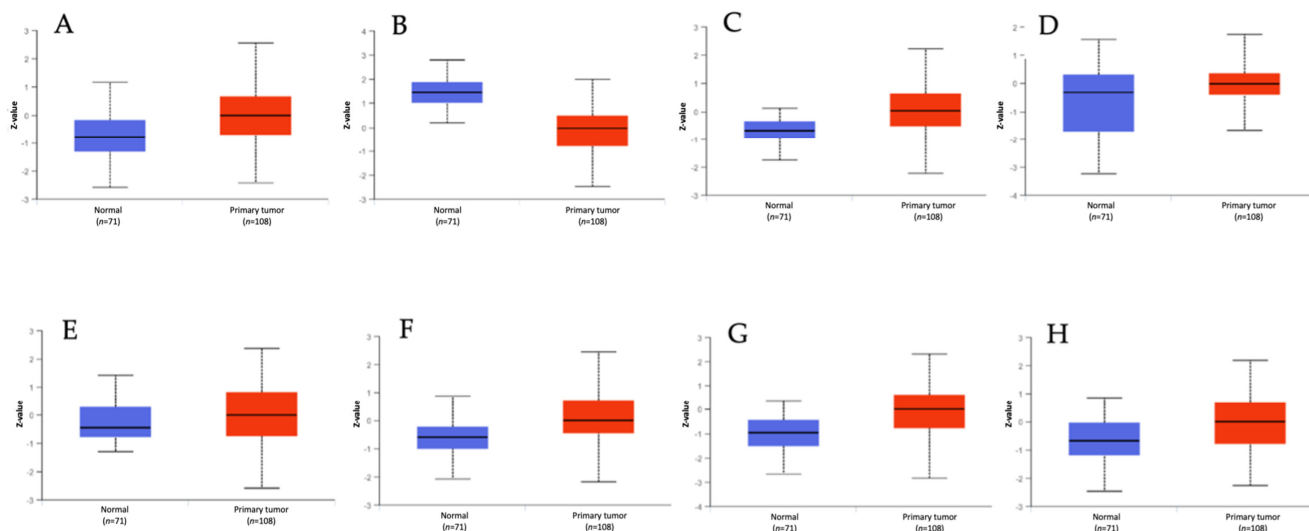
| Galectin     | Tumor Site | SS | T      | N      | M      | IP     | OS     | RFS   | DFS   | RR     | HGM    |
|--------------|------------|----|--------|--------|--------|--------|--------|-------|-------|--------|--------|
| Galectin 1   | GSCC       | 80 |        | Green  |        |        | Green  |       |       |        |        |
|              | OSCC       | 64 |        |        |        | Green  |        |       | Green |        |        |
|              | LSCC       | 62 |        |        |        |        |        |       |       | Green  |        |
|              | TSCC       | 65 |        |        | Green  |        |        |       |       |        |        |
|              | OSCC       | 60 | Green  |        |        | Green  | Orange |       |       | Orange |        |
| Galectin 3   | OSCC       | 60 | Green  |        |        | Green  | Orange |       |       | Orange |        |
|              | OSCC       | 98 | Green  |        |        | Green  |        |       |       |        |        |
|              | OSCC       | 32 | Orange |        |        |        |        |       |       |        |        |
|              | OSCC/LSCC  | 53 |        | Green  |        |        |        |       |       |        |        |
|              | LSCC       | 73 |        |        |        |        | Green  | Green |       |        |        |
| Galectin 3BP | PSCC       | 45 |        | Green  |        | Green  |        |       |       |        |        |
|              | TSCC       | 65 |        |        | Green  |        |        |       |       |        | Green  |
| Galectin 4   | OSCC       | 92 |        |        |        |        | Green  | Green |       |        |        |
| Galectin 7   | TSCC       | 65 |        |        | Orange |        |        |       |       |        | Orange |
|              | OSCC       | 32 | Green  |        |        |        |        |       |       |        |        |
| Galectin 8   | HNSCC      | 81 |        |        |        |        |        |       |       | Green  |        |
|              | TSCC       | 65 |        |        |        | Orange |        |       |       |        | Green  |
|              | HNSCC      | 93 | Orange |        |        |        |        |       |       |        |        |
| Galectin 9   | LSCC       | 77 | Green  |        |        |        |        |       |       |        |        |
|              | OSCC       | 32 |        | Orange |        |        |        |       |       |        |        |

The proteins encoded by the galectin genes were characterized in terms of their impact on survival in this patient population using CPPA as shown in Figure 1 [5–42]. In CPPA, it

could be seen that Gal-4 showed the highest HR relative to other galectins (HR = 14.848,  $p = 1.15 \times 10^{-5}$ ). Gal-3BP was the second galectin with the highest impact on OS in HNSCC (HR = 9.605,  $p = 3.024 \times 10^{-4}$ ), and its profile overlaps with the survival analysis performed in ToPP for the gene encoding Gal-4. CPTAC analysis was conducted to assess the galectin protein expression in HNSCC. We found that all proteins of interest were identified in HNSCC except for Gal-2, Gal-3BP, Gal-7C, Gal-9C, and Gal-S12, as shown in Figure 2. Using CPTAC, we analyzed the protein expression levels in HNSCC tissue compared to normal tissue, and we were able to see that for the galectins with higher impact on HNSCC survival, the following galectins Gal-1, Gal-3BP, and Gal-4 were overexpressed in HNSCC. Gal-3 was shown to be the most upregulated galectin in HNSCC (Figure 2).



**Figure 1.** Survival analysis of galectin proteins on CPPA. Overall survival of LGALS1 (A), LGALS3 (B), LGALS3BP (C), LGALS4 (D), LGALS7 (E), LGALS8 (F), LGALS9 (G), LGALS9B (H) and LGALS9C (I) proteins in HNSCC. The hazard ratios for each galectin were 0.232 (A), 0.000 (B), 14.848 (C), 0.137 (D), 0.696 (E), respectively. The X-axis represents the survival time of the patients and the Y-axis represents the probability of survival.



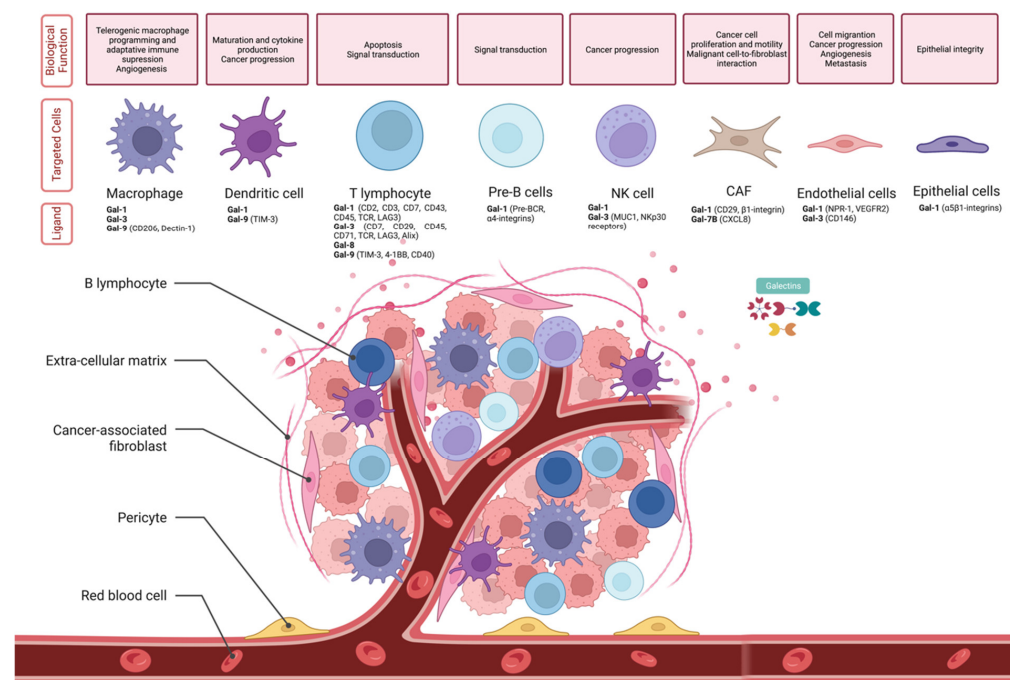
**Figure 2.** Galectin protein expression in HNSCC and healthy patients based on CPTAC. Box plot of the protein expression of LGALS1 (A), LGALS3 (B), LGALS3BP (C), LGALS4 (D), LGALS7 (E), LGALS8 (F), LGALS9 (G), and LGALS9B (H) in HNSCC ( $n = 71$ ) and healthy tissue samples ( $n = 108$ ). All results were statistically significant ( $p < 0.05$ ).

### 3.2. Identification of Galectins with Impact in the TME Remodeling

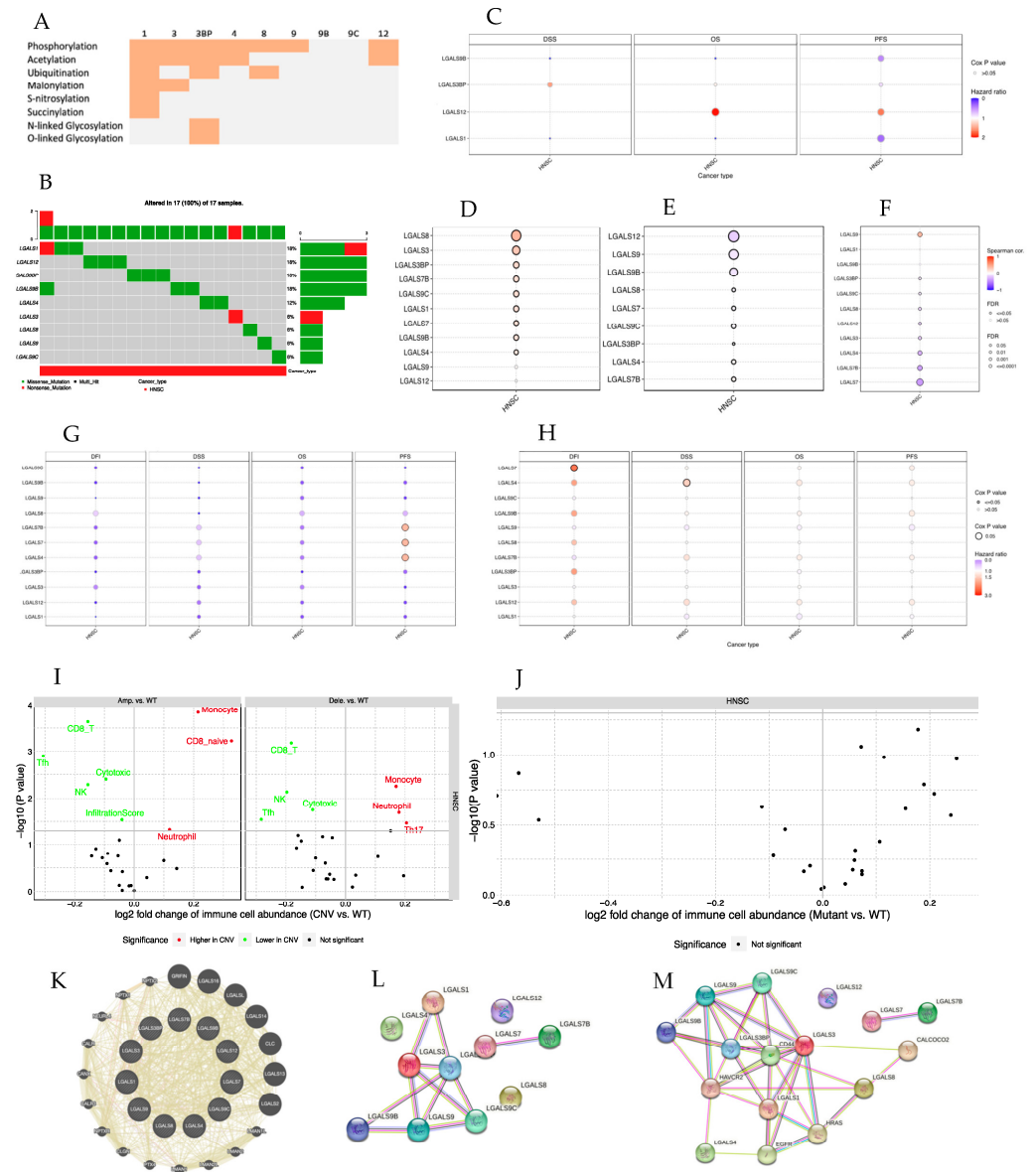
Galectins are involved in several processes related to the regulation of cell-cell interactions in the tumor microenvironment. Cancer cells can interact with various cell types that are part of the tumor microenvironment via galectins to create a more favorable environment for tumor invasion. Abnormal expression of galectins is associated with tumor development. To better understand which ligands and cell types bind to the galectins to support this favorable response to tumor progression, Figure 3 summarizes the key players in this response. When studying the immune subtypes of HNSCC, we found that Gal-1, Gal-3, Gal-3BP, Gal-4, and Gal-9 were mostly expressed in the IFN- $\gamma$  Dominant (C2) immune subtype and to a lesser extent in the Wound Healing (C1) immune subtype, as shown in Table 2. Interestingly, the first four galectins were also the ones with a major impact on survival. Subsequently, the expression of galectins in the tumor microenvironment (TME) was characterized to understand how the TME can determine the expression of each one of the galectins. The results extracted from the TISCH2 web tool for each galectin and each GEO dataset are represented in Supplementary Figures S3–S9 and the main results are illustrated in Supplementary Figure S10. Among all galectins evaluated, Gal-1 had the highest expression levels in the evaluated cell types, including Tprolif, Treg, TCD8, monocytes/macrophages, dendritic cells, natural killer (NK) cells, myofibroblasts, and fibroblasts. Gal-3 and Gal-3BP were upregulated mostly in TCD8 cells, malignant cells, myofibroblasts, and fibroblasts. Gal-9 appeared selectively upregulated in dendritic cells and monocytes/macrophages, over other galectins that were unchanged. When assessing the impact of galectin expression in HNSCC according to immune subtype, it was observed that galectin expression in HNSCC cells with an IFN- $\gamma$  Dominant (C2) profile had the highest impact on survival. Among the galectins expressed in HNSCC cells with this immune phenotype, Gal-1, Gal-3BP, and Gal-8 were the galectins showing a higher HR value in survival analyses, as shown in Supplementary Figure S11.

The most frequent PTM among all galectins was phosphorylation and the galectin that was targeted the most by PTM was Gal-1, as shown in Figure 4A. When analyzing the mutational profile of each of the galectins in HNSCC (Supplementary Table S2 and Figure 4B), it was possible to observe that the most mutated galectins in HNSCC were Gal-3BP, Gal-12, Gal-1, and Gal-4. The results for the SNV impact on HNSCC survival were not statistically significant (Figure 4C). Of these mutations, most are missense. Gal-8 transcription levels were the most influenced by CNV (Figure 4D). Regarding the methylation profile anal-

ysis of galectin genes in HNSCC, *Gal-12* was the most differentially methylated galectin in HNSCC compared to healthy individuals (Figure 4E). The galectin whose expression was most affected by the methylation process is *Gal-7* (Figure 4F). In Figure 4I, we can observe that in the HNSCC wild type group, there is a predominance of monocytes and neutrophils, whereas in the groups expressing galectins that present CNV alterations, there is a predominance of CD8 T cells, NK cells, follicular helper T CD4 cells, and cytotoxic T cells. When the groups in which there is gene amplification are compared to the group in which there is gene deletion, the main difference is the inversion of the proportion of follicular T CD4 cells and NK cells. In the group with galectin gene amplification, the proportion of follicular helper T CD4 cells is higher than NK cells, whereas in the group with galectin gene deletion, there are more NK cells. In Figure 4J, it was possible to observe that in the group of galectins presenting SNV, the immune cell abundance was lower than that in the wild type. The galectins with CNV changes that showed a higher HR in terms of PFS were *Gal-4*, *Gal-7*, and *Gal-7B* (Figure 4G). When the differences in survival were compared according to the methylation profile of each of the galectins, it was possible to observe that the methylation of *Gal-7* was shown to correlate with the highest HR of the galectins studied in terms of DFI (Figure 4H). Furthermore, we used GeneMANIA to obtain potential interaction genes of the galectin family. Figure 4K shows that the galectin genes interact with several genes of the pentraxin family that encode acute-phase inflammatory proteins. The protein-protein networks are shown in Figure 4L, M. It could be seen that the proteins that bind most strongly to the galectins are CD44 (CD44 antigen), HRAS (GTPase HRas), EGFR (Epidermal growth factor receptor), HAVCR2 (Hepatitis A virus cellular receptor 2), and CALCOCO2 (Calcium-binding and coiled-coil domain-containing protein 2). By analyzing these 5 interactors using the REACTOME database, it was possible to observe that they essentially intervene in the EGFR and ERBB2 signaling pathways. *Gal-1* and *Gal-3BP* showed to be the most interesting galectins in HNSCC. These galectins are involved in several processes such as migration, invasion, and angiogenesis of HNSCC, as described in Figure 5.

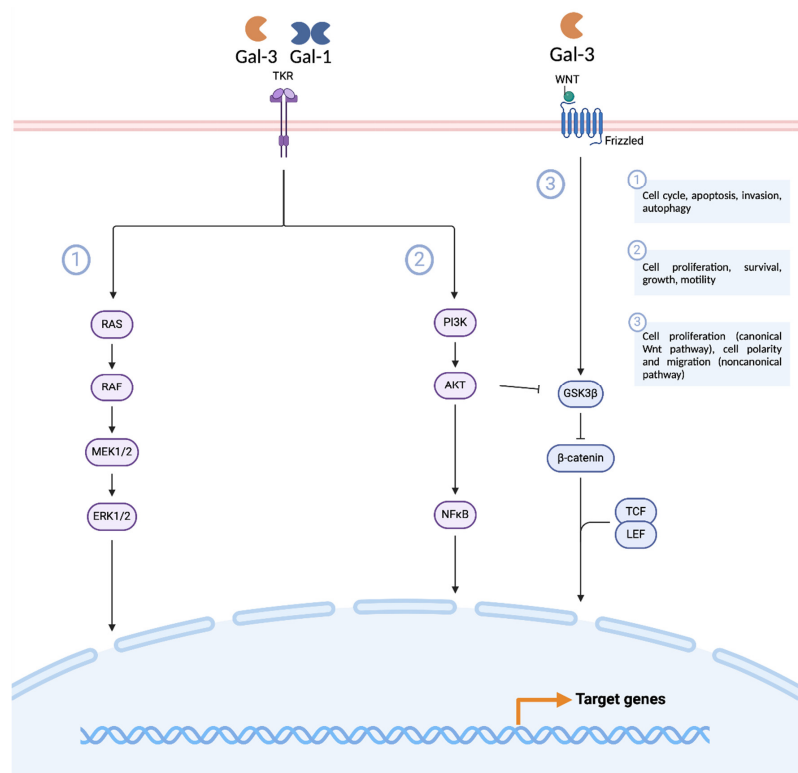


**Figure 3.** Relationship of galectins with tumor microenvironment. Biological function of galectins in each of the major cell types present in the tumor microenvironment [51–102]. Legend: CAF, cancer-associated fibroblasts; Gal-1, galectin-1; Gal-3, galectin-3; Gal-9, galectin-9; and NK cell, natural killer cell. This image was created in BioRender (Toronto, ON, Canada).



**Figure 4.** SNV, CNV, and PTM alterations of the galectins in HNSCC and their impact on survival. PTM of each galectin in HNSCC using cBioPortal (A). SNV frequency for each galectin in HNSCC using GSCALite (B). Survival analysis for galectins SNV in GSCA (C). Pearson correlation between expression of galectins and the corresponding CNV (D). Differential methylation between HNSCC and healthy tissue samples using GSCALite (E). Pearson correlation between expression of galectins and their methylation profile using GSCALite (F). Survival analysis for galectins CNV in GSCA (G). Survival analysis for high and low methylation groups in GSCA (H). Difference of immune infiltration between CNV of galectins and wild type HNSCC in GSCA (I). Difference of immune infiltration between SNV of galectins and wild type HNSCC in GSCA (J). Gene-gene interaction network of galectin of interest in GeneMANIA (K). Protein-protein interaction network analysis of galectins with STRING (L). Protein-protein interaction expanded network analysis of galectins with STRING (M). In GSCALite, only statistically significant results ( $p < 0.05$ ) are represented. In the scatter plots represented in (I,J), the  $p$ -value statistically significant is represented with green color, and the FDR significant results are represented with red color. The set of genes used in GSCA and GSCALite analysis were LGALS1, LGALS12, LGALS3, LGALS3BP, LGALS4, LGALS7, LGALS7B, LGALS8, LGALS9, LGALS9B, LGALS9C, and LGALS12.





**Figure 5.** Overview of the main pathways modulated by the galectins, Gal-1 and Gal-3, found altered in HNSCC. Interaction with the PI3K/Akt pathway is responsible for the migration and proliferation of tumor cells through the activation of β-catenin. When Gal-1 and Gal-3 interact with TKR, they stimulate the H-Ras pathway and promote cell proliferation. Legend: Akt, protein kinase B; ERK, extracellular signal-regulated protein kinase; GSK3β, glycogen synthase kinase 3 beta; ILK, integrin-linked kinase; LEF, lymphoid enhancer-binding factor; MEK, mitogen-activated protein kinase; NF-κB, nuclear factor kappa-light-chain-enhancer of activated B cells; PI3K, phosphoinositide 3-kinase; RAF, rapidly accelerated fibrosarcoma; RAS, rat sarcoma; TCF, T cell factor; and WNT, Wingless-related integration site. This image was created in BioRender (Toronto, ON, Canada).

**Table 2.** Expression of galectins in each immune subtype in HNSCC Dataset in canSAR.ai.

| Cancer Immune Subtype      | Gal-1  | Gal-3  | Gal-3BP | Gal-4  | Gal-7, 7B | Gal-8 | Gal-9  | Gal-9B | Gal-9C | Gal-12 |
|----------------------------|--------|--------|---------|--------|-----------|-------|--------|--------|--------|--------|
| Wound Healing (C1)         | 29/128 | 5/128  | 0/128   | 8/128  | 0/128     | 1/128 | 1/128  | 0/128  | 0/128  | 0/128  |
| IFN-γ Dominant (C2)        | 93/379 | 13/379 | 10/379  | 14/379 | 0/379     | 1/379 | 25/379 | 0/379  | 0/379  | 0/379  |
| Inflammatory (C3)          | 0/2    | 0/2    | 0/2     | 0/2    | 0/2       | 0/2   | 0/2    | 0/2    | 0/2    | 0/2    |
| Lymphocyte Depleted (C4)   | 0/2    | 0/2    | 0/2     | 0/2    | 0/2       | 0/2   | 0/2    | 0/2    | 0/2    | 0/2    |
| Immunologically Quiet (C5) | 0/0    | 0/0    | 0/0     | 0/0    | 0/0       | 0/0   | 0/0    | 0/0    | 0/0    | 0/0    |
| TGF-β Dominant (C6)        | 1/3    | 0/3    | 0/3     | 0/3    | 0/3       | 0/3   | 0/3    | 0/3    | 0/3    | 0/3    |

Legend: 1, galectin-1; 3, galectin-3; 3BP, galectin-3BP; 4, galectin-4; 7, galectin-7; 7B, galectin-7B; 8, galectin-8; 9, galectin-9; 9B, galectin-9B; 9C, galectin-9C; and 12, galectin-12. The values in the table refer to the number of samples where the expression of each of the galectins is seen relative to the total number of samples of each immune phenotype in HNSCC.

### 3.3. Galectins and HNSCC Response to Therapeutics

According to the National Cancer Institute, there are several drugs approved for head and neck cancer, such as bleomycin sulfate, cetuximab, docetaxel, hydroxyurea, methotrexate sodium, pembrolizumab, and nivolumab. Carboplatin with docetaxel and cisplatin with docetaxel and 5-fluorouracil are approved as drug combinations. To evaluate the correlation between the galectins and drugs present in the GDSC and CTRP databases, GSCALite was used. A summary of galectin interaction results with FDA-approved drugs for HNSCC is shown in Table 3. Bleomycin showed a negative Pearson’s correlation with *Gal-1* and cetuximab showed a negative correlation with *Gal-1*, *Gal-3*, and *Gal-3BP* expression. 5-fluorouracil, methotrexate, and paclitaxel were positively correlated with *Gal-3* and *Gal-3BP* expression. 5-fluorouracil and paclitaxel were also positively correlated with *Gal-8* expression.

**Table 3.** Galectin-drug interaction using GDSC and CTRP Databases in GSCALite. Legend: CTRP, Genomics of Drug Sensitivity in Cancer; FDR, False Discovery Rate; and CTRP, Cancer Therapeutics Response Portal. Blue represents a negative Spearman correlation between the gene and the chosen drug. Red represents a positive Spearman correlation. The results shown in this figure were statistically significant ( $p < 0.05$ ).

| Databases | Drugs          | LGALS1                       | LGALS3                       | LGALS3BP                     | LGALS4                       | LGALS8                       | LGALS9 | LGALS12 |
|-----------|----------------|------------------------------|------------------------------|------------------------------|------------------------------|------------------------------|--------|---------|
| GDSC      | Bleomycin      | log <sub>10</sub> (FDR) = 20 |                              |                              |                              |                              |        |         |
|           | 5-fluorouracil |                              | log <sub>10</sub> (FDR) = 10 | log <sub>10</sub> (FDR) = 10 |                              |                              |        |         |
|           | Methotrexate   |                              | log <sub>10</sub> (FDR) = 40 | log <sub>10</sub> (FDR) = 40 |                              | log <sub>10</sub> (FDR) = 10 |        |         |
|           | Docetaxel      | log <sub>10</sub> (FDR) = 20 | log <sub>10</sub> (FDR) = 10 | log <sub>10</sub> (FDR) = 10 |                              |                              |        |         |
|           | Cetuximab      |                              | log <sub>10</sub> (FDR) = 10 |                              |                              |                              |        |         |
| CTRP      | Docetaxel      |                              | log <sub>10</sub> (FDR) = 20 | log <sub>10</sub> (FDR) = 20 | log <sub>10</sub> (FDR) = 10 |                              |        |         |
|           | Paclitaxel     |                              | log <sub>10</sub> (FDR) = 30 | log <sub>10</sub> (FDR) = 30 |                              | log <sub>10</sub> (FDR) = 10 |        |         |

### 4. Discussion

In the present work, the role of galectins was comprehensively studied to compare the contribution of each galectin in the processes of tumor progression and invasion, as well as the added value they may have in the early diagnosis of HNSCC and the evaluation of treatment response. To this end, a literature search was initially conducted to analyze the evidence that exists for these proteins in HNSCC. Most of the published articles focused on the impact of galectin expression on prognosis, with *Gal-3* being the one with the most literature evidence. Taking into account the existing information for *Gal-3* in HNSCC, a systematic review was done in PROSPERO with the following number CRD42023400863. Subsequently, a bioinformatics analysis was conducted, and the expression levels and impact on the prognosis of genes encoding galectins and proteins were assessed using platforms such as ToPP, UALCAN, and CPPA. Based on the results obtained, it was possible to observe that *Gal-1*, *Gal-3*, *Gal-3BP*, and *Gal-4* proved to be the most impactful galectins in the prognosis of HNSCC. The results obtained from the bioinformatics analysis provide supporting evidence for the existing literature, while also paving the way for future research. One of the least studied galectins, *Gal-3BP*, has been found to have a significant impact on the prognosis of HNSCC. The *LGALS3BP* gene exhibited an HR = 2.6 and the *Gal-3BP* protein an HR = 9.605 when evaluating the impact of their expression on the OS of HNSCC patients. High levels of *Gal-3* expression in HNSCC were associated with increased progression, invasiveness, and aggressiveness in part by the activation of pathways such as

the Wnt/B-catenin, ERK1/2, and AKT pathways. *Gal-3* expression is dependent on the cell differentiation process in both normal and neoplastic cells [12,21–23,32,34,38,40,41,103,104]. Colocalization of Gal-3 with desmosomal proteins demonstrated that this protein may play an important role at the cell surface level in mediating intercellular contacts between tumor cells. Thus, it can be used to monitor the degree of cell differentiation in carcinomas that have their genesis in neoplastic transformation cells [11,13,30]. Studies have shown that serum levels of Gal-3 are increased in individuals with HNSCC compared to those of controls, and its expression in individuals with risk factors for this type of cancer has been associated with an approximately 3-fold increased risk of developing HNSCC [6,105]. Thus, it has the potential to be used in HNSCC screening using serum as a source for liquid biopsy. Regarding the treatment, Gal-3 inhibition has shown potential as a therapeutic option in these patients, especially in HPV-driven HNSCC which presents a much higher Gal-3 expression than in HPV-non-driven HNSCC patients, due to the inhibition of this protein particularly interesting in this cancer subset [37]. *Gal-3* seems to have an immunosuppressive effect by promoting M2 macrophage polarization, inducing lymphocyte apoptosis, and inhibiting T-cell activation that contributes to tumor progression. The goal of Gal-3 inhibitors is to restore the immune response that is suppressed by Gal-3 [37,106]. On [ClinicalTrials.gov](https://clinicaltrials.gov), there are 3 clinical trials with Gal-3 inhibitors for the treatment of HNSCC, namely, NCT00054977, NCT02575404, and NCT04987996. Clinical trial NCT00054977 is a phase I, open-label, and non-randomized trial that has been completed and aimed to study the safety of GM-CT-01 alone and in combination with 5-fluorouracil in HNSCC patients. The results have not been published. NCT04987996 is a phase II randomized trial that aims to evaluate the safety and efficacy of Gal-3 inhibitor, GR-MD-02, compared with pembrolizumab in the treatment of HNSCC. This study is currently on hold. NCT02575404 is an ongoing phase I, open-label, and non-randomized clinical trial that aims to evaluate the dose escalation of GR-MD-02 with pembrolizumab in patients with HNSCC. More studies are still needed, but the development of Gal-3 inhibitors that can be administered alone or in conjunction with standard HNSCC treatments to increase their effectiveness is shown to be a promising strategy. By studying the role of each of the galectins in the tumor microenvironment, *Gal-1*, *Gal-3*, and *Gal-3BP* have been shown to be highly overregulated in their expression levels in the major cells that constitute the tumor microenvironment. The development of drugs that inhibit the overexpression of these 3 galectins will allow modulation of the tumor microenvironment, increasing the sensitivity of patients to immunotherapy treatments.

Studies have shown that the high expression of Gal-3BP, compared to other galectins, is associated with a worse prognosis, including lower overall survival, disease-free survival, and relapse-free survival. This protein's involvement in the PI3K/AKT pathway contributes to its impact on tumor progression [107]. Gal-1 is a prototypical galectin that modulates the process of differentiation, immune escape, and tumor progression. Gal-1 levels are modulated by blood oxygen levels. When in hypoxia, Gal-1 expression increases, and this increased expression is associated with the secretion of proteins that modulate the immune response, assuming an active role in the malignant progression and therapeutic response of HNSCC [9]. Considering the protective effect that Gal-1 exerts on cancer cells against the action of the immune system through its deleterious effect on activated T-cells and the activation of oncogenic H-Ras proteins, the ways to reduce its expression to increase the efficiency of T-cell mediated immunotherapy in patients with HNSCC have also been evaluated [9,26,31]. Therefore, high levels of Gal-1 are associated with a worse prognosis of HNSCC. Gal-1 is involved in tumor invasion and metastasis processes. This role is due in part to the increase in certain metalloproteinases in response to increased Gal-1 expression levels and the fact that it is involved in the reorganization of the cytoskeleton in oral cancer [14,16,29,35]. Increased Gal-7 levels have been shown to be associated with tumor progression, a higher recurrence rate, and a worse prognosis of HNSCC [10,18,42]. Gal-8 and Gal-9 were shown to have the potential to differentiate HNSCC from other potentially malignant oral lesions and healthy tissues [17,27,28,60]. The work done in this

study allowed a more comprehensive and detailed characterization of the biomarkers, thereby identifying those with the greatest potential for translation into clinical practice.

## 5. Conclusions

Galectins play a very important role in tumor differentiation, progression, and invasion, being associated with a worse prognosis of HNSCC. *Gal-3* and *Gal-1* are the galectins with more literature evidence pointing to their potential as prognostic biomarkers. From the bioinformatics analysis, *Gal-1* and *Gal-3BP* were shown to be strongly modulated by HPV, *TP53* mutational status, and tumor microenvironment. In addition, they have been shown to be valuable biomarkers of prognosis and treatment response in patients with HNSCC. Future research should prioritize elucidating the intricate role of *Gal-3BP* in HNSCC, as well as investigating the link between the expression of each of the galectins and HPV and *TP53* mutational status. Such investigations hold significant promise for advancing our understanding of HNSCC pathogenesis, improving prognostic assessment, and paving the way for targeted therapeutic interventions in this complex malignancy.

**Supplementary Materials:** The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/biomedicines12030529/s1>, Figure S1: Survival analysis in ToPP Database. Kaplan-Meier curves for patient prognosis when divided by LGALS1 (A), LGALS2 (B), LGALS3 (C), LGAL3BP (D), LGALS4 (E), LGALS7 (F), LGALS7B (G), LGALS8 (H), LGALS9 (I), LGALS9B (J), LGALS9C (L), and LGALS12 (M) expression levels. Black data points represent patients whose HNSCC tumors had a galectin expression below the median level. Red data points represent patients whose HNSCC tumors had a galectin expression above the median level; Figure S2: Differential expression of galectins in HNSCC patients and healthy individuals in ToPP using TCGA Dataset. Box plot of expression of LGALS1 (A), LGALS2 (B), LGALS3 (C), LGAL3BP (D), LGALS4 (E), LGALS7 (F), LGALS7B (G), LGALS8 (H), LGALS9 (I), LGALS9B (J), LGALS9C (K), and LGALS12 (L) in HNSCC (red) and healthy tissue samples (blue). Legend: \*,  $p < 0.05$ ; \*\*,  $p < 0.01$ ; \*\*\*,  $p < 0.001$ ; and NS, not significant; Figure S3: Expression of galectins in GSE150321 Dataset in TISCH2. The color intensity is proportional to the expression of each of the galectins of interest; Figure S4: Expression of galectins in GSE103322 Dataset in TISCH2. The color intensity is proportional to the expression of each of the galectins of interest; Figure S5: Expression of galectins in GSE139324 Dataset in TISCH2. The color intensity is proportional to the expression of each of the galectins of interest; Figure S6: Expression of galectins in GSE150430 Dataset in TISCH2. The color intensity is proportional to the expression of each of the galectins of interest; Figure S7: Expression of galectins in GSE162025 Dataset in TISCH2. The color intensity is proportional to the expression of each of the galectins of interest; Figure S8: Expression of galectins in GSE172577 Dataset in TISCH2. The color intensity is proportional to the expression of each of the galectins of interest; Figure S9: Expression of galectins in GSE180268 Dataset in TISCH2. The color intensity is proportional to the expression of each of the galectins of interest; Figure S10: Expression of galectins in several types of immune cells from HNSCC Dataset using TISCH2. Legend: 1, galectin-1; 3, galectin-3; 3BP, galectin-3BP; 4, galectin-4; 7, galectin-7; 7B, galectin-7B; 8, galectin-8; 9, galectin-9; 9B, galectin-9B; 9C, galectin-9C; 12, galectin-12; B, B cell; DC, dendritic cells; mono/macro, monocyte/macrophage; NK cell; natural killer cell; Tprolif, proliferating T cell; and Treg, regulatory T cell; Figure S11: Survival analysis for each galectin in C1 and C2 immune phenotypes of HNSCC Dataset in ToPP. Abbreviations: \*,  $p < 0.05$ ; \*\*,  $p < 0.01$ ; \*\*\*,  $p < 0.001$ ; 1, galectin-1; 3, galectin-3; 3BP, galectin-3BP; 4, galectin-4; 7, galectin-7; 7B, galectin-7B; 8, galectin-8; 9, galectin-9; 9B, galectin-9B; 9C, galectin-9C; 12, galectin-12; DFS, disease free survival; DSS, disease specific survival; HR; Hazard Ratio; OS, overall survival; and RFS, relapse free survival; Table S1: GEO Datasets of HNSCC used in TISCH2 analysis; Legend: HNSC, head and neck squamous cell carcinoma; LSCC, laryngeal squamous cell carcinoma; NPC, nasopharyngeal carcinoma; OSCC, oropharyngeal squamous cell carcinoma; PMID, and PubMed Unique Identifier; Table S2: Mutations of galectins in cBioPortal. Legend: N/A, non-available information.

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## References

- Machiels, J.P.; Rene Leemans, C.; Golusinski, W.; Grau, C.; Licitra, L.; Gregoire, V.; EHNS Executive Board; ESMO Guidelines Committee; ESTRO Executive Board. Squamous cell carcinoma of the oral cavity, larynx, oropharynx and hypopharynx: EHNS-ESMO-ESTRO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann. Oncol. Off. J. Eur. Soc. Med. Oncol.* **2020**, *31*, 1462–1475. [[CrossRef](#)] [[PubMed](#)]
- Pfister, D.G.; Spencer, S.; Adelstein, D.; Adkins, D.; Anzai, Y.; Brizel, D.M.; Bruce, J.Y.; Busse, P.M.; Caudell, J.J.; Cmelak, A.J.; et al. Head and Neck Cancers, Version 2.2020, NCCN Clinical Practice Guidelines in Oncology. *J. Natl. Compr. Cancer Netw. JNCCN* **2020**, *18*, 873–898. [[CrossRef](#)]
- Anderson, N.M.; Simon, M.C. The tumor microenvironment. *Curr. Biol. CB* **2020**, *30*, R921–R925. [[CrossRef](#)] [[PubMed](#)]
- DeBerardinis, R.J. Tumor Microenvironment, Metabolism, and Immunotherapy. *N. Engl. J. Med.* **2020**, *382*, 869–871. [[CrossRef](#)]
- Cludts, S.; Decaestecker, C.; Mahillon, V.; Chevalier, D.; Kaltner, H.; Andre, S.; Rummelink, M.; Leroy, X.; Gabius, H.J.; Saussez, S. Galectin-8 up-regulation during hypopharyngeal and laryngeal tumor progression and comparison with galectin-1, -3 and -7. *Anticancer. Res.* **2009**, *29*, 4933–4940. [[PubMed](#)]
- Aggarwal, S.; Sharma, S.C.; Das, S.N. Galectin-1 and galectin-3: Plausible tumour markers for oral squamous cell carcinoma and suitable targets for screening high-risk population. *Clin. Chim. Acta Int. J. Clin. Chem.* **2015**, *442*, 13–21. [[CrossRef](#)]
- Xu, X.C.; Sola Gallego, J.J.; Lotan, R.; El-Naggar, A.K. Differential expression of galectin-1 and galectin-3 in benign and malignant salivary gland neoplasms. *Int. J. Oncol.* **2000**, *17*, 271–276. [[CrossRef](#)]
- Dong, G.W.; Kim, J.; Park, J.H.; Choi, J.Y.; Cho, S.I.; Lim, S.C. Galectin-8 expression in laryngeal squamous cell carcinoma. *Clin. Exp. Otorhinolaryngol.* **2009**, *2*, 13–19. [[CrossRef](#)]
- Le, Q.T.; Shi, G.; Cao, H.; Nelson, D.W.; Wang, Y.; Chen, E.Y.; Zhao, S.; Kong, C.; Richardson, D.; O'Byrne, K.J.; et al. Galectin-1: A link between tumor hypoxia and tumor immune privilege. *J. Clin. Oncol. Off. J. Am. Soc. Clin. Oncol.* **2005**, *23*, 8932–8941. [[CrossRef](#)]
- Mesquita, J.A.; Queiroz, L.M.; Silveira, E.J.; Gordon-Nunez, M.A.; Godoy, G.P.; Nonaka, C.F.; Alves, P.M. Association of immunoexpression of the galectins-3 and -7 with histopathological and clinical parameters in oral squamous cell carcinoma in young patients. *Eur. Arch. Oto-Rhino-Laryngol. Off. J. Eur. Fed. Oto-Rhino-Laryngol. Soc. EUFOS Affil. Ger. Soc. Oto-Rhino-Laryngol.-Head Neck Surg.* **2016**, *273*, 237–243. [[CrossRef](#)]
- Alves, P.M.; Godoy, G.P.; Gomes, D.Q.; Medeiros, A.M.; de Souza, L.B.; da Silveira, E.J.; Vasconcelos, M.G.; Queiroz, L.M. Significance of galectins-1, -3, -4 and -7 in the progression of squamous cell carcinoma of the tongue. *Pathol. Res. Pract.* **2011**, *207*, 236–240. [[CrossRef](#)]
- Patru, A.; Surlin, V.; Margaritescu, C.; Ciuca, E.M.; Matei, M.; Dumitrescu, D.; Camen, A. Immunohistochemical evaluation of D2-40, Galectin-3, Maspin and MCM7 expression in palate squamous cell carcinomas. *Rom. J. Morphol. Embryol. Rev. Roum. Morphol. Embryol.* **2021**, *62*, 133–149. [[CrossRef](#)]
- Plzak, J.; Smetana, K., Jr.; Hrdlickova, E.; Kodet, R.; Holikova, Z.; Liu, F.T.; Dvorankova, B.; Kaltner, H.; Betka, J.; Gabius, H.J. Expression of galectin-3-reactive ligands in squamous cancer and normal epithelial cells as a marker of differentiation. *Int. J. Oncol.* **2001**, *19*, 59–64. [[CrossRef](#)] [[PubMed](#)]
- Wu, M.H.; Hong, T.M.; Cheng, H.W.; Pan, S.H.; Liang, Y.R.; Hong, H.C.; Chiang, W.F.; Wong, T.Y.; Shieh, D.B.; Shiau, A.L.; et al. Galectin-1-mediated tumor invasion and metastasis, up-regulated matrix metalloproteinase expression, and reorganized actin cytoskeletons. *Mol. Cancer Res. MCR* **2009**, *7*, 311–318. [[CrossRef](#)]
- Saussez, S.; Decaestecker, C.; Lorfevre, F.; Chevalier, D.; Mortuaire, G.; Kaltner, H.; Andre, S.; Toubeau, G.; Gabius, H.J.; Leroy, X. Increased expression and altered intracellular distribution of adhesion/growth-regulatory lectins galectins-1 and -7 during tumour progression in hypopharyngeal and laryngeal squamous cell carcinomas. *Histopathology* **2008**, *52*, 483–493. [[CrossRef](#)] [[PubMed](#)]

16. Chang, S.L.; Li, C.F.; Lin, C.; Lin, Y.S. Galectin-1 overexpression in nasopharyngeal carcinoma: Effect on survival. *Acta Oto-Laryngol.* **2014**, *134*, 536–542. [[CrossRef](#)]
17. Ghasemi, M.; Vahedi Larijani, L.; Yazdani-Charati, J.; Kamali Hakim, E. Reduced Expression of Galectin-8 May Contribute in Carcinogenic Pathway of Head and Neck Squamous Cell Carcinoma. *Iran. J. Pathol.* **2021**, *16*, 195–204. [[CrossRef](#)] [[PubMed](#)]
18. Guo, J.P.; Li, X.G. Galectin-7 promotes the invasiveness of human oral squamous cell carcinoma cells via activation of ERK and JNK signaling. *Oncol. Lett.* **2017**, *13*, 1919–1924. [[CrossRef](#)]
19. Fik, Z.; Valach, J.; Chovanec, M.; Mazanek, J.; Kodet, R.; Kodet, O.; Tachezy, R.; Foltynova, E.; Andre, S.; Kaltner, H.; et al. Loss of adhesion/growth-regulatory galectin-9 from squamous cell epithelium in head and neck carcinomas. *J. Oral Pathol. Med. Off. Publ. Int. Assoc. Oral Pathol. Am. Acad. Oral Pathol.* **2013**, *42*, 166–173. [[CrossRef](#)]
20. Matsukawa, S.; Morita, K.; Negishi, A.; Harada, H.; Nakajima, Y.; Shimamoto, H.; Tomioka, H.; Tanaka, K.; Ono, M.; Yamada, T.; et al. Galectin-7 as a potential predictive marker of chemo- and/or radio-therapy resistance in oral squamous cell carcinoma. *Cancer Med.* **2014**, *3*, 349–361. [[CrossRef](#)]
21. Plzak, J.; Betka, J.; Smetana, K., Jr.; Chovanec, M.; Kaltner, H.; Andre, S.; Kodet, R.; Gabius, H.J. Galectin-3—An emerging prognostic indicator in advanced head and neck carcinoma. *Eur. J. Cancer* **2004**, *40*, 2324–2330. [[CrossRef](#)]
22. Tokmak, S.; Arik, D.; Pinarbasli, O.; Gurbuz, M.K.; Acikalin, M.F. Evaluation and Prognostic Significance of Galectin-3 Expression in Oral Squamous Cell Carcinoma. *Ear Nose Throat J.* **2021**, *100*, 578S–583S. [[CrossRef](#)]
23. Acikalin, M.F.; Etiz, D.; Gurbuz, M.K.; Ozudogru, E.; Canaz, F.; Colak, E. Prognostic significance of galectin-3 and cyclin D1 expression in undifferentiated nasopharyngeal carcinoma. *Med. Oncol.* **2012**, *29*, 742–749. [[CrossRef](#)] [[PubMed](#)]
24. Miranda, F.A.; Hassumi, M.K.; Guimaraes, M.C.; Simoes, R.T.; Silva, T.G.; Lira, R.C.; Rocha, A.M.; Mendes, C.T., Jr.; Donadi, E.A.; Soares, C.P.; et al. Galectin-3 overexpression in invasive laryngeal carcinoma, assessed by computer-assisted analysis. *J. Histochem. Cytochem. Off. J. Histochem. Soc.* **2009**, *57*, 665–673. [[CrossRef](#)] [[PubMed](#)]
25. Noda, Y.; Kondo, Y.; Sakai, M.; Sato, S.; Kishino, M. Galectin-1 is a useful marker for detecting neoplastic squamous cells in oral cytology smears. *Hum. Pathol.* **2016**, *52*, 101–109. [[CrossRef](#)] [[PubMed](#)]
26. Noda, Y.; Kishino, M.; Sato, S.; Hirose, K.; Sakai, M.; Fukuda, Y.; Murakami, S.; Toyosawa, S. Galectin-1 expression is associated with tumour immunity and prognosis in gingival squamous cell carcinoma. *J. Clin. Pathol.* **2017**, *70*, 126–133. [[CrossRef](#)] [[PubMed](#)]
27. Muniz, J.M.; Bibiano Borges, C.R.; Beghini, M.; de Araujo, M.S.; Miranda Alves, P.; de Lima, L.M.; Pereira, S.A.; Nogueira, R.D.; Napimoga, M.H.; Rodrigues, V., Jr.; et al. Galectin-9 as an important marker in the differential diagnosis between oral squamous cell carcinoma, oral leukoplakia and oral lichen planus. *Immunobiology* **2015**, *220*, 1006–1011. [[CrossRef](#)]
28. Chan, S.W.; Kallarakkal, T.G.; Abraham, M.T. Changed expression of E-cadherin and galectin-9 in oral squamous cell carcinomas but lack of potential as prognostic markers. *Asian Pac. J. Cancer Prev. APJCP* **2014**, *15*, 2145–2152. [[CrossRef](#)] [[PubMed](#)]
29. Zhong, L.P.; Wei, K.J.; Yang, X.; Pan, H.Y.; Ye, D.X.; Wang, L.Z.; Zhang, Z.Y. Overexpression of Galectin-1 is negatively correlated with pathologic differentiation grade in oral squamous cell carcinoma. *J. Cancer Res. Clin. Oncol.* **2010**, *136*, 1527–1535. [[CrossRef](#)] [[PubMed](#)]
30. Ferrazzo, K.L.; Alves, S.M., Jr.; Santos, E.; Martins, M.T.; de Sousa, S.M. Galectin-3 immunoprofile in adenoid cystic carcinoma and polymorphous low-grade adenocarcinoma of salivary glands. *Oral Oncol.* **2007**, *43*, 580–585. [[CrossRef](#)]
31. Saussez, S.; Decaestecker, C.; Lorfevre, F.; Cucu, D.R.; Mortuaire, G.; Chevalier, D.; Wacreniez, A.; Kaltner, H.; Andre, S.; Toubeau, G.; et al. High level of galectin-1 expression is a negative prognostic predictor of recurrence in laryngeal squamous cell carcinomas. *Int. J. Oncol.* **2007**, *30*, 1109–1117. [[CrossRef](#)] [[PubMed](#)]
32. Wang, L.P.; Chen, S.W.; Zhuang, S.M.; Li, H.; Song, M. Galectin-3 accelerates the progression of oral tongue squamous cell carcinoma via a Wnt/beta-catenin-dependent pathway. *Pathol. Oncol. Res. POR* **2013**, *19*, 461–474. [[CrossRef](#)]
33. Aimjongjun, S.; Reamtong, O.; Janvilisri, T. Lectin affinity chromatography and quantitative proteomic analysis reveal that galectin-3 is associated with metastasis in nasopharyngeal carcinoma. *Sci. Rep.* **2020**, *10*, 16462. [[CrossRef](#)] [[PubMed](#)]
34. Teymoortash, A.; Pientka, A.; Schrader, C.; Tiemann, M.; Werner, J.A. Expression of galectin-3 in adenoid cystic carcinoma of the head and neck and its relationship with distant metastasis. *J. Cancer Res. Clin. Oncol.* **2006**, *132*, 51–56. [[CrossRef](#)] [[PubMed](#)]
35. Chiang, W.F.; Liu, S.Y.; Fang, L.Y.; Lin, C.N.; Wu, M.H.; Chen, Y.C.; Chen, Y.L.; Jin, Y.T. Overexpression of galectin-1 at the tumor invasion front is associated with poor prognosis in early-stage oral squamous cell carcinoma. *Oral Oncol.* **2008**, *44*, 325–334. [[CrossRef](#)] [[PubMed](#)]
36. Hossaka, T.A.; Focchi, G.R.; Oshima, C.T.; Ribeiro, D.A. Detection of galectins during malignant transformation of oral cells. *Dent. Res. J.* **2013**, *10*, 428–433.
37. Coppock, J.D.; Mills, A.M.; Stelow, E.B. Galectin-3 Expression in High-Risk HPV-Positive and Negative Head & Neck Squamous Cell Carcinomas and Regional Lymph Node Metastases. *Head Neck Pathol.* **2021**, *15*, 163–168. [[CrossRef](#)]
38. Piantelli, M.; Iacobelli, S.; Almadori, G.; Iezzi, M.; Tinari, N.; Natoli, C.; Cadoni, G.; Lauriola, L.; Ranelletti, F.O. Lack of expression of galectin-3 is associated with a poor outcome in node-negative patients with laryngeal squamous-cell carcinoma. *J. Clin. Oncol. Off. J. Am. Soc. Clin. Oncol.* **2002**, *20*, 3850–3856. [[CrossRef](#)]
39. Valach, J.; Fik, Z.; Strnad, H.; Chovanec, M.; Plzak, J.; Cada, Z.; Szabo, P.; Sachova, J.; Hroudova, M.; Urbanova, M.; et al. Smooth muscle actin-expressing stromal fibroblasts in head and neck squamous cell carcinoma: Increased expression of galectin-1 and induction of poor prognosis factors. *Int. J. Cancer* **2012**, *131*, 2499–2508. [[CrossRef](#)]

40. Wehrhan, F.; Buttner-Herold, M.; Distel, L.; Ries, J.; Moebius, P.; Preidl, R.; Geppert, C.I.; Neukam, F.W.; Kesting, M.; Weber, M. Galectin 3 expression in regional lymph nodes and lymph node metastases of oral squamous cell carcinomas. *BMC Cancer* **2018**, *18*, 823. [[CrossRef](#)] [[PubMed](#)]
41. Weber, M.; Buttner-Herold, M.; Distel, L.; Ries, J.; Moebius, P.; Preidl, R.; Geppert, C.I.; Neukam, F.W.; Wehrhan, F. Galectin 3 expression in primary oral squamous cell carcinomas. *BMC Cancer* **2017**, *17*, 906. [[CrossRef](#)]
42. Saussez, S.; Cucu, D.R.; Decaestecker, C.; Chevalier, D.; Kaltner, H.; Andre, S.; Wacreniez, A.; Toubeau, G.; Camby, I.; Gabius, H.J.; et al. Galectin 7 (p53-induced gene 1): A new prognostic predictor of recurrence and survival in stage IV hypopharyngeal cancer. *Ann. Surg. Oncol.* **2006**, *13*, 999–1009. [[CrossRef](#)] [[PubMed](#)]
43. Chandrashekar, D.S.; Bashel, B.; Balasubramanya, S.A.H.; Creighton, C.J.; Ponce-Rodriguez, I.; Chakravarthi, B.; Varambally, S. UALCAN: A Portal for Facilitating Tumor Subgroup Gene Expression and Survival Analyses. *Neoplasia* **2017**, *19*, 649–658. [[CrossRef](#)] [[PubMed](#)]
44. Ouyang, J.; Qin, G.; Liu, Z.; Jian, X.; Shi, T.; Xie, L. ToPP: Tumor online prognostic analysis platform for prognostic feature selection and clinical patient subgroup selection. *iScience* **2022**, *25*, 104190. [[CrossRef](#)] [[PubMed](#)]
45. Coker, E.A.; Mitsopoulos, C.; Tym, J.E.; Komianou, A.; Kannas, C.; Di Micco, P.; Villasclaras Fernandez, E.; Ozer, B.; Antolin, A.A.; Workman, P.; et al. canSAR: Update to the cancer translational research and drug discovery knowledgebase. *Nucleic Acids Res.* **2019**, *47*, D917–D922. [[CrossRef](#)]
46. Han, Y.; Wang, Y.; Dong, X.; Sun, D.; Liu, Z.; Yue, J.; Wang, H.; Li, T.; Wang, C. TISCH2: Expanded datasets and new tools for single-cell transcriptome analyses of the tumor microenvironment. *Nucleic Acids Res.* **2023**, *51*, D1425–D1431. [[CrossRef](#)] [[PubMed](#)]
47. Warde-Farley, D.; Donaldson, S.L.; Comes, O.; Zuberi, K.; Badrawi, R.; Chao, P.; Franz, M.; Grouios, C.; Kazi, F.; Lopes, C.T.; et al. The GeneMANIA prediction server: Biological network integration for gene prioritization and predicting gene function. *Nucleic Acids Res.* **2010**, *38*, W214–W220. [[CrossRef](#)]
48. Szklarczyk, D.; Gable, A.L.; Nastou, K.C.; Lyon, D.; Kirsch, R.; Pyysalo, S.; Doncheva, N.T.; Legeay, M.; Fang, T.; Bork, P.; et al. The STRING database in 2021: Customizable protein-protein networks, and functional characterization of user-uploaded gene/measurement sets. *Nucleic Acids Res.* **2021**, *49*, D605–D612. [[CrossRef](#)]
49. Gillespie, M.; Jassal, B.; Stephan, R.; Milacic, M.; Rothfels, K.; Senff-Ribeiro, A.; Griss, J.; Sevilla, C.; Matthews, L.; Gong, C.; et al. The reactome pathway knowledgebase 2022. *Nucleic Acids Res.* **2022**, *50*, D687–D692. [[CrossRef](#)]
50. Liu, C.J.; Hu, F.F.; Xia, M.X.; Han, L.; Zhang, Q.; Guo, A.Y. GSCALite: A web server for gene set cancer analysis. *Bioinformatics* **2018**, *34*, 3771–3772. [[CrossRef](#)]
51. Sturgill, E.R.; Rolig, A.S.; Linch, S.N.; Mick, C.; Kasiewicz, M.J.; Sun, Z.; Traber, P.G.; Shlevin, H.; Redmond, W.L. Galectin-3 inhibition with belapectin combined with anti-OX40 therapy reprograms the tumor microenvironment to favor anti-tumor immunity. *Oncoimmunology* **2021**, *10*, 1892265. [[CrossRef](#)]
52. Balakrishnan, B.; Subramanian, S.; Mallia, M.B.; Repaka, K.; Kaur, S.; Chandan, R.; Bhardwaj, P.; Dash, A.; Banerjee, R. Multifunctional Core-Shell Glyconanoparticles for Galectin-3-Targeted, Trigger-Responsive Combination Chemotherapy. *Biomacromolecules* **2020**, *21*, 2645–2660. [[CrossRef](#)]
53. Dos Santos, S.N.; Sheldon, H.; Pereira, J.X.; Paluch, C.; Bridges, E.M.; El-Cheikh, M.C.; Harris, A.L.; Bernardes, E.S. Galectin-3 acts as an angiogenic switch to induce tumor angiogenesis via Jagged-1/Notch activation. *Oncotarget* **2017**, *8*, 49484–49501. [[CrossRef](#)] [[PubMed](#)]
54. Toti, A.; Santi, A.; Pardella, E.; Nesi, I.; Tomasini, R.; Mello, T.; Paoli, P.; Caselli, A.; Cirri, P. Activated fibroblasts enhance cancer cell migration by microvesicles-mediated transfer of Galectin-1. *J. Cell Commun. Signal.* **2021**, *15*, 405–419. [[CrossRef](#)]
55. Nakajima, K.; Kho, D.H.; Yanagawa, T.; Harazono, Y.; Hogan, V.; Chen, W.; Ali-Fehmi, R.; Mehra, R.; Raz, A. Galectin-3 Cleavage Alters Bone Remodeling: Different Outcomes in Breast and Prostate Cancer Skeletal Metastasis. *Cancer Res.* **2016**, *76*, 1391–1402. [[CrossRef](#)] [[PubMed](#)]
56. Qi, Y.; Chang, Y.; Wang, Z.; Chen, L.; Kong, Y.; Zhang, P.; Liu, Z.; Zhou, Q.; Chen, Y.; Wang, J.; et al. Tumor-associated macrophages expressing galectin-9 identify immunoevasive subtype muscle-invasive bladder cancer with poor prognosis but favorable adjuvant chemotherapeutic response. *Cancer Immunol. Immunother. CII* **2019**, *68*, 2067–2080. [[CrossRef](#)] [[PubMed](#)]
57. Pereira, J.X.; Dos Santos, S.N.; Pereira, T.C.; Cabanel, M.; Chammas, R.; de Oliveira, F.L.; Bernardes, E.S.; El-Cheikh, M.C. Galectin-3 Regulates the Expression of Tumor Glycosaminoglycans and Increases the Metastatic Potential of Breast Cancer. *J. Oncol.* **2019**, *2019*, 9827147. [[CrossRef](#)] [[PubMed](#)]
58. Muller, J.; Duray, E.; Lejeune, M.; Dubois, S.; Plougonven, E.; Leonard, A.; Storti, P.; Giuliani, N.; Cohen-Solal, M.; Hempel, U.; et al. Loss of Stromal Galectin-1 Enhances Multiple Myeloma Development: Emphasis on a Role in Osteoclasts. *Cancers* **2019**, *11*, 261. [[CrossRef](#)]
59. Giesbrecht, K.; Former, S.; Sahr, A.; Heeg, K.; Hildebrand, D. Streptococcal Pyrogenic Exotoxin A-Stimulated Monocytes Mediate Regulatory T-Cell Accumulation through PD-L1 and Kynurenine. *Int. J. Mol. Sci.* **2019**, *20*, 3933. [[CrossRef](#)]
60. Chen, T.C.; Chen, C.H.; Wang, C.P.; Lin, P.H.; Yang, T.L.; Lou, P.J.; Ko, J.Y.; Wu, C.T.; Chang, Y.L. The immunologic advantage of recurrent nasopharyngeal carcinoma from the viewpoint of Galectin-9/Tim-3-related changes in the tumour microenvironment. *Sci. Rep.* **2017**, *7*, 10349. [[CrossRef](#)]

61. You, Y.; Tan, J.X.; Dai, H.S.; Chen, H.W.; Xu, X.J.; Yang, A.G.; Zhang, Y.J.; Bai, L.H.; Bie, P. MiRNA-22 inhibits oncogene galectin-1 in hepatocellular carcinoma. *Oncotarget* **2016**, *7*, 57099–57116. [[CrossRef](#)]
62. Nambiar, D.K.; Aguilera, T.; Cao, H.; Kwok, S.; Kong, C.; Bloomstein, J.; Wang, Z.; Rangan, V.S.; Jiang, D.; von Eyben, R.; et al. Galectin-1-driven T cell exclusion in the tumor endothelium promotes immunotherapy resistance. *J. Clin. Investig.* **2019**, *129*, 5553–5567. [[CrossRef](#)]
63. Kouo, T.; Huang, L.; Pucsek, A.B.; Cao, M.; Solt, S.; Armstrong, T.; Jaffee, E. Galectin-3 Shapes Antitumor Immune Responses by Suppressing CD8+ T Cells via LAG-3 and Inhibiting Expansion of Plasmacytoid Dendritic Cells. *Cancer Immunol. Res.* **2015**, *3*, 412–423. [[CrossRef](#)]
64. Daley, D.; Mani, V.R.; Mohan, N.; Akkad, N.; Ochi, A.; Heindel, D.W.; Lee, K.B.; Zambirinis, C.P.; Pandian, G.S.B.; Savadkar, S.; et al. Dectin 1 activation on macrophages by galectin 9 promotes pancreatic carcinoma and peritumoral immune tolerance. *Nat. Med.* **2017**, *23*, 556–567. [[CrossRef](#)]
65. Croci, D.O.; Cerliani, J.P.; Dalotto-Moreno, T.; Mendez-Huergo, S.P.; Mascanfroni, I.D.; Dergan-Dylon, S.; Toscano, M.A.; Caramelo, J.J.; Garcia-Vallejo, J.J.; Ouyang, J.; et al. Glycosylation-dependent lectin-receptor interactions preserve angiogenesis in anti-VEGF refractory tumors. *Cell* **2014**, *156*, 744–758. [[CrossRef](#)]
66. Severson, J.J.; Serracino, H.S.; Mateescu, V.; Raeburn, C.D.; McIntyre, R.C., Jr.; Sams, S.B.; Haugen, B.R.; French, J.D. PD-1+Tim-3+ CD8+ T Lymphocytes Display Varied Degrees of Functional Exhaustion in Patients with Regionally Metastatic Differentiated Thyroid Cancer. *Cancer Immunol. Res.* **2015**, *3*, 620–630. [[CrossRef](#)]
67. Vaitaitis, G.M.; Wagner, D.H., Jr. Galectin-9 controls CD40 signaling through a Tim-3 independent mechanism and redirects the cytokine profile of pathogenic T cells in autoimmunity. *PLoS ONE* **2012**, *7*, e38708. [[CrossRef](#)] [[PubMed](#)]
68. Nam, K.; Son, S.H.; Oh, S.; Jeon, D.; Kim, H.; Noh, D.Y.; Kim, S.; Shin, I. Binding of galectin-1 to integrin beta1 potentiates drug resistance by promoting survivin expression in breast cancer cells. *Oncotarget* **2017**, *8*, 35804–35823. [[CrossRef](#)]
69. AbuSamra, D.B.; Mauris, J.; Argueso, P. Galectin-3 initiates epithelial-stromal paracrine signaling to shape the proteolytic microenvironment during corneal repair. *Sci. Signal.* **2019**, *12*, eaaw7095. [[CrossRef](#)] [[PubMed](#)]
70. Madireddi, S.; Eun, S.Y.; Lee, S.W.; Nemcovicova, I.; Mehta, A.K.; Zajonc, D.M.; Nishi, N.; Niki, T.; Hirashima, M.; Croft, M. Galectin-9 controls the therapeutic activity of 4-1BB-targeting antibodies. *J. Exp. Med.* **2014**, *211*, 1433–1448. [[CrossRef](#)] [[PubMed](#)]
71. Sandberg, T.P.; Oosting, J.; van Pelt, G.W.; Mesker, W.E.; Tollenaar, R.; Morreau, H. Molecular profiling of colorectal tumors stratified by the histological tumor-stroma ratio—Increased expression of galectin-1 in tumors with high stromal content. *Oncotarget* **2018**, *9*, 31502–31515. [[CrossRef](#)]
72. Vuong, L.; Kouverianou, E.; Rooney, C.M.; McHugh, B.J.; Howie, S.E.M.; Gregory, C.D.; Forbes, S.J.; Henderson, N.C.; Zetterberg, F.R.; Nilsson, U.J.; et al. An Orally Active Galectin-3 Antagonist Inhibits Lung Adenocarcinoma Growth and Augments Response to PD-L1 Blockade. *Cancer Res.* **2019**, *79*, 1480–1492. [[CrossRef](#)]
73. Zhong, S.; Jeong, J.H.; Chen, Z.; Chen, Z.; Luo, J.L. Targeting Tumor Microenvironment by Small-Molecule Inhibitors. *Transl. Oncol.* **2020**, *13*, 57–69. [[CrossRef](#)]
74. Corapi, E.; Carrizo, G.; Compagno, D.; Laderach, D. Endogenous Galectin-1 in T Lymphocytes Regulates Anti-prostate Cancer Immunity. *Front. Immunol.* **2018**, *9*, 2190. [[CrossRef](#)]
75. Jin, M.Z.; Jin, W.L. The updated landscape of tumor microenvironment and drug repurposing. *Signal Transduct. Target. Ther.* **2020**, *5*, 166. [[CrossRef](#)]
76. Pereira, J.X.; Azeredo, M.C.; Martins, F.S.; Chammas, R.; Oliveira, F.L.; Santos, S.N.; Bernardes, E.S.; El-Cheikh, M.C. The deficiency of galectin-3 in stromal cells leads to enhanced tumor growth and bone marrow metastasis. *BMC Cancer* **2016**, *16*, 636. [[CrossRef](#)] [[PubMed](#)]
77. Enninga, E.A.L.; Harrington, S.M.; Creedon, D.J.; Ruano, R.; Markovic, S.N.; Dong, H.; Dronca, R.S. Immune checkpoint molecules soluble program death ligand 1 and galectin-9 are increased in pregnancy. *Am. J. Reprod. Immunol.* **2018**, *79*, e12795. [[CrossRef](#)] [[PubMed](#)]
78. Luo, Z.; Ji, Y.; Tian, D.; Zhang, Y.; Chang, S.; Yang, C.; Zhou, H.; Chen, Z.K. Galectin-7 promotes proliferation and Th1/2 cells polarization toward Th1 in activated CD4+ T cells by inhibiting The TGFbeta/Smad3 pathway. *Mol. Immunol.* **2018**, *101*, 80–85. [[CrossRef](#)]
79. Li, Y.; Gong, S.; Pan, W.; Chen, Y.; Liu, B.; Li, N.; Tang, B. A tumor acidity activatable and Ca(2+)-assisted immuno-nanoagent enhances breast cancer therapy and suppresses cancer recurrence. *Chem. Sci.* **2020**, *11*, 7429–7437. [[CrossRef](#)]
80. Li, X.; Chen, Y.; Liu, X.; Zhang, J.; He, X.; Teng, G.; Yu, D. Tim3/Gal9 interactions between T cells and monocytes result in an immunosuppressive feedback loop that inhibits Th1 responses in osteosarcoma patients. *Int. Immunopharmacol.* **2017**, *44*, 153–159. [[CrossRef](#)] [[PubMed](#)]
81. Baker, G.J.; Chockley, P.; Zamlar, D.; Castro, M.G.; Lowenstein, P.R. Natural killer cells require monocytic Gr-1(+)/CD11b(+) myeloid cells to eradicate orthotopically engrafted glioma cells. *Oncoimmunology* **2016**, *5*, e1163461. [[CrossRef](#)] [[PubMed](#)]
82. Colomb, F.; Wang, W.; Simpson, D.; Zafar, M.; Beynon, R.; Rhodes, J.M.; Yu, L.G. Galectin-3 interacts with the cell-surface glycoprotein CD146 (MCAM, MUC18) and induces secretion of metastasis-promoting cytokines from vascular endothelial cells. *J. Biol. Chem.* **2017**, *292*, 8381–8389. [[CrossRef](#)] [[PubMed](#)]



83. Seki, M.; Oomizu, S.; Sakata, K.M.; Sakata, A.; Arikawa, T.; Watanabe, K.; Ito, K.; Takeshita, K.; Niki, T.; Saita, N.; et al. Galectin-9 suppresses the generation of Th17, promotes the induction of regulatory T cells, and regulates experimental autoimmune arthritis. *Clin. Immunol.* **2008**, *127*, 78–88. [[CrossRef](#)] [[PubMed](#)]
84. Demotte, N.; Wieers, G.; Van Der Smissen, P.; Moser, M.; Schmidt, C.; Thielemans, K.; Squifflet, J.L.; Weynand, B.; Carrasco, J.; Lurquin, C.; et al. A galectin-3 ligand corrects the impaired function of human CD4 and CD8 tumor-infiltrating lymphocytes and favors tumor rejection in mice. *Cancer Res.* **2010**, *70*, 7476–7488. [[CrossRef](#)] [[PubMed](#)]
85. Suzuki, Y.; Sutoh, M.; Hatakeyama, S.; Mori, K.; Yamamoto, H.; Koie, T.; Saitoh, H.; Yamaya, K.; Funyu, T.; Habuchi, T.; et al. MUC1 carrying core 2 O-glycans functions as a molecular shield against NK cell attack, promoting bladder tumor metastasis. *Int. J. Oncol.* **2012**, *40*, 1831–1838. [[CrossRef](#)]
86. Wang, W.; Guo, H.; Geng, J.; Zheng, X.; Wei, H.; Sun, R.; Tian, Z. Tumor-released Galectin-3, a soluble inhibitory ligand of human NKp30, plays an important role in tumor escape from NK cell attack. *J. Biol. Chem.* **2014**, *289*, 33311–33319. [[CrossRef](#)]
87. Cedeno-Laurent, F.; Opperman, M.J.; Barthel, S.R.; Hays, D.; Schatton, T.; Zhan, Q.; He, X.; Matta, K.L.; Supko, J.G.; Frank, M.H.; et al. Metabolic inhibition of galectin-1-binding carbohydrates accentuates antitumor immunity. *J. Investig. Dermatol.* **2012**, *132*, 410–420. [[CrossRef](#)]
88. Peng, W.; Wang, H.Y.; Miyahara, Y.; Peng, G.; Wang, R.F. Tumor-associated galectin-3 modulates the function of tumor-reactive T cells. *Cancer Res.* **2008**, *68*, 7228–7236. [[CrossRef](#)]
89. Rabinovich, G.A.; Ramhorst, R.E.; Rubinstein, N.; Corigliano, A.; Daroqui, M.C.; Kier-Joffe, E.B.; Fainboim, L. Induction of allogenic T-cell hyporesponsiveness by galectin-1-mediated apoptotic and non-apoptotic mechanisms. *Cell Death Differ.* **2002**, *9*, 661–670. [[CrossRef](#)]
90. Kovacs-Solyom, F.; Blasko, A.; Fajka-Boja, R.; Katona, R.L.; Vegh, L.; Novak, J.; Szebeni, G.J.; Krenacs, L.; Uher, F.; Tubak, V.; et al. Mechanism of tumor cell-induced T-cell apoptosis mediated by galectin-1. *Immunol. Lett.* **2010**, *127*, 108–118. [[CrossRef](#)]
91. Camby, I.; Le Mercier, M.; Lefranc, F.; Kiss, R. Galectin-1: A small protein with major functions. *Glycobiology* **2006**, *16*, 137R–157R. [[CrossRef](#)] [[PubMed](#)]
92. Koguchi, K.; Anderson, D.E.; Yang, L.; O'Connor, K.C.; Kuchroo, V.K.; Hafler, D.A. Dysregulated T cell expression of TIM3 in multiple sclerosis. *J. Exp. Med.* **2006**, *203*, 1413–1418. [[CrossRef](#)]
93. Dai, S.Y.; Nakagawa, R.; Itoh, A.; Murakami, H.; Kashio, Y.; Abe, H.; Katoh, S.; Kontani, K.; Kihara, M.; Zhang, S.L.; et al. Galectin-9 induces maturation of human monocyte-derived dendritic cells. *J. Immunol.* **2005**, *175*, 2974–2981. [[CrossRef](#)] [[PubMed](#)]
94. Paron, I.; Scaloni, A.; Pines, A.; Bachi, A.; Liu, F.T.; Puppini, C.; Pandolfi, M.; Ledda, L.; Di Loreto, C.; Damante, G.; et al. Nuclear localization of Galectin-3 in transformed thyroid cells: A role in transcriptional regulation. *Biochem. Biophys. Res. Commun.* **2003**, *302*, 545–553. [[CrossRef](#)]
95. Elad-Sfadia, G.; Haklai, R.; Balan, E.; Kloog, Y. Galectin-3 augments K-Ras activation and triggers a Ras signal that attenuates ERK but not phosphoinositide 3-kinase activity. *J. Biol. Chem.* **2004**, *279*, 34922–34930. [[CrossRef](#)]
96. Stillman, B.N.; Hsu, D.K.; Pang, M.; Brewer, C.F.; Johnson, P.; Liu, F.T.; Baum, L.G. Galectin-3 and galectin-1 bind distinct cell surface glycoprotein receptors to induce T cell death. *J. Immunol.* **2006**, *176*, 778–789. [[CrossRef](#)] [[PubMed](#)]
97. Chung, C.D.; Patel, V.P.; Moran, M.; Lewis, L.A.; Miceli, M.C. Galectin-1 induces partial TCR zeta-chain phosphorylation and antagonizes processive TCR signal transduction. *J. Immunol.* **2000**, *165*, 3722–3729. [[CrossRef](#)]
98. Sanchez-Ruderisch, H.; Detjen, K.M.; Welzel, M.; Andre, S.; Fischer, C.; Gabius, H.J.; Rosewicz, S. Galectin-1 sensitizes carcinoma cells to anoikis via the fibronectin receptor alpha5beta1-integrin. *Cell Death Differ.* **2011**, *18*, 806–816. [[CrossRef](#)]
99. Rossi, B.; Espeli, M.; Schiff, C.; Gauthier, L. Clustering of pre-B cell integrins induces galectin-1-dependent pre-B cell receptor relocalization and activation. *J. Immunol.* **2006**, *177*, 796–803. [[CrossRef](#)]
100. Walzel, H.; Fahmi, A.A.; Eldesouky, M.A.; Abou-Eladab, E.F.; Waitz, G.; Brock, J.; Tiedge, M. Effects of N-glycan processing inhibitors on signaling events and induction of apoptosis in galectin-1-stimulated Jurkat T lymphocytes. *Glycobiology* **2006**, *16*, 1262–1271. [[CrossRef](#)]
101. Hsieh, S.H.; Ying, N.W.; Wu, M.H.; Chiang, W.F.; Hsu, C.L.; Wong, T.Y.; Jin, Y.T.; Hong, T.M.; Chen, Y.L. Galectin-1, a novel ligand of neuropilin-1, activates VEGFR-2 signaling and modulates the migration of vascular endothelial cells. *Oncogene* **2008**, *27*, 3746–3753. [[CrossRef](#)] [[PubMed](#)]
102. Anderson, A.C.; Anderson, D.E.; Bregoli, L.; Hastings, W.D.; Kassam, N.; Lei, C.; Chandwaskar, R.; Karman, J.; Su, E.W.; Hirashima, M.; et al. Promotion of tissue inflammation by the immune receptor Tim-3 expressed on innate immune cells. *Science* **2007**, *318*, 1141–1143. [[CrossRef](#)] [[PubMed](#)]
103. Saussez, S.; Decaestecker, C.; Mahillon, V.; Cludts, S.; Capouillez, A.; Chevalier, D.; Vet, H.K.; Andre, S.; Toubeau, G.; Leroy, X.; et al. Galectin-3 upregulation during tumor progression in head and neck cancer. *Laryngoscope* **2008**, *118*, 1583–1590. [[CrossRef](#)] [[PubMed](#)]
104. Li, M.; Chen, Y.B.; Liu, F.; Qu, J.Q.; Ren, L.C.; Chai, J.; Tang, C.E. Galectin-3 facilitates the proliferation and migration of nasopharyngeal carcinoma cells via activation of the ERK1/2 and Akt signaling pathways, and is positively correlated with the inflammatory state of nasopharyngeal carcinoma. *Mol. Med. Rep.* **2021**, *23*, 370. [[CrossRef](#)] [[PubMed](#)]
105. Andisheh-Tadbir, A.; Mardani, M.; Malekzadeh, M.; Amirbeigi Tafti, T.; Khademi, B. Galectin-3 Serum Levels Could Help Clinicians Screen for Salivary Gland Tumor Patients. *Asian Pac. J. Cancer Prev. APJCP* **2018**, *19*, 689–692. [[CrossRef](#)]

106. Curti, B.D.; Koguchi, Y.; Leidner, R.S.; Rolig, A.S.; Sturgill, E.R.; Sun, Z.; Wu, Y.; Rajamanickam, V.; Bernard, B.; Hilgart-Martiszus, I.; et al. Enhancing clinical and immunological effects of anti-PD-1 with belapectin, a galectin-3 inhibitor. *J. Immunother. Cancer* **2021**, *9*, e002371. [[CrossRef](#)]
107. Zhang, X.; Ding, H.; Lu, Z.; Ding, L.; Song, Y.; Jing, Y.; Hu, Q.; Dong, Y.; Ni, Y. Increased LGALS3BP promotes proliferation and migration of oral squamous cell carcinoma via PI3K/AKT pathway. *Cell. Signal.* **2019**, *63*, 109359. [[CrossRef](#)]

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