

## Rapid Nickel-based Isolation of Extracellular Vesicles from Different Biological Fluids

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**[Abstract]** Extracellular vesicles (EVs) are membranous structures that cells massively release in extracellular fluids. EVs are cargo of cellular components such as lipids, proteins, and nucleic acids that can work as a formidable source in liquid biopsy studies searching for disease biomarkers. We recently demonstrated that nickel-based isolation (NBI) is a valuable method for fast, efficient, and easy recovery of heterogeneous EVs from biological fluids. NBI exploits nickel cations to capture negatively charged vesicles. Then, a mix of balanced chelating agents elutes EVs while preserving their integrity and stability in solution. Here, we describe steps and quality controls to functionalize a matrix of agarose beads, obtain an efficient elution of EVs, and extract nucleic acids carried by them. We demonstrate the versatility of NBI method in isolating EVs from media of primary mouse astrocytes, from human blood, urine, and saliva processed in parallel, as well as outer membrane vesicles (OMVs) from cultured Gram-negative bacteria.

**Keywords:** Extracellular vesicles, OMVs, Nickel, Biomarkers, Liquid biopsies

**[Background]** Extracellular vesicles (EVs) are heterogeneous membranous structures that cells release in the extracellular fluids. Besides the generation of apoptotic bodies, we now recognize at least two mechanisms that generate EVs, comprising the formation of multivesicular bodies (MVBs) in mature endosomes that fuse with the plasma membrane and discharge exosomes, and the direct release of membrane-budding vesicles known as microvesicles or ectosomes (van Niel *et al.*, 2018). EVs are increasingly referred to as intercellular communicators which are able to influence the metabolism of recipient cells through the horizontal exchange of different components such as lipids, proteins, and nucleic acids (Raposo and Stoorvogel, 2013; Hirshman *et al.*, 2016; Jeppesen *et al.*, 2019). Besides the research efforts aiming to convert EVs into novel carriers of therapeutics, the incredible density and heterogeneity of EVs present in biological fluids hold promise for the interception of pathological conditions such as cancer or neurodegenerative diseases (Schiera *et al.*, 2015; Bandu *et al.*, 2019; Latifkar *et al.*, 2019). Especially in cancer, this perspective is nowadays expanding the field of liquid biopsy tests, where the non-invasive detection of biomarkers through a systematic screening of circulating tumor cells, cell-free DNA, and EVs could advance early detection and monitoring, with novel opportunities for patients' stratification and prediction of disease progression.

EVs have been detected in blood, urine, cerebrospinal fluid, saliva, tears, breast milk, as well as from cell culture media (Lässer *et al.*, 2011; Rolfo *et al.*, 2014; Salih *et al.*, 2014; Pieragostino *et al.*, 2019; Palviainen *et al.*, 2019). Multiple strategies have been presented to isolate them, with protocols based on differential ultracentrifugation, density gradient, or polymer precipitation, allowing for a massive enrichment of dimensionally heterogeneous vesicles depending on their size, buoyant density, or lipid structure (Chen *et al.*, 2019). Alternatively, immunocapture approaches have been exploited to enrich for EV sub-populations, virtually deriving from the same tissue of origin (Kowal *et al.*, 2016).

In research laboratories, the most widely used method for EV isolation is the differential ultracentrifugation, in which large EVs of about 200-1,000 nm sediment at low speed (10,000 x *g*), while small EVs of about 50-200 nm pellet at high speed (100,000 x *g*), as described by Théry *et al.*, 2006. This size-based classification cannot be exploited to infer the intracellular pathways of their biogenesis, as ectosomes generated by the outward budding of the plasma membrane can overlap in size with exosomes derived from the MVBs (multivesicular bodies). Nevertheless, specific tetraspanins (CD9, CD81, CD63) or lumen proteins (Alix, synthenin) are found to be enriched in the small EVs, including exosomes (Jeppesen *et al.*, 2019; Latifkar *et al.*, 2019). Ultracentrifugation has some drawbacks related to reproducibility of results when comparing different instruments/rotors, or to the physical force applied to balance yield *versus* integrity (Livshits *et al.*, 2015). Typically, this technique is combined with size-exclusion chromatography or floatation in Optiprep gradient to separate EVs from co-sedimenting protein aggregates (Théry *et al.*, 2018). However, these approaches are inapplicable in view of a routine clinical practice, where the recovery of EVs needs to be fast and the biological sample not exposed to chemicals that could contribute experimental variability but interfere in downstream analysis. For these reasons, other microfluidic- or electromagnetic-based technologies that exploit other biophysical features of EVs have been implemented (Hou *et al.*, 2019). The strategy of capturing EVs through electrostatic interactions (negative zeta potential values) has recently shown the advantages of rapidity and versatility in terms of device implementation, combination with other protocols like size-exclusion chromatography, and recovery of heterogeneous suspended vesicles (Petersen *et al.*, 2018; Heath *et al.*, 2018).

Here, we describe the procedure of nickel-based isolation (NBI) we recently developed as a low-cost and convenient method for isolating EVs (Notarangelo *et al.*, 2019). This method is based on nickel cations to capture EVs and on the use of chelating agents that synergistically elute vesicles in a physiological pH (or pH-controlled) solution. We demonstrated that NBI efficiently captures EVs and isolates them as non-aggregated vesicles in solution while preserving their integrity and minimizing the co-elution of exogenous protein contaminants (Notarangelo *et al.*, 2019). Importantly, NBI method is fast, scalable, and easy to apply, offering a valuable option for liquid biopsy investigations and EV preparations that could benefit from increased stability and a non-aggregated distribution in solution. We describe the procedure starting from the functionalization of a matrix of agarose beads with nickel ions, the steps for efficient elution, and the protocol we use to extract nucleic acids from eluted EVs. We show here the versatility of NBI method in isolating EVs from different biological fluids, including media

from primary mouse astrocytes, human plasma, urine, and saliva processed in parallel, as well as outer membrane vesicles (OMVs) from cultured Gram-negative bacteria.

## **Materials and Reagents**

1. Low-binding protein tubes: LoBind Tubes, Protein LoBind, 2.0 ml, PCR clean (Eppendorf, catalog number: 0030108132)
2. Ni Sepharose® High Performance, pack of 25 ml (GE Healthcare, catalog number: 17-5268-01), store at 4 °C
3. PVDF Syringe Filters (Euroclone, Primo® Syringe Filters, 0.22 µm pour size, catalog number: EPSPV2230)
4. 50 ml tube
5. 15 ml Falcon tube
6. 1.5 ml centrifuge tube
7. Water prepared by ultrafiltration, reverse osmosis, deionization and sterile filtration (e.g., Lonza®, catalog number: 17-724Q)
8. Phosphate-buffered saline (PBS 1x, pH 7.4) (Thermo Fisher®, catalog number: 10010023)
9. Nickel sulphate powder, NiSO<sub>4</sub> (Sigma®, catalog number: 656895)
10. NaCl powder (Sigma®, catalog number: 450006)
11. EDTA [0.5] M (Thermo Fisher®, UltraPure pH 8.0, catalog number: 15575020)
12. Citric acid (Sigma®, catalog number: 251275)
13. TRIzol™ Reagent (Thermo Fisher®, catalog number: 15596026)
14. Agilent RNA 6000 Pico Kit (Agilent, catalog number: 5067-1513)
15. Chloroform (Sigma®, catalog number: C7559)
16. Isopropanol Molecular Biology Reagent (Sigma®, catalog number: I9516)
17. Ethanol (Sigma®, catalog number: 51976)
18. UltraPure™ DNase/RNase-Free Distilled Water (Thermo Fisher®, catalog number: 10977035)
19. Glycogen, RNA grade (Thermo Fisher®, catalog number: R0551)
20. Buffer S (Stripping buffer) (see Recipe 1)
21. Buffer N (see Recipe 2)
22. Solution A 5x (see Recipe 3)
23. Solution B 5x (see Recipe 4)

## **Equipment**

1. Laminar flow hood
2. Burkholder chamber (VWR, MARI0640230)

3. Eppendorf® Centrifuge 5702, non-refrigerated, with Swing-bucket Rotor (A-4-38) included adapters for 15/50 ml conical tubes (Eppendorf, catalog number: 5702 000.32) (used in paragraphs A-B-C of the procedure)
4. Eppendorf® Thermomixer Comfort (Eppendorf, catalog number: 5355.000.011)
5. VWR® Tube Rotator (VWR, catalog number: 10136-084) with interchangeable rotisserie assembly for: 10x 10/15 and 16x 5/7 ml tubes (VWR, catalog number: 444-0504), 6x 50 ml tubes (VWR, catalog number: 444-0505), 36x 1.5/2.0 ml tubes (VWR, catalog number: 444-0503)
6. Eppendorf® Centrifuge 5415 R (Eppendorf, model: Centrifuge 5415 R, catalog number: 22 62 140-8), refrigerated, with Rotor (Eppendorf, catalog number: F-45-24-11) (used in Procedure D)
7. 2100 Bioanalyzer Instrument (Agilent Technologies, catalog number: G2939BA)
8. -80 °C freezer

## **Procedure**

### **A. Stripping and functionalization of agarose beads**

*Note: Steps A1-A3 are mostly known as part of the standard “stripping/re-charge” protocol well described by manufacturer’s selling nickel-functionalized matrices. We still include these details as these are part of the procedure we follow.*

1. Transfer the beads to a 50 ml tube, centrifuge at 600-800 x g for 2 min at room temperature (RT), and discard the supernatant without disturbing the bead pellet.
2. Add 1:1 bead volume of pure water and mix by gentle rotation and inversion of the tube until beads are homogeneously resuspended. Then recover beads by centrifugation at 600-800 x g for 2 min and discard the supernatant.
3. Add 1:1 bead volume of Buffer S (Stripping buffer, Recipe 1) and resuspend beads by inverting the tube several times. Incubate for 5 min at RT and recover beads again by centrifugation (at 600-800 x g for 2 min). Discard the supernatant.

*Note: This step removes nickel ions from the beads, which turn white indicating the nickel stripping from the matrix. Remember to pay attention to appropriate waste disposals for solutions containing nickel, especially after Step A4.*

4. Add 1:1 bead volume of Buffer N (Recipe 2). Gently rotate the tube to obtain a homogeneous solution of beads. Incubate at room temperature for 5 min, then recover beads by centrifugation at 600-800 x g for 2 min.

*Note: The concentration of Nickel sulphate (0.4 M) used to functionalize beads confers a positive charge to the matrix able to capture a number of vesicles at least equivalent to that obtained by ultracentrifugation of the same biological sample (Notarangelo et al., 2019). At this step, beads appear in light blue color.*

5. Wash beads with 1:1 bead volume of pure water. Recover them again after centrifugation (at 600-800 x g for 2 min) and discard the supernatant.

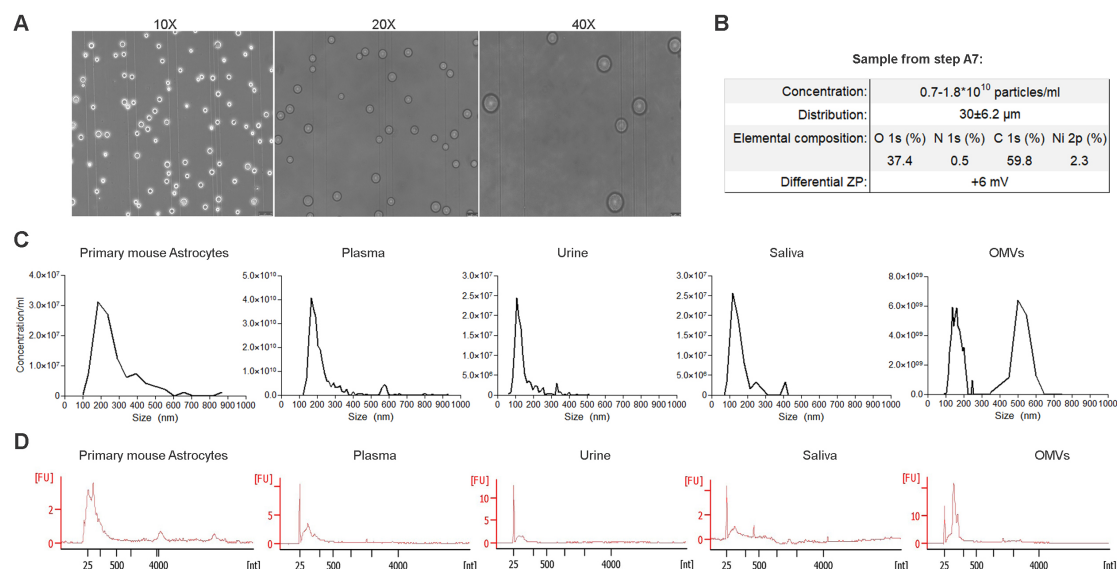
*Note: We usually perform Steps A5-A7 under the laminar flow hood to reduce the chances of biological contamination.*

- Wash beads with 1:1 bead volume of PBS 1x. Gently rotate the tube to homogenize beads in solution. Always recover them by centrifugation at 600-800 x g for 2 min. Repeat this step twice and carefully remove the supernatant using a serological pipet with a slow, constant aspiration of the volume to avoid disturbing the bead pellet.

*Note: The removal of unbound nickel residues is crucial to avoid contamination of the biological sample.*

- Resuspend beads with PBS 1x to obtain a 20 mg/mx final concentration, corresponding to a range of  $0.7 \times 10^{10}$ - $1.8 \times 10^{10}$  particles/ml. Store at 4 °C.

*Note: Figure 1 indicates: i) the concentration of beads, calculated using a Burkert chamber (MARI0640230, VWR) under the inverted microscope with 10x, 20x, and 40x objectives; ii) the size-distribution of beads, with a mean diameter of  $30 \pm 6.2 \mu\text{m}$ ; calculated as the mean diameter of beads with respect to the reference scale bars in 20x and 40x acquisitions; iii) the elemental composition of beads as determined by X-ray photoelectron spectroscopy (XPS) in terms of oxygen, carbon, nitrogen, and nickel; iv) the differential zeta potential (ZP) of +6 mV in comparison to the stripped beads, as determined by electrophoretic light scattering (ELS).*



**Figure 1. Quality controls and parallel recovery of EVs and EV-RNA from different biological fluids.** A. Representative images of beads prepared in Step A7, diluted 1:500 in PBS, taken with a microscope Leica DM IL LED with a LAS X software. Scale bar is 50  $\mu\text{m}$ . B. Parameters of beads prepared in Step A7. *Distribution* was assessed by ImageJ software, *elemental composition* by X-ray photoelectron spectroscopy (XPS), *differential ZP* by electrophoretic light scattering (ELS) in stripped (Step A3) and functionalized (Step A7) beads diluted 1:50 in PBS 1x (standard error <8%). C. Distribution of EVs recovered from media incubated with primary mouse astrocytes, 2 ml of plasma, 30 ml of urine, and 3 ml of saliva, all

collected from the same healthy volunteer and processed in parallel. Luria-Bertani media incubated with pure culture of *E. coli* cells. A representative EV distribution was assessed by TRPS using NP250 nanopores (A58835, A59044). D. Bioanalyzer profiles of the relative RNA profile loading 1.5 µl out of 10 in an Agilent Pico RNA Chip.

#### Annex 1:

**XPS analysis:** A drop of beads, either stripped (Step A3) or functionalized with nickel sulphate (Step A7) was dried at room temperature on a silicon substrate and measured at 0° take-off angle. Whilst the samples were similar in terms of oxygen (35.3% vs 37.4%), carbon (64.3% vs 59.8%), and nitrogen (present in EDTA; 0.4% vs 0.5%), the nickel content instead varied from stripped beads (undetected) and functionalized beads (2.3%). The standard error did not exceed the 1-2% of the value reported in Figure 1B.

**Sample details of Figures 1C and 1D.** Primary astrocytes were derived from cortex of neonatal mice (P0-1) and were grown under standard conditions (10% FBS in DMEM) in 6-well plates for two weeks (Basso *et al.*, 2013). Forty-eight hours before the collection, 36 ml of serum-free media were incubated with approximately  $4 \times 10^6$  cells seeded (2 ml per well). In case of OMVs, we processed 50 ml of LB that fed overnight a BL21 strain already characterized for the secretion of OMVs (kindly provided by Prof. Grandi's lab) (Fantappiè *et al.*, 2017).

#### B. NBI: EV capture

1. Add at least 1:40 volume of beads prepared in Step A7, drop by drop, on the surface of the biological fluid containing EVs/exosome-like liposomes/OMVs and incubate for 30 min at RT in gentle shaking (using the VWR Tube Rotator) to avoid sedimentation of beads.

*Note: The biological fluids include: cell culture medium, plasma, cerebrospinal fluid (CSF), urine, saliva, as well as LB medium to recover OMVs from E. coli strains. We typically use 25 µl of beads per ml of:*

- a. Cell culture medium (the presence of more than 2% FBS in the medium will cause bead aggregation during incubation, affecting the yield of EVs; in the presence of 5% FBS for example, we usually dilute it 1:5 in PBS to efficiently recover EVs). Importantly, refer to FBS characterization (Wei *et al.*, 2016) as it normally contains small vesicles of non-human origin;
- b. Human plasma diluted 1:3 in PBS 1x (Figure 1C);
- c. Human urine, un-diluted (Figure 1C);
- d. Human saliva diluted 1:3 in PBS 1x (Figure 1C).

*All the biological fluids are subjected to centrifugation at 2,800 x g for 10 min at RT to remove potential cellular contaminants (i.e., platelets in plasma or leukocytes in urine) before starting incubation with beads.*

2. Centrifuge the mixture at 600-800 x g for 2 min at RT.



*Note: To avoid disturbing the bead pellet, it is advisable to use 15 ml Falcon tubes during incubation of cell supernatant/biological fluid with beads. Moreover, it is advisable to use swing-bucket rotors during centrifugation steps, to better locate the bead-pellet on the bottom of the tube and allow gentle sedimentation of beads.*

- Carefully remove (transfer or discard) the supernatant.

*Note: We usually call this supernatant as “EV-depleted medium” and use it in internal control experiments for EV enumeration or downstream RNA analyses, especially in case we compare the potential enrichment of specific EV-transcripts.*

### C. NBI: EV elution

- Prepare the elution buffer 1x by diluting Solution A concentrated 5x (Recipe 3) and Solution B concentrated 5x (Recipe 4) in 0.2  $\mu$ m filtered PBS 1x. Gently mix the solution obtained.

*Note: This buffer must be freshly prepared at RT prior usage and should be used within 15-20 min to obtain an efficient release of polydisperse EVs from the beads.*

- Add 0.5 to 2 bead-volumes of elution buffer 1x to the bead-pellet and transfer them in a low-binding tube. Incubate the tube in a Thermomixer for 15 min, in orbital shaking at 600 rpm and 28 °C, to keep beads in suspension and favor the dissociation of bound vesicles. During the incubation, flick the tube every 2-3 min to avoid sedimentation of beads.

*Note: Low-binding tubes are designed for proteins, but are useful for minimizing interaction with vesicles. The volume of elution buffer 1x determines the concentration of EVs obtained.*

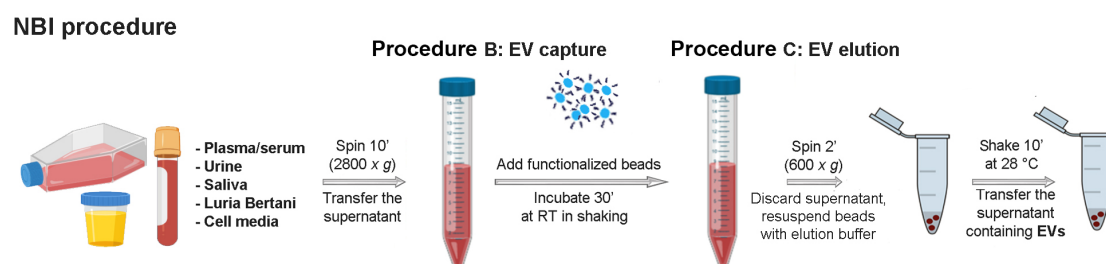
- Pellet beads at 600-800 x g for 30 s and carefully transfer the EV-containing supernatant in a new labeled 1.5 ml centrifuge tube.

*Note: Optionally, we perform an additional centrifugation step to pellet any residual beads.*

- Optional: Repeat Step C2 to recover EVs in case you have used less than two bead-volumes of elution buffer 1x.

*Note: In addition to Step C2, it ensures an efficient elution of bead-captured EVs. Figure 1C shows a distribution of EVs recovered from media of primary mouse astrocytes, from 2 ml of plasma, 30 ml of urine, and 3 ml of saliva all collected from the same healthy volunteer and processed in parallel, as well as from E. coli cells.*

A schematic representation of the NBI procedure is indicated in Figure 2.



**Figure 2. Schematic representation of the NBI method described in Procedures B and C.**

The NBI procedure it is entirely performed at room temperature, except the Step C2.

Functionalized agarose beads prepared in Procedure A are stable for at least four months at 4 °C. The procedures described in Procedures B and C are routinely performed to isolate EVs and last approximately 45 min, depending on the number of samples to be processed.

#### D. RNA extraction from EVs

To extract nucleic acids preserving the majority of RNA classes enclosed in EVs, we typically use the TRIzol™ Reagent protocol (Rio *et al.*, 2010) with minor modification in the procedure and following the general guidelines to avoid/minimize RNase contamination.

1. Add 0.5 ml of TRIzol™ Reagent per 1 ml of sample (a suspension of EVs in the elution buffer 1x) and briefly vortex to homogenize. Incubate at room temperature for 2-3 min.
2. Add 0.2 ml of chloroform per 1 ml of TRIzol™ Reagent used, then homogenize by inverting the tube several times and incubate for 2-3 min.
3. Centrifuge the sample for 10 min, at 13,000 x g at 4 °C, to obtain a lower red phenol-chloroform, an interphase, and a colorless upper aqueous phase.

*Note: In case of EVs the interphase appears as a slightly visible ring.*

4. Carefully transfer the aqueous phase to a new tube, avoiding any contamination from the interphase or organic layer into the pipette.
5. Add 0.6 ml of isopropanol to the aqueous phase, per 1 ml of TRIzol™ Reagent used in Step D1. Briefly vortex and incubate 5 min on ice.
6. Centrifuge for 15 min, at 13,000 x g and 4 °C, to precipitate nucleic acids.

*Note: A nucleic acid pellet derived from EVs is hardly visible. Optionally, with glycogen you can facilitate visualization of gel-like pellet at the bottom of the tube.*

7. Wash the pellet with 70% Ethanol and air dry for 5-10 min.

*Note: We optionally skip the step of 70% Ethanol to avoid the potential waste of the precipitated material in case of a recovery of EVs < 10<sup>7</sup> particles/ml.*

8. Resuspend the pellet in 7-15 µl of RNase-free water, by pipetting several times, and keep it on ice until stored at -80 °C.

*Note: Transfer 1.5 µl of the solution in a new 0.2 ml tube for incubation at 70 °C and ice-killing to profile the nucleic acid fragment distribution by Bioanalyzer Pico Chip. Figure 1D shows the size distribution of nucleic acid fragments recovered from the different biological fluids indicated in Figure 1C.*

#### Data analysis

Image analysis of Figure 1A was performed using ImageJ software. TRPS data displayed in Figure 1C were analyzed by Izon Control Suite software v3 and CSV data were plotted using GraphPad Prism Software v5.1. Nucleic acid profile of samples in Figure 1D was examined with Agilent 2100 Expert Software.



## **Recipes**

### 1. Buffer S (Stripping buffer)

PBS 1x

Under the biological hood, filter with a 0.22 µm disposal unit

Store at room temperature

PBS 1x + 0.5 M NaCl + 50 mM EDTA pH 8.0

Filter with 0.2 µm disposals

Store at room temperature

### 2. Buffer N

0.4 M of NiSO<sub>4</sub> in pure water

Filter with 0.2 µm disposals

Store at 4 °C.

### 3. Solution A 5x

PBS 1x + 16 mM EDTA

Filter with 0.2 µm disposals

Store at 4 °C

*Note: This storage ensures stability for months. A pH of 7.8 is a quality control.*

### 4. Solution B 5x

PBS 1x + 10 mM NaCl + 225 µM citric acid

Filter with 0.2 µm disposals

*Note: This storage ensures stability for months. A pH of 7.2 is a quality control.*

## **Acknowledgments**

We acknowledge the previous work (Notarangelo *et al.*, 2019) describing the novelty and the application of NBI procedure. We are grateful to Prof. G. Grandi's lab (University of Trento, CIBIO) for providing us the *E. coli* strain that we used in this work to isolate OMVs and OMV-nucleic acids.

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## **Competing interests**

MN, AP, AQ, and VGD declare a potential conflict of interest for a patent application regarding the NBI method (P019950WO).

## **Ethics**

Human specimens involved in this study derived from healthy subjects who gave informed consent before blood and urine sampling in the frame of a study approved by the Local Ethics Committee of the University of Trento. Mice were used to obtain astrocytes in the frame of a study approved by the Italian Ministry of Health (project number 603/2017-PR).

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